

## Review Article

# Sesquiterpenoids from the sunflower family as potential anti-inflammatory candidates: a review

Cheng Chen<sup>a,1</sup>, Zheling Feng<sup>a,b,1</sup>, Jovana Petrović<sup>c</sup>, Marina Soković<sup>c</sup>, Yang Ye<sup>b</sup> and Ligen Lin<sup>a,\*</sup>

<sup>a</sup>State Key Laboratory of Quality Research in Chinese Medicine, Institute of Chinese Medical Sciences, University of Macau, Taipa, Macau, China

<sup>b</sup>State Key Laboratory of Drug Research and Natural Products Chemistry Department, Shanghai Institute of Materia Medica, Chinese Academy of Sciences, Shanghai 201203, China

<sup>c</sup>China-Serbia "Belt and Road" Joint Laboratory for Natural Products and Drug Discovery, Institute for biological research "Siniša Stanković", University of Belgrade, Bulevar Despota Stefana 142, 11000 Belgrade, Serbia

<sup>1</sup>Cheng Chen and Zheling Feng contributed equally to this work.

\*Correspondence: [ligen@um.edu.mo](mailto:ligen@um.edu.mo) (L. Lin)

Received: 12 July 2023; Revised: 29 August 2023; Accepted: 13 September 2023

Published online: 29 September 2023

DOI 10.15212/AMM-2023-0026

## ABSTRACT

Inflammation is an essential part of the immune response to injury and infection. Emerging evidence indicates that long-term low-grade inflammation is positively correlated with many diseases, such as cancer, metabolic disorders, and cardiovascular diseases. Due to common anti-inflammatory drugs are suitable for treating acute inflammation and cause severe adverse effects, new safe and effective drug candidates are urgently needed for treating chronic inflammation. Plants of the Asteraceae family have been widely used in traditional medicines for relieving fever symptoms and killing pathogens. The anti-inflammatory properties of sesquiterpenoids from plants in the Asteraceae family have attracted increasing attention in recent decades because of their structural complexity and potent bioactivities. Herein, we provide a comprehensive and up-to-date summary of sesquiterpenoids from the Asteraceae family with anti-inflammatory properties, including their drug likeness and druggability, as analyzed with the SwissADME and ADMETlab online tools. In the future, some sesquiterpenoids might serve as therapeutic agents to treat inflammation-associated diseases.

**Keywords:** Asteraceae, sesquiterpenoids, anti-inflammation, drug-likeness, SwissADME, ADMETlab

## 1. INTRODUCTION

Inflammation is a key response of the immune system [1, 2]. Long-term low-grade inflammation is considered a culprit of cancer, metabolic disorders, and cardiovascular diseases [3-5]. During chronic inflammation, the profiles of pro-inflammatory and anti-inflammatory mediators greatly vary [6, 7]. Macrophages are present in most organs and tissues, and play important roles in tissue development, inflammation, and defense against pathogens [8-10]. M0 type macrophages can differentiate into either pro-inflammatory M1 type or anti-inflammatory M2 type macrophages [8-11]. The nuclear factor- $\kappa$ B (NF- $\kappa$ B) pathway and mitogen-activated protein kinase (MAPK) pathway are the main regulators of inflammatory responses. In the unstimulated state, the p50/p65 heterodimer is retained by the inhibitory protein I $\kappa$ B in the cytoplasm. Under external stimuli, I $\kappa$ B

kinase (IKK) is activated and subsequently phosphorylates serine in the I $\kappa$ B subunit, resulting in I $\kappa$ B release and ubiquitination. The liberated NF- $\kappa$ B dimer then translocates to the nucleus and initiates downstream transcription [12]. Increased MAPK activity, particularly p38, c-Jun N-terminal kinases (JNKs), and extracellular signal-regulated kinases (ERKs), and their involvement in the regulation of the synthesis of inflammation mediators, make them potential targets for anti-inflammatory therapeutics [13].

Current clinically applied anti-inflammatory drugs include glucocorticoids and non-steroidal anti-inflammatory drugs, all of which can cause serious adverse reactions [14-16]. Thus, safe and effective anti-inflammatory drugs must urgently be developed. Natural products have become a promising alternative to identify drug leads, owing to their efficiency and safety. Growing evidence indicates the great potential

of natural medicines in treating inflammation-associated diseases [17, 18]. Drug-likeness is determined from structural or physicochemical inspection of a molecule, and used to qualitatively evaluate its bioavailability, and enable its use as an oral drug. Drug-likeness can be applied in the prediction of drug absorption, distribution, metabolism, and excretion (ADME) properties [19]. Drug-likeness and ADME properties of hits can be predicted in early stages of drug discovery, and compounds with ideal properties can be selected for experimental validation, thus decreasing costs and increasing the success rate of lead compound discovery.

Asteraceae is a very large family of flowering plants comprising more than 1,620 genera and 23,600 accepted species. Plants in the Asteraceae family are widely distributed worldwide except in Antarctica, and are particularly abundant in North America, the Andes in South America, eastern Brazil, the Mediterranean, Central Asia, and southwestern China [20, 21]. Many Asteraceae plants possess substantial therapeutic value in traditional medicines, such as *Artemisia annua* and dandelion [22, 23]. In China, many Asteraceae plants are used for treating inflammatory responses occurring after pathogens invade the human body. For example, chrysanthemum is widely used to treat throat inflammation, fever, headache, and oral ulcers [24], and dandelion is used to treat upper respiratory tract infections, lung infections, and purulent diseases [25]. Modern pharmacological studies have indicated that Asteraceae plants possess anti-cancer, cardiovascular-protective, hepatoprotective, anti-diabetic, anti-bacterial, and anti-viral properties [26-31]. Chemical investigations have been widely performed on Asteraceae plants to identify terpenoids, flavonoids, lignans, polyacetylenes, and polyphenols. Among these compounds, sesquiterpenoids are notable for their great structural complexity and diverse bioactivities [32]. Sesquiterpenoids consist of three isoprene units, which can be linear, monocyclic, bicyclic, or tricyclic. Most sesquiterpenoids exist in the form of volatile oils, as oxygen-containing derivatives, such as alcohols, ketones, lactones, etc. Several sesquiterpenes have been successfully developed as medicines, such as artemisinin for the treatment of malignant malaria [33].

Interestingly, many sesquiterpenoids from plants in the Asteraceae family have been reported with anti-inflammatory properties in *in vitro* or *in vivo* models [34, 35]. Herein, we provide a comprehensive and up-to-date summary of these sesquiterpenoids. We further analyzed their drug likeness by using the SwissADME and ADMETlab online tools, to aid in their development as candidates to treat inflammation-associated diseases.

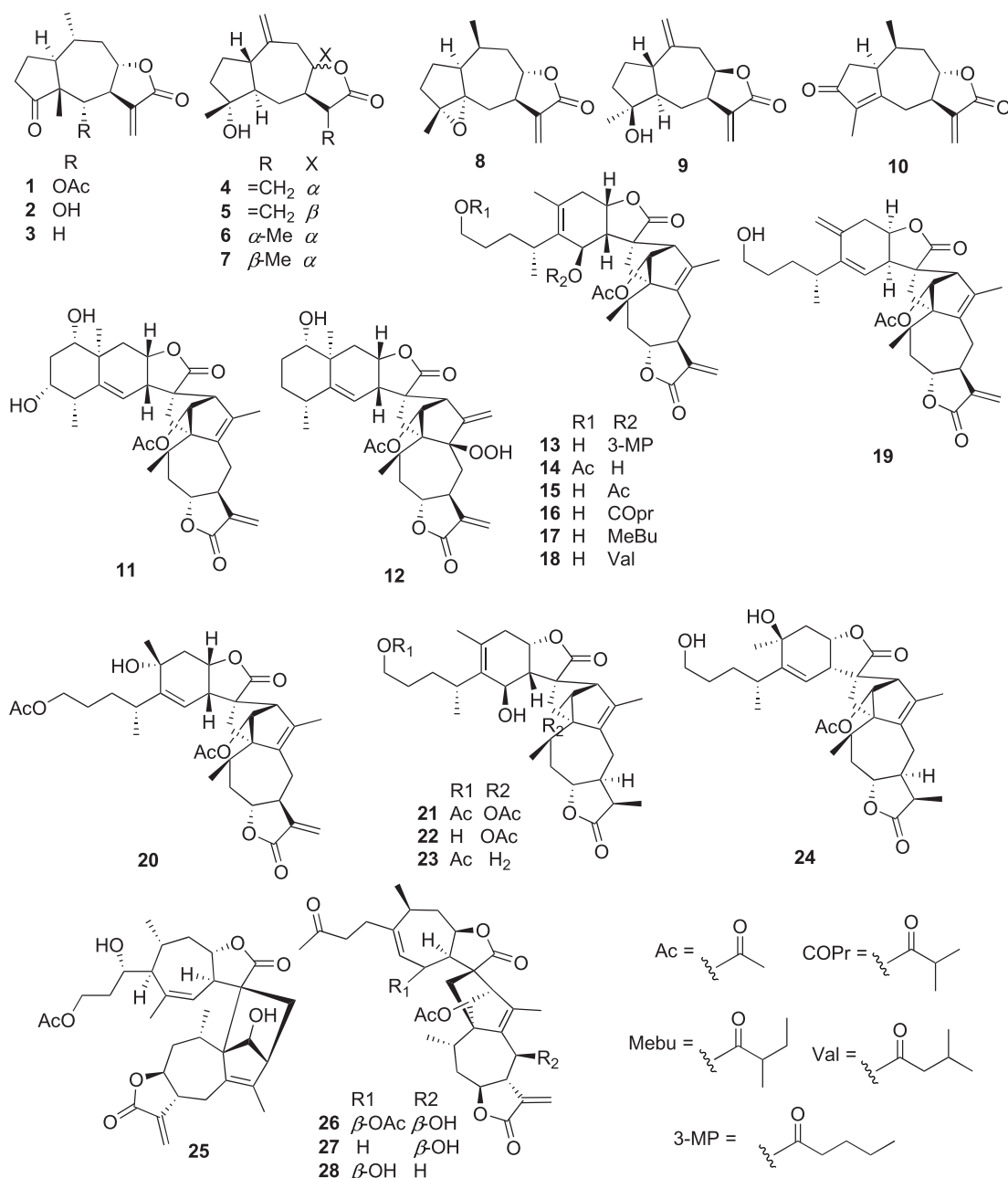
## 2. SESQUITERPENOIDS FROM ASTERACEAE PLANTS WITH ANTI-INFLAMMATORY PROPERTIES

We searched the Google Scholar, Web of Science, Scopus, and PubMed databases with the keywords sesquiterpenoids, Asteraceae, and inflammation. A total

of 88 sesquiterpenoids from Asteraceae plants with anti-inflammatory properties were identified (Figures 1–4). According to their carbon skeletons, these compounds can be classified into guaiane-type sesquiterpenoids (1–65, Figures 1–3), oplopane-type sesquiterpenoids (66–75), bisabolane-type sesquiterpenoids (76–80), eudesmanolide-type sesquiterpenoids (81–83), cadinane-type sesquiterpenoids (84–87), and a caryophyllane-type sesquiterpenoid (88) (Figure 4). The anti-inflammatory activities of these sesquiterpenoids are summarized in Table 1.

Lipopolysaccharides (LPS) are composed of lipids and polysaccharides, consisting of O-antigens, an outer nucleus and an inner nucleus linked by covalent bonds; they are present in the outer membranes of Gram-negative bacteria [69, 70]. LPS bind Toll-like receptor 4 (TLR4) or CD14, and consequently induce inflammatory responses in macrophages. LPS have been widely applied to identify anti-inflammatory agents and investigate their underlying mechanisms. Compounds 1–28, 32, 33, 52, 54–56, and 61–80 exhibit anti-inflammatory activities in LPS-stimulated RAW264.7 macrophages [36–40, 42, 52, 53, 56, 58, 59, 61, 62, 71, 72]. Among them, compounds 1–28, 32, 33, and 63–80 exhibit inhibitory effects against nitric oxide (NO) production, and compounds 66 and 70 down regulate heme oxygenase expression [57]. Compound 56 was found to suppress the mRNA expression of tumor necrosis factor- $\alpha$  (TNF- $\alpha$ ), interleukin-1 $\beta$  (IL-1 $\beta$ ), IL-6, inducible NO synthase (iNOS), and cyclooxygenase-2 (COX-2) [53]. Compounds 52, 54, 55, 61, and 62 decrease the production of inflammatory factors through inhibiting the NF- $\kappa$ B pathway. Notably, compound 52 blocks NF- $\kappa$ B signaling through directly interacting with cysteine 46 of IKK $\alpha$ / $\beta$ , according to activity-based protein profiling [35]. In LPS- and ATP-induced THP-1 macrophages, compounds 29–31 and 53 suppress inflammatory responses; among them, compounds 29 and 53 inhibit the activation of NOD-like receptor thermal protein domain associated protein 3 (NLRP3) inflammasomes, thus decreasing the secretion of IL-1 $\beta$  [34, 41]. Interestingly, compound 53, a structural analogue of compound 52 isolated from the same plant (*Ainsliaea macrocephala*), activates the deacetylase Sirtuin1, and consequently decreases the acetylation of NLRP3 and inhibits the activation of NLRP3 inflammasomes [34]. Furthermore, compound 53 decreases adipose tissue inflammation in C57BL/6J mice stimulated with LPS plus ATP [34]. LPS-induced BV-2 cells are often used as a model of neuroinflammation. In this model, compounds 34–47 were found to effectively inhibit NO production, suggesting their potential in the treatment of neuroinflammation [44, 45]. In addition, compounds 57–60 were found to inhibit NO production in LPS-induced 293T cells [55]. Ameliorating inflammation in macrophages is critical for a variety of human inflammatory disorders, such as sepsis-associated multiple organ dysfunction, acute brain/lung/hepatic/renal injuries, microbial infection, tumorigenesis, neurodegenerative

## Review Article



**Figure 1 | Structures of guaian-type sesquiterpenoids from plants of the genus *Inula* with anti-inflammatory activity.**

disorders, cardiovascular and metabolic diseases, and autoimmune diseases. Thus, these compounds possess great potential to be further developed to treat a variety of inflammatory diseases.

TNF- $\alpha$  is a key pro-inflammatory cytokine [73]. Compound **48** was found to decrease TNF- $\alpha$ -induced inflammation in Jurkat cells through inhibiting the NF- $\kappa$ B pathway [46, 74]. In particular, compound **48** directly alkylates the p65 subunit of NF- $\kappa$ B at Cys38 and Cys120 [46], and might be used to treat acute T cell

leukemia. COX-2 is an enzyme responsible for inflammation and pain [75]. In human colon HT29 cells, compounds **49** and **51** were found to suppress COX-2 levels, thereby alleviating inflammation [47-49]. Compounds **81-83**, were found to reduce inflammatory pain in a rat model of carrageenan-induced foot swelling [63]. Compound 12-O-tetradecanoylphorbol 13-acetate (TPA) induces skin inflammation through promoting the production of TNF- $\alpha$  and the formation of leukotriene B<sub>4</sub> (LTB<sub>4</sub>) [66, 76]. Prolonged TPA exposure results in ear

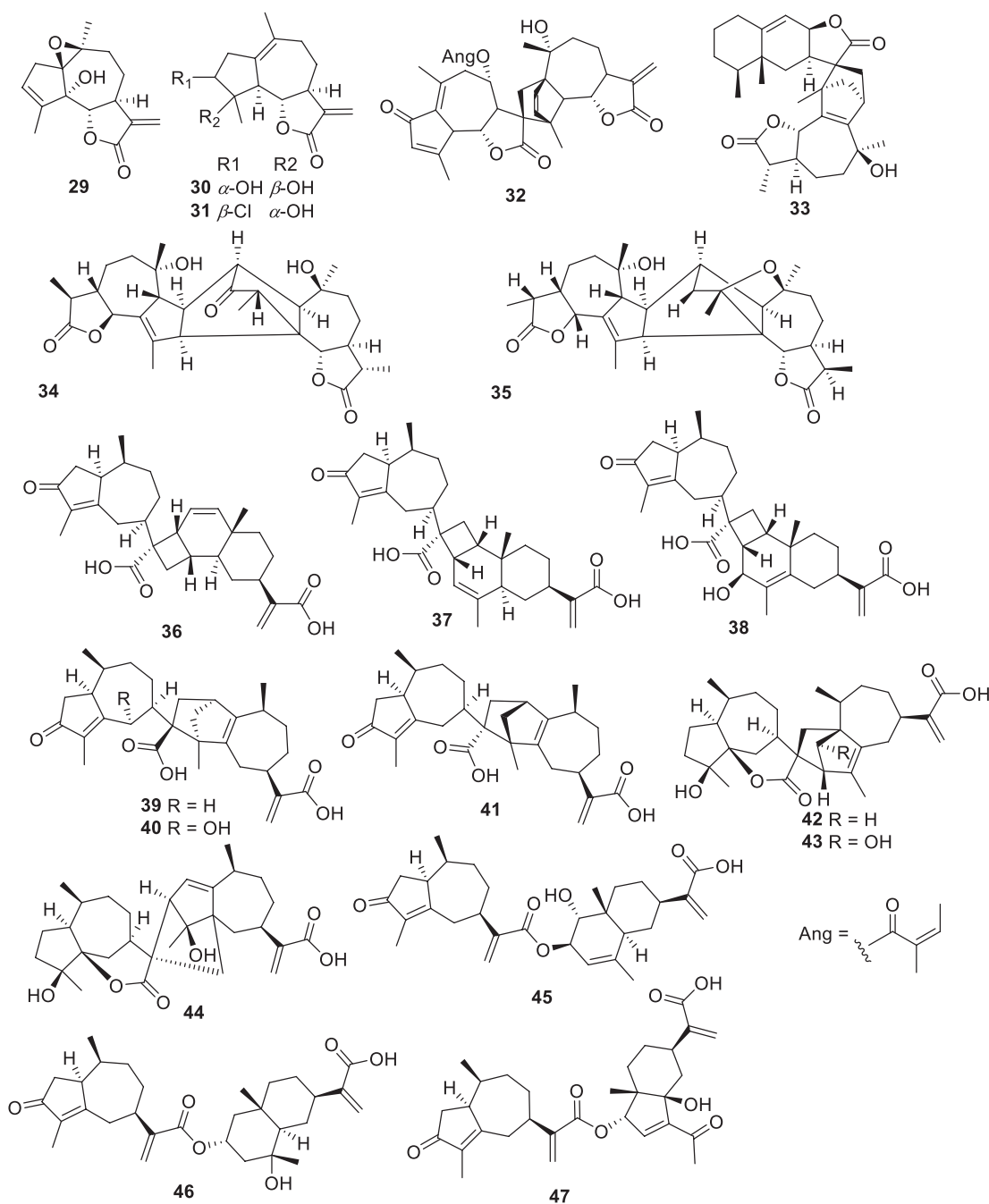


Figure 2 | Structures of guaiane-type sesquiterpenoids from plants of the genus *Artemisia* with anti-inflammatory activity.

swelling, inflammatory cell infiltration, and epidermal hyperplasia in mice [77]. In TPA-induced ear swelling mice, compounds **84–86** and **87** effectively alleviate ear inflammation [64–66, 76, 78]. These compounds could be developed to alleviate swelling and pain.

Acute lung injury (ALI) is a respiratory failure disease whose main pathological changes include excessive recruitment of pro-inflammatory mediators and activation of neutrophils. Compound **50** was found to decrease

inflammatory cell infiltration and pro-inflammatory cytokine levels in an LPS-induced mouse model of ALI [50, 51]. Furthermore, compound **50** treatment suppresses phosphorylation of key regulators of the NF- $\kappa$ B signaling pathway in lung epithelial BEAS-2B cells and alveolar macrophage MH-S cells. In a murine model of colitis induced by dextran sulfate sodium, compound **88** attenuates inflammatory responses through suppressing NF- $\kappa$ B activity [67], and activating the endogenous

## Review Article

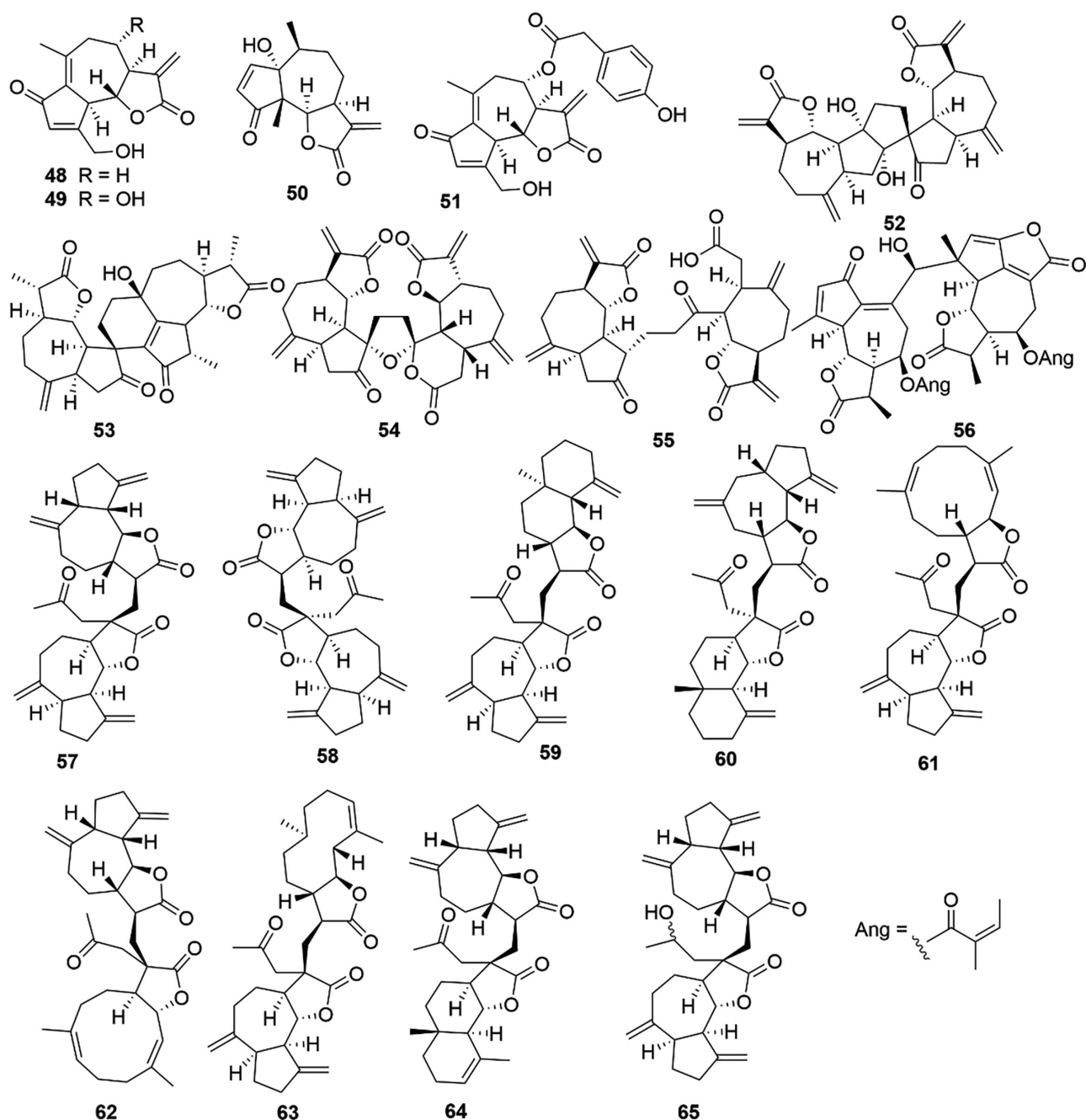


Figure 3 | Structures of guaianane-type sesquiterpenoids from other Asteraceae plants with anti-inflammatory activity.

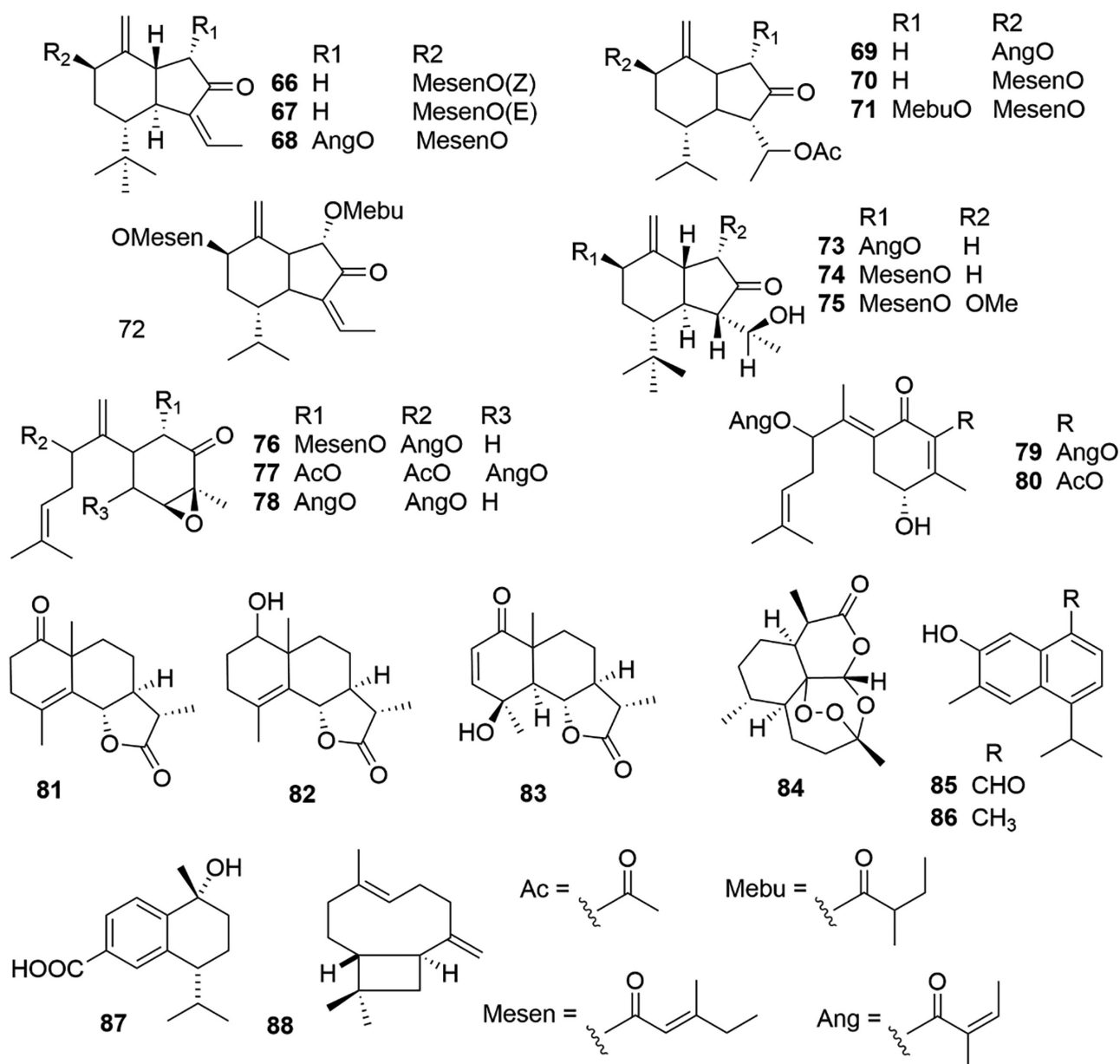
cannabinoid 2 receptor and peroxisome proliferator-activated receptor- $\gamma$  (PPAR $\gamma$ ) pathway [68].

### 3. COMPARISON OF THE DRUG-LIKENESS OF ANTI-INFLAMMATORY SESQUITERPENOID FROM ASTERACEAE PLANTS WITH RESPECT TO MARKETED DRUGS

We used SwissADME, an online tool developed by the Swiss Institute of Bioinformatics [79], to predict the

molecular descriptors of the anti-inflammatory sesquiterpenoids from Asteraceae plants. The simplified molecular input line entry system (SMILES) files of the compounds were uploaded to the website, and descriptors were predicted, including molecular weight (MW), number of chiral centers, numbers of hydrogen bond acceptors (HBA) and donors (HBD), number of rotatable bonds (RBs), number of rings, fraction of  $sp^3$  carbons ( $F_{sp^3}$ , the ratio of  $sp^3$  hybridized carbons to the total number of carbons), and the fraction of aromatic





**Figure 4 | Structures of oplopane-type sesquiterpenoids (66–75), bisabolane-type sesquiterpenoids (76–80), eudesmanolide-type sesquiterpenoids (81–83), cadinane-type sesquiterpenoids (84–87), and a caryophyllane-type sesquiterpenoid (88) from Asteraceae plants with anti-inflammatory activity.**

heavy carbon atoms (Far, the number of heavy aromatic atoms divided by the total number of heavy atoms) [80]. In addition, the database supports simultaneous prediction and comparison of pharmacokinetic properties of multiple compounds. Seven molecular descriptors for each compound, including the mean and median values, were shown in [Supplementary Information Table S1](#). ADME, drug-likeness, and medicinal chemistry friendliness are important factors in drug development [81]. The physicochemical properties, pharmacokinetics, polar surface area (PSA), logS, logP, and bioavailability

properties of these sesquiterpenoids were documented in [Supplementary Information Table S2](#). In addition, multiple algorithmic procedures were used to determine the values of logP and logS [82, 83].

We classified marketed drugs into synthetic compounds, assumed synthetic compounds, natural-product-type macrocycles, polycyclic compounds, natural products, and natural-product derivatives. Their molecular descriptors, including MW, HBA, HBD, logP, PSA, RBs, and number of chiral centers, were obtained from previous publications ([Supplementary Information](#)

## Review Article

**Table 1** | Sesquiterpenoids from Asteraceae plants with anti-inflammatory activity.

Skeleton	Source	No name	Model	Dose	Outcomes	References
Guaiane-type	<i>Inula falconeri</i>	1 Ergolide	LPS-induced RAW264.7 macrophages	0.07 $\mu\text{M}^*$	Inhibition of NO production	[36]
		2 Carpesiolin		2.18 $\mu\text{M}^*$		
		3 2,3-Dihydroaromaticin		0.40 $\mu\text{M}^*$		
		4 Inuviscolide		9.64 $\mu\text{M}^*$		
		5 8-Epinuvisolide		3.94 $\mu\text{M}^*$		
		6 11 $\beta$ ,13-Dihydroinuviscolide		41.20 $\mu\text{M}^*$		
		7 11 $\alpha$ ,13-Dihydroinuviscolide		19.53 $\mu\text{M}^*$		
		8 4 $\alpha$ ,5 $\alpha$ -Epoxy-10 $\alpha$ ,14H-1- <i>epi</i> -inuviscolide		0.11 $\mu\text{M}^*$		
		9 4,8-Bis- <i>epi</i> -inuviscolide		7.30 $\mu\text{M}^*$		
		10 Xerantholide		12.86 $\mu\text{M}^*$		
<i>Inula japonica</i>		11 Japonicone R	LPS-induced RAW264.7 macrophages	8.90 $\mu\text{M}^*$	Inhibition of NO production	[37]
		12 Japonicone T		4.30 $\mu\text{M}^*$		
		13 Japonicone Q		8.50 $\mu\text{M}^*$		
		14 Inulanolide A		4.20 $\mu\text{M}^*$		
		15 Japonicone M		12.0 $\mu\text{g/ml}$		
		16 Japonicone N		-		
		17 Japonicone O		-		
		18 Japonicone P		-		
		19 Inulanolide C		4.90 $\mu\text{M}^*$		
		20 Japonicone S		4.30 $\mu\text{M}^*$		
<i>Inula linearifolia</i>		21 Dibritannilactone B	LPS-induced RAW264.7 macrophages	43.77 $\mu\text{M}^*$	Inhibition of NO production	[40]
		22 Dibritannilactone C		49.44 $\mu\text{M}^*$		
		23 Dibritannilactone D		25.08 $\mu\text{M}^*$		
		24 Dibritannilactone E		29.18 $\mu\text{M}^*$		
		25 Linearifolianoid I		1.02 $\mu\text{M}^*$		
		26 Linearifolianoid J		1.79 $\mu\text{M}^*$		
		27 Linearifolianoid K		10.02 $\mu\text{M}^*$		
		28 Linearifolianoid L		10.16 $\mu\text{M}^*$		

Table 1 | Continued

Skeleton	Source	No name	Model	Dose	Outcomes	References
	<i>Artemisia codonocephala</i>	29 Lavandolide D	LPS- plus ATP-induced THP-1 cells	3.31 $\mu\text{M}^*$	Inactivation of NLRP3 inflammasomes	[41]
		30 3 $\alpha$ ,4 $\beta$ -Dihydroxy-5,7 $\alpha$ ,6 $\beta$ (H)-guaia-1(10),11(13)-dien-12,6-olide		3.68 $\mu\text{M}^*$	Inhibition of NO production	
		31 3 $\beta$ -Chloro-4 $\alpha$ -hydroxy-5,7 $\alpha$ ,6 $\beta$ (H)-guaia-1(10),11(13)-dien-12,6-olide		0.51 $\mu\text{M}^*$		
	<i>Artemisia argyi</i>	32 Dsf-27	LPS-induced RAW264.7 macrophages	10 $\mu\text{M}$	Inhibition of NO production	[42]
	<i>Artemisia freyniana</i>	33 Artefrenisin A		38.30 $\mu\text{M}^*$		[43]
	<i>Artemisia absinthium</i>	34 Absinthin C	LPS-induced BV-2 cells	1.52 $\mu\text{M}^*$	Inhibition of NO production	[44]
		35 Isoanabsinthin		1.98 $\mu\text{M}^*$		
	<i>Artemisia rupestris</i>	36 Artepestrin A	LPS-induced BV-2 cells	27.30 $\mu\text{M}^*$	Inhibition of NO production	[45]
		37 Artepestrin B		39.80 $\mu\text{M}^*$		
		38 Artepestrin C		29.80 $\mu\text{M}^*$		
		39 Artepestrin D		32.70 $\mu\text{M}^*$		
		40 6 $\alpha$ -Hydroxyartepestrin D		23.00 $\mu\text{M}^*$		
		41 Artepestrin E		38.60 $\mu\text{M}^*$		
		42 Artepestrin F		17.00 $\mu\text{M}^*$		
		43 2' $\alpha$ -Hydroxyartepestrin F		71.80 $\mu\text{M}^*$		
		44 Artepestrin G		43.60 $\mu\text{M}^*$		
		45 Rupestrinate A		33.00 $\mu\text{M}^*$		
		46 Rupestrinate B		40.60 $\mu\text{M}^*$		
		47 Rupestrinate C		30.10 $\mu\text{M}^*$		
	<i>Cichorium intybus</i>	48 8-Deoxylactucin	TNF- $\alpha$ -induced Jurkat T-cells	50 $\mu\text{M}^{\#}$	Alkylation of Cys38 and Cys120 of the p65 subunit of NF- $\kappa\text{B}$ , and inhibition of NF- $\kappa\text{B}$ activation	[46, 47]
		49 Lactucin	Human colon HT29 cells	16 $\mu\text{g/ml}$	Inhibition of COX-2	[48, 49]
	<i>Tanacetum vulgare</i>	50 (3a <i>s</i> ,6 <i>S</i> ,6 <i>a</i> <i>s</i> ,9 <i>a</i> <i>s</i> ,9 <i>br</i> )-6 <i>a</i> -Hydroxy-6,9 <i>a</i> -dimethyl-3-methylene-4,5,6,9 <i>b</i> -tetrahydro-3 <i>ah</i> -azulenol[8,7- <i>b</i> ]furan-2,9-dione	Lung epithelial BEAS-2B cells and alveolar macrophage MH-S cells	20 $\mu\text{M}$	Inhibition of NF- $\kappa\text{B}$ pathway, decreased airway permeability and production of pro-inflammatory cytokines	[50, 51]
			LPS-induced ALI mice	10 mg/Kg		



## Review Article

Table 1 | Continued

Skeleton	Source	No name	Model	Dose	Outcomes	References
<i>Cichorium intybus</i>	<i>Ainsliadimer macrocephala</i>	51 Lactucopicrin	Human colon HT29 cells	-	Inhibition of COX-2	[48, 49]
		52 Ainsliadimer A	LPS-induced RAW264.7 macrophages	8 $\mu\text{M}$	Targeting of Cys46 in IKK $\alpha$ / $\beta$ to block NF- $\kappa$ B signalling	[35, 52]
<i>Eupatorium perfoliatum</i>	<i>Ainsliadimer C</i>	53 Ainsliadimer C	LPS- plus ATP-induced THP-1 cells/LPS plus ATP induced C57BL/6J mice	80 $\mu\text{M}$	Inactivation of NLRP3 inflammasomes	[34, 52]
		54 Macrocephaliolide A	LPS-induced RAW264.7 macrophages	0.99 $\mu\text{M}^*$	Inhibition of NF- $\kappa$ B pathway	[52]
<i>Eupatorium perfoliatum</i>	<i>Macrocephaliolide B</i>	55 Macrocephaliolide B	LPS-induced RAW264.7 macrophages	6.13 $\mu\text{M}^*$		
		56 Diguaiaperfolin	LPS-induced RAW264.7 macrophages	16.5 $\mu\text{M}^*$	Suppression of mRNA expression of TNF- $\alpha$ , IL-1 $\beta$ , IL-6, iNOS, and COX-2	[53, 54]
<i>Vladimiria souliei</i>	<i>Viasouliolide A</i>	57 Viasouliolide A	LPS-induced 293T cells	1.14 $\mu\text{M}^*$		[55]
		58 Viasouliolide B		2.53 $\mu\text{M}^*$		
<i>Vladimiria souliei</i>	<i>Viasouliolide C</i>	59 Viasouliolide C		1.57 $\mu\text{M}^*$		
		60 Viasouliolide D		3.19 $\mu\text{M}^*$		
<i>Vladimiria souliei</i>	<i>Viasouliolide E</i>	61 Viasouliolide E	LPS-induced RAW264.7 macrophages	1.88 $\mu\text{M}^*$	Inhibition of NF- $\kappa$ B pathway	[56]
		62 Viasouliolide F		4.89 $\mu\text{M}^*$		
<i>Vladimiria souliei</i>	<i>Viasouliolide G</i>	63 Viasouliolide G		-	Inhibition of NO production	
		64 Viasouliolide H		7.24 $\mu\text{M}^*$		
<i>Vladimiria souliei</i>	<i>Viasouliolide I</i>	65 Viasouliolide I		2.46 $\mu\text{M}^*$		
		66 Tussilagone	LPS-induced RAW264.7 macrophages	11.86 $\mu\text{M}^*$	Inhibition of NO production by regulation of heme oxygenase	[57, 58]
<i>Tussilago farfara</i>	<i>7<math>\beta</math>-(4-Methylseneciolyloxy)-oplopa-3(14)E,8(10)-dien-2-one</i>	67 7 $\beta$ -(4-Methylseneciolyloxy)-oplopa-3(14)E,8(10)-dien-2-one	LPS-induced RAW264.7 macrophages	10.80 $\mu\text{M}^*$	Inhibition of NO production	[58]
		68 1 $\alpha$ -Angelyloxy-7 $\beta$ -(4-methylseneciolyloxy)-oplopa-3(14)Z,8(10)-dien-2-one		13.87 $\mu\text{M}^*$		
<i>Tussilago farfara</i>	<i>Tussilagone</i>	69 Tussilagone		71.57 $\mu\text{M}^*$		[58, 59]
		70 14-Acetoxy-7 $\beta$ -angelyloxy-notonipetranone	LPS-induced RAW264.7 macrophages	5.60 $\mu\text{M}^*$	Inhibition of NO production by regulation of heme oxygenase	[59, 60]

Table 1 | Continued

Skeleton	Source	No name	Model	Dose	Outcomes	References	
Bisabolene-type	<i>Tussilago farfara</i>	<b>71</b> 14-Acetoxy-7β-(3-ethyl-cis-crotonyloxy)-1α-(2-methylbutyryloxy)-notonipetranone	LPS-induced RAW264.7 macrophages	3.50 μM*	Inhibition of NO production	[59]	
		<b>72</b> 7β-(3-Ethyl-cis-crotonyloxy)-1α-(2-methylbutyryloxy)-3,14-dehydro-Z-notonipetranone		4.60 μM*			
		<b>73</b> 7β-Angeloyloxy-14-hydroxy-notonipetranone	LPS-induced RAW264.7 macrophages	60.29 μM*	Inhibition of NO production	[58]	
		<b>74</b> 7β-(3'-Ethyl-cis-crotonyloxy)-14-hydroxy-notonipetranone		32.87 μM*			
		<b>75</b> 7β-(3'-Ethyl-cis-crotonyloxy)-14-hydroxy-1α-(2'-methylbutyryloxy)-notonipetranone		15.64 μM*			
		<b>76</b> 1α-(3'-Ethyl-cis-crotonyloxy)-8-angeloyloxy-3β,4β-epoxy-bisabola-7(14),10-diene	LPS-induced RAW264.7 macrophages	20.86 μM*	Inhibition of NO production	[58]	
		<b>77</b> 1α,5α-Bisacetoxy-8-angeloyloxy-3β,4β-epoxy-bisabola-7(14),10-dien-2-one	LPS-induced RAW264.7 macrophages	5.51 μM	Inhibition of NO production	[61]	
		<b>78</b> 1β,8-Bisangeloyloxy-3α,4α-epoxybisabola-7(14),10-dien-2-one	LPS-induced RAW264.7 macrophages	-	Inhibition of NO production	[62]	
Eudesmanolide-type	<i>Artemisia barrelieri</i>	<b>79</b> Tussfararin F		13.60 μM*			
		<b>80</b> (4R,6E)-2-Acetoxy-8-angeloyloxy-4-hydroxybisabola-2,6,10-trien-1-one		17.70 μM*			
		<b>81</b> Barrelierin	Carrageenan-induced foot swelling in rats	90 mg/Kg	-	[63]	
		<b>82</b> Artemalin		90 mg/Kg			
Cadinane-type	<i>Artemisia annua</i>	<b>83</b> Barrelin		15 mg/Kg			
		<b>84</b> Artemisinin	TPA-induced ear swelling in mice	200 μM	Inhibition of NF-κB pathway	[64]	
		<b>85</b> 7-Hydroxy-14-cadalenal		> 228 μg/ear		[65]	
		<b>86</b> 7-Hydroxycadalene		>0.5 mg/ear		[66]	
Caryophyllane-type	<i>Heterotheca inuloides</i>	<b>87</b> 1α-Hydroxy-4αh-1,2,3,4-tetrahydrocadalen-15-oic acid		>0.5 mg/ear			
		<b>88</b> B-caryophyllene	Dextran sulfate sodium-induced colitis in mice	300 mg/Kg	Suppression of NF-κB activity, and activation of the endogenous CB2 receptor and PPARγ pathway	[67, 68]	

\*<sup>1</sup>C<sub>50</sub>, <sup>1</sup>C<sub>100</sub>, maximum dosage for those without asterisk or pound marker.

## Review Article

**Table S3)** [84]. We summarized the biophysicochemical properties of 88 sesquiterpenoids with anti-inflammatory properties from Asteraceae plants and compared with those of marketed drugs (Figure 5).

### 3.1 Size: molecular weight

Lipinski's rules have been widely used to evaluate drug candidates. These rules include no more than five HBD, no more than ten HBA, an MW <500 Daltons, and an octanol-water partition coefficient (logP)  $\leq 5$  [85]. The MWs of approved drugs showed an increasing trend from 300–360 Daltons (in 1959–1964) to 360–440 Daltons (in 2005–2009); in 2017, the MWs of 64% Food and Drug Administration (FDA)-approved drugs exceeded 440 Daltons [86]. The mean MW for anti-inflammatory sesquiterpenoids from Asteraceae plants is 436.00 Daltons, thus meeting Lipinski's rules (Figure 5a). Among these compounds, 60% of sesquiterpenoids had an MW <500 Daltons.

### 3.2 Chirality: number of chiral centers

A total of 56% of FDA-approved drugs contain one or more chiral centers, of which 88% exist in the form of a racemate [87]. Chirality is highly important for the interaction between a molecule and its protein target. The mean number of chiral centers of anti-inflammatory sesquiterpenoids from Asteraceae plants is 7.86, a number exceeding those of synthetic compounds, assumed synthetic compounds, polycyclic compounds, natural products, and natural-product derivatives, and less than those of natural-product-type macrocycles, owing to the structural complexity of sesquiterpenoids, particularly sesquiterpenoid dimers (Figure 5b). In general, total synthesis of a molecules containing multiple chiral centers is difficult [88, 89]. Thus, total synthesis of these sesquiterpenoids poses major challenges.

### 3.3 Polarity: PSA and HBD/HBA

PSA, defined as the molecular surface area of oxygen or nitrogen atoms plus hydrogen atoms connected to nitrogen or oxygen atoms, reflects the transportability of a molecule in the gastrointestinal (GI) tract. Topological PSA is often used in actual drug prediction [90]. Most molecules with more than five HBD and/or more than ten HBA are glycosylated compounds or amino acids, which cannot easily cross the blood-brain barrier (BBB) or any cellular membrane. Shultz's dataset analysis has indicated that 90% of drugs contain four or fewer HBD. The number of HBA increases significantly with increasing MW. The numbers of HBA and HBD affect the lattice energy and melting point, and thus are associated with solubility. In macromolecules, intramolecular hydrogen bonds are associated with permeability [91]. Owing to hydrogen bonding, PSA is highly correlated with the quantity of HBA but is less correlated with the quantity of HBD [89]. The mean PSA values are 84.36 Å<sup>2</sup> for anti-inflammatory sesquiterpenoids from Asteraceae plants, 86.9 Å<sup>2</sup> for new polycyclic drugs, and 105.3 Å<sup>2</sup> for

natural products. The mean numbers of HBA and HBD for these sesquiterpenoids are 5.64 and 1.01. Similarly, HBA, HBD, and PSA values increase with increasing MW (Figures 5c-e and 6). Most anti-inflammatory sesquiterpenoids from Asteraceae plants meet Lipinski's rules and thus might exhibit good oral bioavailability.

### 3.4 Molecular flexibility: RBs and aromatic character

RBs, bonds with free rotation, are single bonds that are not adjacent to triple bonds, do not connect hydrogen or halogen atoms, and are not included in rings containing fewer than five single bonds [92]. Aromatic compounds, particularly hetero-aromatic compounds, have made substantial contributions to drug discovery. However, the amide C-N bond is excluded from RBs because of its high rotational energy barrier. Aromaticity and RBs, but not MW, greatly affect solubility. Low molecular flexibility, as measured by the number of RBs, and low PSA or total HB, are important predictors of good oral bioavailability [93]. A higher RB number decreases the permeation rate, and the permeation rate is a prerequisite for oral bioavailability. The data of 164 FDA-approved small molecules in the past 5 years indicated a high proportion of N-heterocycles (88%) and aromatic skeletons (87%), followed by chiral molecules (63%) [94].

The mean number of RBs in the anti-inflammatory sesquiterpenoids from Asteraceae plants is 4.06, and the mean number of aromatic heavy atoms is 0.36. The mean values of RBs for polycyclic compounds, natural products, natural-product derivatives, and synthetic drugs are 7.4, 9.4, 7.4, and 5.4, respectively. The RBs for most of these sesquiterpenoids are below those of polycyclic natural products, thus indicating a good permeation rate (Figure 5f).

Fsp<sup>3</sup> is the ratio of sp<sup>3</sup> hybridized carbons to the total number of carbons [95]. Fsp<sup>3</sup> is used to characterize the carbon saturation, aliphatic degree, and complexity of the spatial structures of molecules: a higher Fsp<sup>3</sup> value indicates less aromatic character [95, 96]. The complexity of the spatial structure of molecules is highly correlated with target affinity and specificity. Molecules with more complex 3D shapes commonly possess higher druggability, whereas molecules with high aromaticity show planar features, which are not conducive to target binding. Compared with synthetic compounds (mean Fsp<sup>3</sup> of 0.27) and natural products (mean Fsp<sup>3</sup> of 0.55), the anti-inflammatory sesquiterpenoids exhibit a mean Fsp<sup>3</sup> of 0.68, indicating complex spatial characteristics, less aromatic character, and higher druggability [97].

### 3.5 Lipophilicity: logP

The parameter logP, also termed hydrophobicity, is important in drug discovery and design [98]. The logP is closely associated with water solubility, membrane permeability, bioactive potency, and target selectivity and heterogeneity, and it affects the pharmacokinetic and pharmacodynamic properties of compounds. Proper lipid solubility, low MW, and PSA are the main

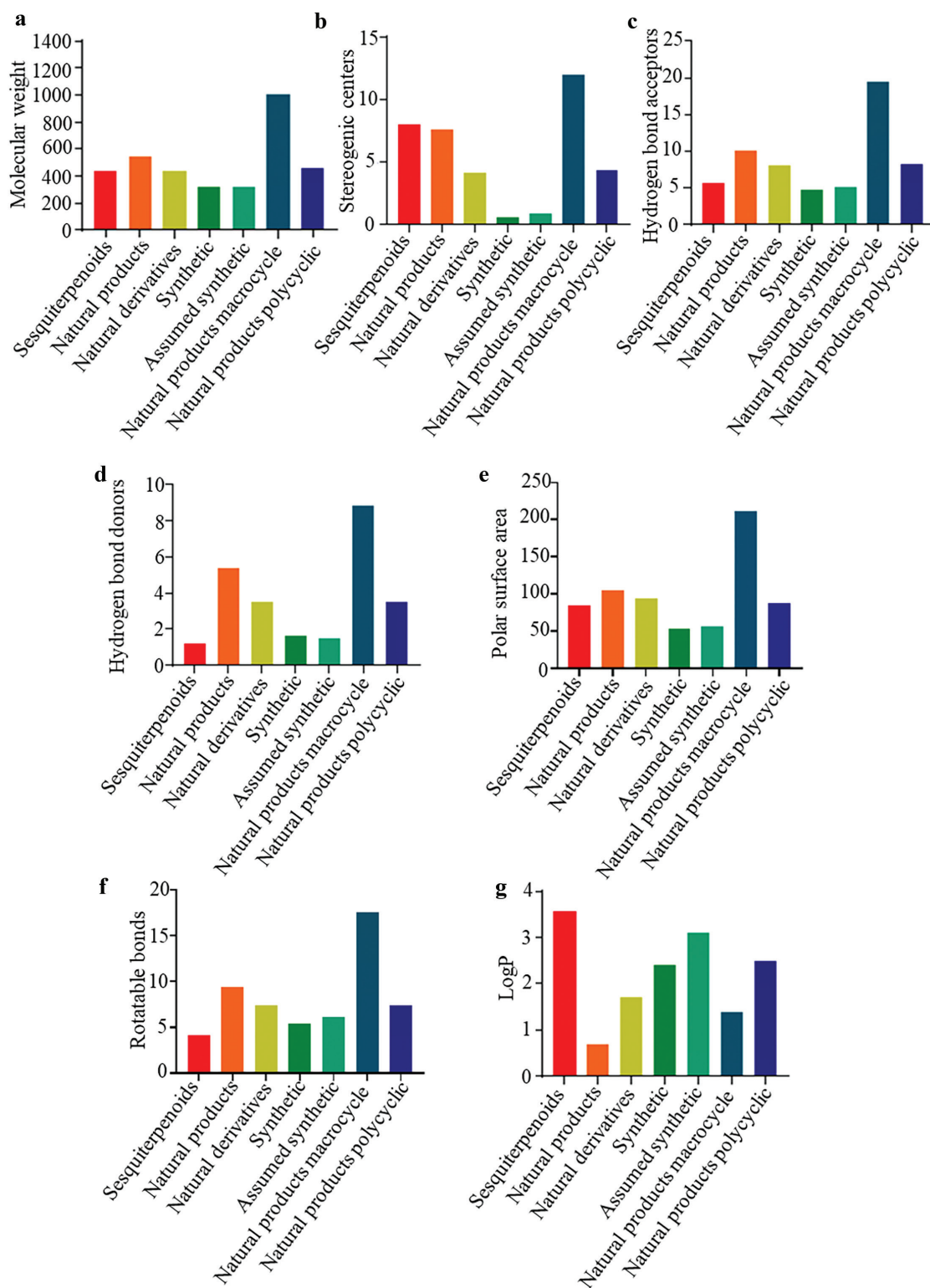
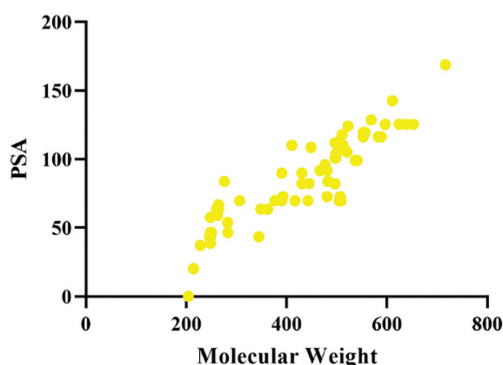


Figure 5 | Mean values of MW (a), stereogenic centers (b), HBA (c), HBD (d), PSA (e), RBs (f), and logP (g), for anti-inflammatory sesquiterpenoids from Asteraceae plants (red), natural products (orange), natural derivatives (yellowish brown), synthetic compounds (dark green), assumed synthetic compounds (aquamarine), macrocycle natural products (indigo), and polycyclic natural products (royal blue).

## Review Article



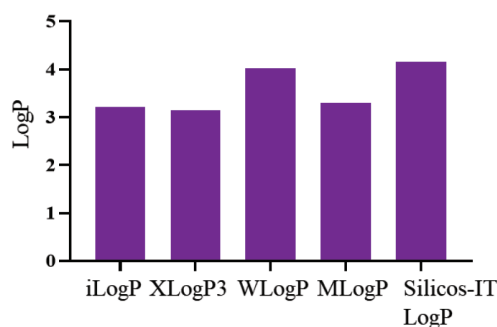
**Figure 6** | PSA values of the anti-inflammatory sesquiterpenoids from Asteraceae plants vs MW.

forces driving good oral absorption of a compound. Lipophilicity is the ratio between the equilibrium concentrations of a molecule in 1-octanol and water, commonly described as  $\log D$ , which represents the ratio for all species (unionized + ionized) of the molecule at a given pH value.  $D$  is therefore pH dependent for ionizable drugs. The distribution coefficient ( $D$ ) is replaced with a partition coefficient ( $P$ ) at any given pH if only one species (typically neutral) is present. The  $\log P$  value of an ideal drug is generally between 1 and 5 [99].

The  $\log P$  values of anti-inflammatory sesquiterpenoids from Asteraceae plants substantially vary depending on the predictive method in SwissADME. SwissADME supplies five available predictive models: 1)  $i\log P$ , an in-house physics-based method relying on free energies of solvation in  $n$ -octanol and water, calculated by the generalized Born and solvent-accessible surface area (GB/SA) model [98]; 2)  $X\log P3$ , an atomistic knowledge-based method [100]; 3)  $W\log P$ , a purely atomistic method based on the fragmental system [101]; 4)  $M\log P$ , an archetypal topological method relying on a linear relationship with 13 molecular descriptors [102]; and 5) Silicos-IT  $\log P$ , a hybrid method relying on 27 fragments and seven topological descriptors [79]. The calculated  $\log P$  values are averages of all five predictions. The  $\log P$  value of anti-inflammatory sesquiterpenoids (3.56) satisfies the criteria for good oral bioavailability (Figure 5g). For each sesquiterpenoid, we compared the  $\log P$  values obtained from the above five methods with the average  $\log P$  value. The greatest differences (maximum discreteness) of  $i\log P$ ,  $X\log P3$ ,  $W\log P$ ,  $M\log P$ , and Silicos-IT  $\log P$  were 18, 30, 7, 16, and 17, respectively. Thus,  $X\log P3$  is the most discrepant method among all  $\log P$  indexes (Figure 7 and Supplementary Information Table S2).

### 3.6 Solubility: $\log S$

To be absorbed into the body, drugs must be dissolved in water and be able to penetrate biofilms. The solubility of a compound is expressed in  $\log S$  (mol/L). The commonly used water solubility prediction methods include the free energy calculation method based on quantum



**Figure 7** | Mean  $\log P$  values of the anti-inflammatory sesquiterpenoids from Asteraceae plants, calculated with different methods.

mechanics and molecular mechanics, and the quantitative structure-activity relationship method, which has relatively high computational complexity [103]. The development of poorly water-soluble compounds would be risky and time-consuming. A compound with a solubility of at least 100  $\mu\text{g/mL}$  can be considered to have good water solubility.  $\log S$  values greater than -4 are acceptable for drugs. In fact, only 39% of anti-inflammatory sesquiterpenoids from Asteraceae plants meet this requirement, indicating that most compounds would face solubility challenges (Figure 8).

## 4. COMPLIANCE OF ANTI-INFLAMMATORY SESQUITERPENOID FROM ASTERACEAE PLANTS WITH DRUG-LIKENESS RULES

Drugs can be effectively assessed according to Lipinski's rules, as described above. Candidates with poor oral bioavailability can be quickly excluded. Most anti-inflammatory sesquiterpenoids from Asteraceae plants show good drug-likeness, according to the results of SwissADME global assessment (Supplementary Information Tables S1-S4). However, FDA-approved drugs with high MW and high  $\log P$  account for only 1%. Only a few oral active drugs violate two or more Lipinski's rule parameters. Recently, GlaxoSmithKline has considered additional physicochemical data for small drug molecules and discussed behavioral patterns, refinements, and implications of these five rules, providing new insights and principles for drug discovery [91]. According to ADMETlab 2.0 [104] prediction results, only 22% of these compounds simultaneously meet the Lipinski, Pfizer, GlaxoSmithKline, and Golden Triangle rules (Figure 9 and Supplementary Information Table S5).

## 5. TRENDS IN THE PHARMACOKINETIC BEHAVIOR OF ANTI-INFLAMMATORY SESQUITERPENOID FROM PLANTS OF THE ASTERACEAE FAMILY

Beyond efficacy and toxicity, a great number of failures in drug development are attributable to poor



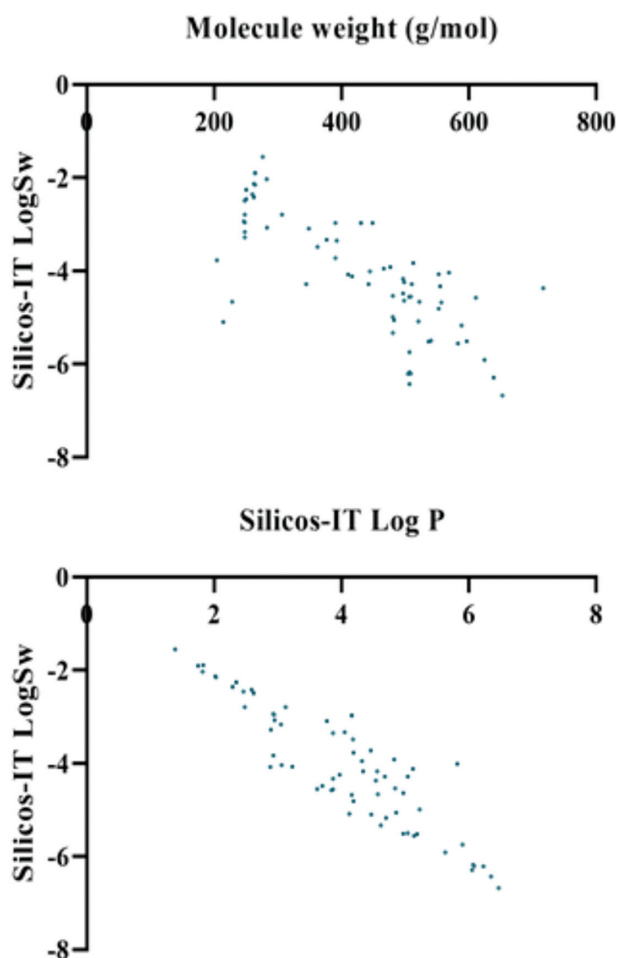


Figure 8 |  $\log S$  (SILICON-IT) of anti-inflammatory sesquiterpenoids from Asteraceae plants vs MW and  $\log P$  (SILICON-IT).

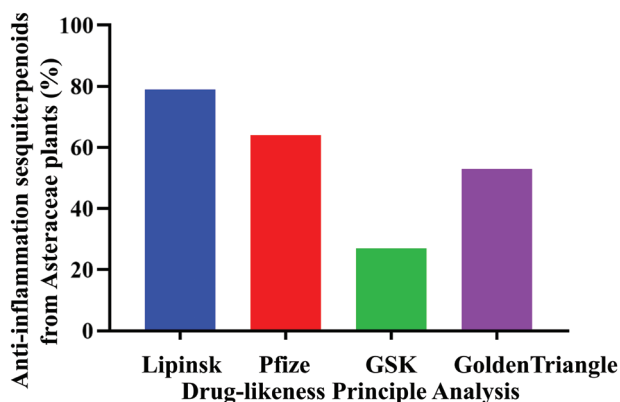


Figure 9 | Drug-likeness principle analysis of the sesquiterpenoids from Asteraceae plants.

pharmacokinetics and bioavailability. GI absorption and the BBB are critical considerations in various stages of the drug discovery process. BOILED-Egg was proposed

as an accurate predictive model that calculates the lipophilicity and polarity of small molecules [105]. The evaluation and optimization of active chemicals from the perspective of drug metabolism and pharmacokinetics, the selection of clinical candidates, and applications for new investigational drugs provide indispensable knowledge for scientific and management personnel involved in research and development of new drugs. These methods can be applied in a variety of settings, from chemical library filtering in the early stages of drug discovery to evaluation [106].

Approximately 82% of the anti-inflammatory sesquiterpenoids from Asteraceae plants exhibit a high probability of being absorbed in the GI (Figure 10a and Supplementary Information Figure S1). Approximately 72 sesquiterpenoids exhibit high GI absorption, possibly because of their relatively low MW. In addition, 51 sesquiterpenoids show a high probability of being a substrate for P-gp (Figure 10b). The BBB, formed by the brain capillary endothelium, is the most important barrier in the human body, and excludes 100% of macromolecular drugs and more than 98% of small-molecule drugs [107]. Most anti-inflammatory sesquiterpenoids possess a low probability of being able to cross the BBB (Figure 10c), suggesting the possibility of their development as neuroinflammatory drugs.

The pharmacokinetic properties of drugs are substantially determined by their physical and chemical properties. However, metabolic enzymes and transporters in the human body greatly affect the pharmacokinetic properties of drugs. Metabolic enzymes can metabolize drugs into more water-soluble metabolites, thus accelerating drug excretion from the body, whereas membrane transporters regulate the drug distribution in the body through the selective transport of some drugs. SwissADME indicates the potential ability of these sesquiterpenoids to act as P-gp substrates, thus inhibiting one of five major isoforms of cytochrome p450 (CYP450, including CYP1A2, CYP2C19, CYP2C9, CYP2D6, CYP3A4) [80, 108]. CYP450 is widely involved in drug metabolism. The predicted results were shown in Supplementary Information Table S4. The anti-inflammatory sesquiterpenoids from Asteraceae plants provide substantial opportunities to serve as CYP450 enzyme inhibitors, particularly for CYP2C9 (Figure 11). If a drug metabolized by CYP450 is used in combination with its inducer or inhibitor, the effectiveness and safety of drug interaction should be noted. If necessary, the dosage should be adjusted, or other drugs should be chosen.

## 6. TOXICITY

With ADMETlab 2.0, the toxicity of these sesquiterpenoids was predicted and analyzed (Supplementary Information Table S6). We selected three representative toxicity evaluation indicators: carcinogenicity, human Ether-a-go-go-associated Gene (hERG) toxicity,



## Review Article

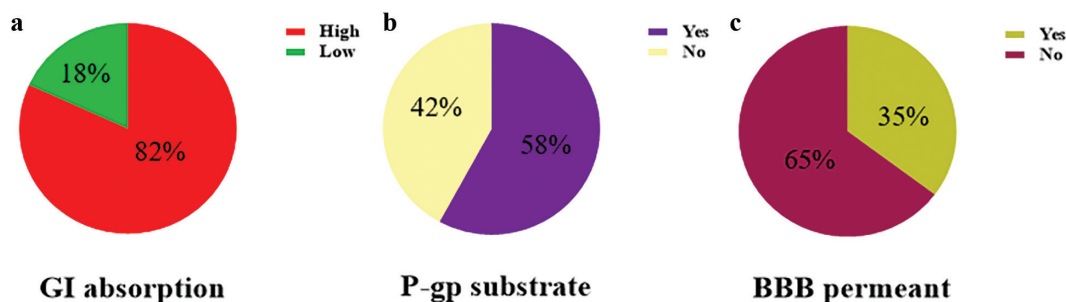


Figure 10 | (a) GI absorption of sesquiterpenoids from Asteraceae plants. (b) Classification of P-gp substrates for compounds with high GI absorption. (c) BBB permeability of sesquiterpenoids from Asteraceae plants.

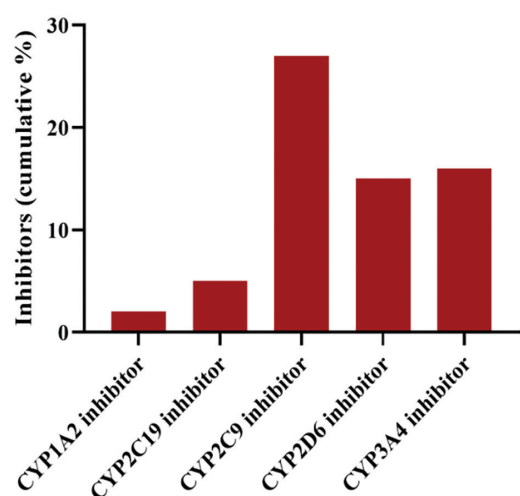


Figure 11 | Numbers of CYP450 enzyme inhibitors from the sesquiterpenoids from Asteraceae plants.

and drug-induced liver injury (DILI). Carcinogenicity is an important indicator in drug safety evaluation [109]. Carcinogenic compounds increase the incidence of tumors, thus severely threatening human health. The predictive models of the carcinogenicity of compounds can be classified into local models and global models. Local models focus on N-nitroso compounds,

aromatic amines, and polycyclic aromatic hydrocarbon equivalents. hERG is a human delayed rectifier potassium channel gene. The potassium channel encoded by this gene plays an important role in repolarization of the action potential. Many drugs increase the cardiac QT interval through their action on hERG potassium channels, resulting in serious adverse effects and potentially sudden death [110]. Several marketed drugs show a tendency to inhibit hERG, leading to a risk of adverse drug reactions associated with sudden death, such as terfenadine and cisapride. DILI is a common adverse drug reaction [111] and a serious threat to human health which is a major reason for failures in drug research and development, restriction of use, and withdrawal from the market. Because the molecular mechanism of drug-induced hepatotoxicity is complex, predicting DILI is very difficult. Bioinformatics and computational models based on drug molecular structure are widely used to predict drug-induced hepatotoxicity, such as the Bayesian method, deep learning, and the substructure pattern-recognition method. These models have yielded useful results in practical applications for predicting hepatotoxicity.

Approximately 44% of these sesquiterpenoids might exhibit carcinogenicity (Figure 12a), 9% sesquiterpenoids might possess hERG toxicity (Figure 12b), and, more importantly, 74% may cause hepatotoxicity. Critical attention should be paid to the toxicity of these compounds in pre-clinical research.

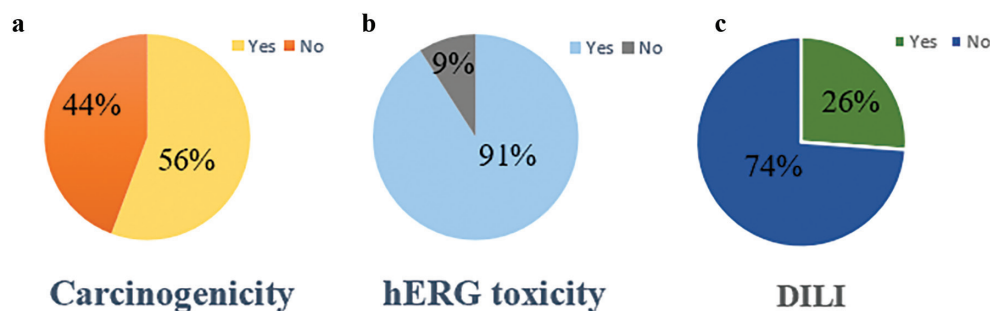


Figure 12 | Toxicity analysis of the sesquiterpenoids from Asteraceae plants.

## 7. CLINICAL RESEARCH

By searching the [ClinicalTrials.gov](https://www.clinicaltrials.gov) website, we identified four relevant clinical studies (compounds **50**, **51**, **84**, and **88**). Parthenolide (**50**) is to be developed in an allergic contact dermatitis detection panel. As part of the appetite suppressor in dietary supplement, the effects of lactucopicrin (**51**) in regulating incretin (GIP and GLP-1) and peptide hormones in human plasma have been evaluated. A total of 233 relevant clinical studies have investigated artemisinin (**84**), many of which have focused on its application in the treatment of malaria. Its effect on schizophrenia induced by *Toxoplasma gondii* infection was studied. Notably, artemisinin has been evaluated in the treatment of hospitalized patients with severe Coronavirus Disease 2019. The National Center for Complementary and Integrative Health at the University of Washington investigated whether  $\beta$ -caryophyllene (**88**) inhalation has positive feedback effects on the stress-reducing effects of forest bathing.

## 8. CONCLUSIONS AND PERSPECTIVES

Drug discovery is a complex interdisciplinary process. On average, only one of every 80,000–100,000 compounds is approved for marketing. Plants in the Asteraceae family are widely distributed and have been traditionally used for the treatment of inflammation-associated diseases. In this review, 88 anti-inflammatory sesquiterpenoids from plants in the Asteraceae family were summarized according to their carbon skeletons, origins, and pharmacological activities. Although many *in vitro* and *in vivo* studies have demonstrated these sesquiterpenoids with anti-inflammatory activities, practical applications of these compounds remain less. More clinical trials are necessary for development of these compounds as anti-inflammatory drugs.

In new drug research and development, most candidates fail during clinical trials, owing to low efficacy and selectivity, adverse pharmacokinetic properties, or unexpected adverse effects. Among these factors, pharmacokinetics and toxicity issues account for around 50% of failures in drug development. ADME determines the bioavailability, duration of action, and required dosage of a drug *in vivo*. In addition, toxicity is closely associated with pharmacokinetics, which is usually considered in combination with ADME, termed ADME/T. Owing to the poor consistency in findings between animal models and *in vitro* cellular models in evaluating the potential toxicity of candidate drugs, and the inability to simulate the complex environment in human body, the accuracy of ADME/T is low. With the development of combinatorial chemistry and high-throughput screening technology, extensive data have been generated, thus enabling progress in artificial intelligence and learning algorithms. Artificial intelligence and machine learning tools are increasingly being applied to predict drug pharmacokinetics properties and drug-induced toxicity. With

the large amount of *in vitro* toxicity data generated by high-throughput experiments, as well as the development of neural networks and learning algorithms, artificial intelligence may become a powerful tool for mining hidden information in big data, and play major roles in predicting the pharmacokinetics and toxicity of drug candidates.

Herein, the anti-inflammatory sesquiterpenoids were virtually predicted with SwissADME to obtain their ADME and drug-likeness properties. Many sesquiterpenoids show potential to become drug candidates for treating inflammatory diseases. More phytochemical, structural modification, and pharmaceuticals studies are needed to enhance potency and selectivity, improve bioavailability, and decrease toxicity. SwissADME is a powerful tool to predict the druggability of a molecule, and may decrease the likelihood of failure in clinical research.

## ACKNOWLEDGEMENTS

Financial support from the National Natural Science Foundation of China (81872754 and 82073715), Shanghai Pujiang Program (22PJD102), Science and Technology Development Fund, Macau SAR (file no. FDCT 0064/2021/AGJ and SKL-QRCM(UM)-2023-2025), Open Research Fund of Chengdu University of Traditional Chinese Medicine Key Laboratory of Systematic Research of Distinctive Chinese Medicine Resources in Southwest China (2022ZYXK2011007), National Key R&D Program "Strategic Scientific and Technological Innovation Cooperation" Key Project (2022YFE0203600) released by the Ministry of Science and Technology, and Research Fund of University of Macau (MYRG2020-00091-ICMS) is gratefully acknowledged.

## CONFLICTS OF INTEREST

The authors declare no competing financial interests.

## REFERENCES

- [1] Amor S, Puentes F, Baker D, van der Valk P: Inflammation in Neurodegenerative Diseases. *Immunology* 2010, 129:154–169.
- [2] Miller AH, Raison CL: The Role of Inflammation in Depression: From Evolutionary Imperative to Modern Treatment Target. *Nature Review Immunology* 2016, 16:22–34.
- [3] Danesh J, Whincup P, Walker M, Lennon L, Thomson A, Appleby P, et al.: Low Grade Inflammation and Coronary Heart Disease: Prospective Study and Updated Meta-Analyses. *British Medical Journal* 2000, 321:199–204.
- [4] Candore G, Caruso C, Jirillo E, Magrone T, Vasto S: Low Grade Inflammation as a Common Pathogenetic Denominator in Age-Related Diseases: Novel Drug Targets for Anti-Ageing Strategies and Successful Ageing Achievement. *Current Pharmaceutical Design* 2010, 16:584–596.
- [5] Cani PD, Osto M, Geurts L, Everard A: Involvement of Gut Microbiota in the Development of Low-Grade Inflammation and Type 2 Diabetes Associated with Obesity. *Gut Microbes* 2012, 3:279–288.
- [6] Miller GE, Cohen S, Ritchey AK: Chronic Psychological Stress and the Regulation of Pro-Inflammatory Cytokines: A Glucocorticoid-Resistance Model. *Health Psychology* 2002, 21:531–541.

## Review Article

- [7] Raghavendra V, Tanga FY, DeLeo JA: Complete Freund's Adjuvant-Induced Peripheral Inflammation Evokes Glial Activation and Proinflammatory Cytokine Expression in the CNS. *European Journal of Neuroscience* 2004, 20:467–473.
- [8] Christie GE, Allison HE, Kuzio J, McShan WM, Waldor MK, Kropinski AM: Prophage-Induced Changes in Cellular Cytochemistry and Virulence. In: Hyman P, Abedon ST, (Eds), *Bacteriophages in Health and Disease: Advances in Molecular and Cellular Microbiology*. CABI Publishing; 2012:33–60.
- [9] Gordon S: Alternative Activation of Macrophages. *Nature Review Immunology* 2003, 3:23–35.
- [10] Sunderkotter C, Steinbrink K, Goebeler M, Bhardwaj R, Sorg C: Macrophages and Angiogenesis. *Journal of Leukocyte Biology* 1994, 55:410–422.
- [11] Davies LC, Jenkins SJ, Allen JE, Taylor PR: Tissue-Resident Macrophages. *Nature Immunology* 2013, 14:986–995.
- [12] Imbert V, Peyron JF: NF-kappa B in Hematological Malignancies. *Biomedicines* 2017, 5:27.
- [13] Kim EK, Choi EJ: Compromised MAPK Signaling in Human Diseases: An Update. *Archives of Toxicology* 2015, 89:867–882.
- [14] Zheng XQ, Li K, Wei YD, Tie HT, Yi XY, Huang W: Nonsteroidal Anti-Inflammatory Drugs versus Corticosteroid for Treatment of Shoulder Pain: A Systematic Review and Meta-Analysis. *Archives of Physical Medicine and Rehabilitation* 2014, 95:1824–1831.
- [15] Wolfensberger TJ, Herbort CP: Treatment of Cystoid Macular Edema with Non-Steroidal Anti-Inflammatory Drugs and Corticosteroids. *Documenta Ophthalmologica* 1999, 97:381–386.
- [16] Rodríguez LAG, Hernández-Díaz S: The Risk of Upper Gastrointestinal Complications Associated with Nonsteroidal Anti-Inflammatory Drugs, Glucocorticoids, Acetaminophen, and Combinations of these Agents. *Arthritis Research and Therapy* 2001, 3:98.
- [17] Molinari GL: Natural Products in Drug Discovery: Present Status and Perspectives. *Advances in Experimental Medicine and Biology* 2009, 655:13–27.
- [18] Gerwick WH, Moore BS: Lessons from the Past and Charting the Future of Marine Natural Products Drug Discovery and Chemical Biology. *Chemical Biology* 2012, 19:85–98.
- [19] Roberts DJ, Hall RI: Drug Absorption, Distribution, Metabolism and Excretion Considerations in Critically Ill Adults. *Expert Opinion on Drug Metabolism and Toxicology* 2013, 9:1067–1084.
- [20] Zhang C, Huang CH, Liu M, Hu Y, Panero JL, Luebert F, et al.: Phylotranscriptomic Insights into Asteraceae Diversity, Polyploidy, and Morphological Innovation. *Journal of Integrative Plant Biology* 2021, 63:1273–1293.
- [21] Xiao-hui T, Jing H, Chao-xiang R, Jie S, Jin P, Qing-hua W, et al.: Analysis of Genetic Diversity and Relationship in Medicinal Plants of Compositae by ISSR. *Natural Products Research and Development* 2018, 30:1764–1768.
- [22] Juteau F, Masotti V, Bessière JM, Dherbomez M, Viano J: Antibacterial and Antioxidant Activities of *Artemisia annua* Essential Oil. *Fitoterapia* 2002, 73:532–535.
- [23] Ding A, Wen X: Dandelion Root Extract Protects NCM460 Colonic Cells and Relieves Experimental Mouse Colitis. *Journal of Natural Medicine* 2018, 72:857–866.
- [24] Li Y, Liu XJ, Su SL, Yan H, Guo S, Qian DW, et al.: Evaluation of Anti-Inflammatory and Antioxidant Effects of *Chrysanthemum* Stem and Leaf Extract on Zebrafish Inflammatory Bowel Disease Model. *Molecules* 2022, 27:2114.
- [25] Liu Q, Zhao H, Gao Y, Meng Y, Zhao XX, Pan SN: Effects of Dandelion Extract on the Proliferation of Rat Skeletal Muscle Cells and the Inhibition of a Lipopolysaccharide-Induced Inflammatory Reaction. *Chinese Medical Journal* 2018, 131:1724–1731.
- [26] Mouhid L, de Cedron MG, Vargas T, Garcia-Carrascosa E, Herranz N, Garcia-Risco M, et al.: Identification of Antitumoral Agents Against Human Pancreatic Cancer Cells from Asteraceae and Lamiaceae Plant Extracts. *BMC Complementary and Alternative Medicine* 2018, 18:254.
- [27] Michel J, Abd Rani NZ, Husain K: A Review on the Potential Use of Medicinal Plants From Asteraceae and Lamiaceae Plant Family in Cardiovascular Diseases. *Frontiers in Pharmacology* 2020, 11:1–26.
- [28] Soliman GA, Ansari MN, Alqarni MH, Foudah AI, Alam A, Salkini MA, et al.: Analgesic, Antipyretic, Anti-Inflammatory, and Hepatoprotective Activities of *Pulicaria crispa* (Forssk.) Oliv. (Asteraceae). *Brazilian Journal of Pharmaceutical Sciences* 2023, 59:e18851.
- [29] Aslan M, Orhan DD, Orhan N, Sezik E, Yesilada E: A Study of Antidiabetic and Antioxidant Effects of *Helichrysum graveolens* Capitulum in Streptozotocin-Induced Diabetic Rats. *Journal of Medical Food* 2007, 10:396–400.
- [30] Ugur A, Sarac N, Ceylan O, Duru ME: Chemical Composition of Endemic *Centaurea austro-anatolica* and Studies of its Antimicrobial Activity Against Multi-Resistant Bacteria. *Acta Pharmaceutica* 2009, 59:463–472.
- [31] Linh NTT, Ha NTT, Tra NT, Anh LT, Van Tuyen N, Son NT: Medicinal Plant *Centipeda minima*: A Resource of Bioactive Compounds. *Mini-Reviews in Medicinal Chemistry* 2021, 21:273–287.
- [32] Zhan ZJ, Ying YM, Ma LF, Shan WG: Natural Disesquiterpenoids. *Natural Product Reports* 2011, 28: 594–629.
- [33] Ma N, Zhang ZY, Liao FL, Jiang TL, Tu YY: The Birth of Artemisinin. *Pharmacology and Therapeutics* 2020, 216:107658.
- [34] Chen C, Ren YM, Zhu JZ, Chen JL, Feng ZL, Zhang T, et al.: Ainsliadimer C, a Disesquiterpenoid Isolated from *Ainsliaea macrocephala*, Ameliorates Inflammatory Responses in Adipose Tissue via Sirtuin 1-NLRP3 Inflammasome Axis. *Acta Pharmacologica Sinica* 2022, 43:1780–1792.
- [35] Dong T, Li C, Wang X, Dian L, Zhang X, Li L, et al.: Ainsliadimer A Selectively Inhibits IKK $\alpha/\beta$  by Covalently Binding a Conserved Cysteine. *Nature Communications* 2015, 6:6522.
- [36] Cheng XR, Zeng Q, Ren J, Qin JJ, Zhang SD, Shen YH, et al.: Sesquiterpene Lactones from *Inula falconeri*, a Plant Endemic to the Himalayas, as Potential Anti-Inflammatory Agents. *European Journal of Medicinal Chemistry* 2011, 46:5408–5415.
- [37] Zhu JX, Qin JJ, Jin HZ, Zhang WD: Japonicones Q-T, Four New Dimeric Sesquiterpene Lactones from *Inula japonica* Thunb. *Fitoterapia* 2013, 84:40–46.
- [38] Qin JJ, Wang LY, Zhu JX, Jin HZ, Fu JJ, Liu XF, et al.: Neojaponicone A, a Bioactive Sesquiterpene Lactone Dimer with an Unprecedented Carbon Skeleton from *Inula japonica*. *Chemical Communication* 2011, 47:1222–1224.
- [39] Zhang XF, Ren J, Cheng XR, Jin HZ, Zhang WD: One New Unusual Sesterterpenoid and Four New Sesquiterpene Dimers from *Inula britannica*. *RSC Advances* 2015, 5:1979–1982.

- [40] Chen LP, Wu GZ, Dong HY, Yang N, Zhang WD, Li HL: Linearifolianoids I-L, Four Rare Sesquiterpene Lactone Dimers Inhibiting NO Production from *Inula linearifolia*. *RSC Advances* 2016, 6:103296–103298.
- [41] Wang Q, Zhang T, Ke C-Q, Tang C, Yao S, Lin L, et al.: Guaianolides from *Artemisia codonocephala* Suppress Interleukine-1 $\beta$  Secretion in Macrophages. *Phytochemistry* 2021, 192:112955.
- [42] Zeng KW, Wang S, Dong X, Jiang Y, Jin HW, Tu PF: Sesquiterpene Dimmer (DSF-27) Inhibits the Release of Neuroinflammatory Mediators from Microglia by Targeting Spleen Tyrosine Kinase (Syk) and Janus Kinase 2 (Jak2): Two Major Non-Receptor Tyrosine Signaling Proteins Involved in Inflammatory Events. *Toxicological and Applied Pharmacology* 2014, 275:244–256.
- [43] Zhang C, Wen R, Ma XL, Zeng KW, Xue Y, Zhang PM, et al.: Nitric Oxide Inhibitory Sesquiterpenoids and its Dimers from *Artemisia freyniana*. *Journal of Natural Products* 2018, 81:866–878.
- [44] Turak A, Shi SP, Jiang Y, Tu PF: Dimeric Guaianolides from *Artemisia absinthium*. *Phytochemistry* 2014, 105:109–114.
- [45] Zhang C, Wang S, Zeng KW, Li J, Ferreira D, Zjawiony JK, et al.: Nitric Oxide Inhibitory Dimeric Sesquiterpenoids from *Artemisia rupestris*. *Journal of Natural Products* 2016, 79:213–223.
- [46] Siedle B, Garcia-Pineres AJ, Murillo R, Schulte-Monting J, Castro V, Rungeler P, et al.: Quantitative Structure-Activity Relationship of Sesquiterpene Lactones as Inhibitors of the Transcription Factor NF-kappa B. *Journal of Medicinal Chemistry* 2004, 47:6042–6054.
- [47] Malarz J, Stojakowska A, Kisiel W: Sesquiterpene Lactones in a Hairy Root Culture of *Cichorium intybus*. *Zeitschrift für Naturforschung C* 2002, 57:994–997.
- [48] Cavin C, Delannoy M, Malnoe A, Debefve E, Touche A, Courtois D, et al.: Inhibition of the Expression and Activity of Cyclooxygenase-2 by Chicory Extract. *Biochemical and Biophysical Research Communications* 2005, 327:742–749.
- [49] Vanbeek TA, Maas P, King BM, Leclercq E, Voragen AGJ, Degroot A: Bitter Sesquiterpene Lactones from Chicory Roots. *Journal of Agricultural and Food Chemistry* 1990, 38:1035–1038.
- [50] Jang YJ, Back MJ, Fu Z, Lee JH, Won JH, Ha HC, et al.: Protective Effect of Sesquiterpene Lactone Parthenolide on LPS-Induced Acute Lung Injury. *Archives of Pharmacal Research* 2016, 39:1716–1725.
- [51] Chadwick M, Trewin H, Gawthrop F, Wagstaff C: Sesquiterpenoids Lactones: Benefits to Plants and People. *International Journal of Molecular Sciences* 2013, 14:12780–12805.
- [52] Ren Y, Zhou S, Zhang T, Qian M, Zhang R, Yao S, et al.: Targeted Isolation of Two Disesquiterpenoids Macrocephadiolides A and B from *Ainsliaea macrocephala* using Molecular Networking-Based Dereplication Strategy. *Organic Chemistry Frontiers* 2020, 7:1481–1489.
- [53] Qin JJ, Wang W, Zhang RW: Novel Natural Product Therapeutics Targeting Both Inflammation and Cancer. *Chinese Journal of Natural Medicines* 2017, 15:401–416.
- [54] Maas M, Deters AM, Hensel A: Anti-Inflammatory Activity of *Eupatorium perfoliatum* L. Extracts, Eupafolin, and Dimeric Guaianolide via iNOS Inhibitory Activity and Modulation of Inflammation-Related Cytokines and Chemokines. *Journal of Ethnopharmacology* 2011, 137:371–381.
- [55] Chen LP, Wu GZ, Zhang JP, Ye J, Liu QX, Shen YH, et al.: Vlasouliolides A-D, Four Rare C17/C15 Sesquiterpene Lactone Dimers with Potential Anti-Inflammatory Activity from *Vladimiria souliei*. *Scientific Reports* 2017, 7:43837.
- [56] Wu ZL, Wang JX, Chen LP, Dong HY, Li HL, Zhang WD: Five Rare C-32 Sesquiterpene Lactone Dimers with Anti-Inflammation Activity from *Vladimiria souliei*. *Fitoterapia* 2018, 125:117–122.
- [57] Lee J, Kang U, Seo EK, Kim YS: Heme Oxygenase-1-Mediated Anti-Inflammatory Effects of Tussilagonone on Macrophages and 12-O-Tetradecanoylphorbol-13-Acetate-Induced Skin Inflammation in Mice. *International Immunopharmacology* 2016, 34:155–164.
- [58] Li W, Huang X, Yang XW: New Sesquiterpenoids from the Dried Flower Buds of *Tussilago farfara* and their Inhibition on NO Production in LPS-Induced RAW264.7 Cells. *Fitoterapia* 2012, 83:318–322.
- [59] Jang H, Lee JW, Lee C, Jin Q, Choi JY, Lee D, et al.: Sesquiterpenoids from *Tussilago farfara* Inhibit LPS-Induced Nitric Oxide Production in Macrophage RAW 264.7 Cells. *Archives of Pharmacal Research* 2016, 39:127–132.
- [60] Hwangbo C, Lee HS, Park J, Choe J, Lee JH: The Anti-Inflammatory Effect of Tussilagone, from *Tussilago farfara*, is Mediated by the Induction of Heme Oxygenase-1 in Murine Macrophages. *International Immunopharmacology* 2009, 9:1578–1584.
- [61] Ryu JH, Jeong JS, Sohn DH: A New Bisabolene Epoxide from *Tussilago farfara*, and Inhibition of Nitric Oxide Synthesis in LPS-Activated Macrophages. *Journal of Natural Products* 1999, 62:1437–1438.
- [62] Qin ZB, Zhang J, Wu XD, He J, Ding LF, Peng LY, et al.: Sesquiterpenoids from *Tussilago farfara* and their Inhibitory Effects on Nitric Oxide Production. *Planta Medica* 2014, 80:703–709.
- [63] Zafrapolo MC, Blazquez MA: Antiinflammatory Activity of Sesquiterpene Lactones from *Artemisia barrelieri* in Rats. *Phytotherapy Research* 1991, 5:91–93.
- [64] Wang KS, Li JB, Wang Z, Mi CL, Ma J, Piao LX, et al.: Artemisinin Inhibits Inflammatory Response via Regulating NF- $\kappa$ B and MAPK Signaling Pathways. *Immunopharmacology and Immunotoxicology* 2017, 39:28–36.
- [65] Egas V, Toscano RA, Linares E, Bye R, Espinosa-Garcia FJ, Delgado G: Cadinane-Type Sesquiterpenoids from *Heterotheca inuloides*: Absolute Configuration and Anti-Inflammatory Activity. *Journal of Natural Products* 2015, 78:2634–2641.
- [66] Delgado G, del Socorro Olivares M, Chávez MI, Ramírez-Apan T, Linares E, Bye R, et al.: Antiinflammatory Constituents from *Heterotheca inuloides*. *Journal of Natural Products* 2001, 64:861–864.
- [67] Cho JY, Kim HY, Kim S-K, Park JHY, Lee HJ, Chun HS:  $\beta$ -Caryophyllene Attenuates Dextran Sulfate Sodium-Induced Colitis in Mice via Modulation of Gene Expression Associated Mainly with Colon Inflammation. *Toxicology Reports* 2015, 2:1039–1045.
- [68] Bento AF, Marcon R, Dutra RC, Claudino RF, Cola M, Leite DF, et al.:  $\beta$ -Caryophyllene Inhibits Dextran Sulfate Sodium-Induced Colitis in Mice through CB2 Receptor Activation and PPAR $\gamma$  Pathway. *American Journal of Pathology* 2011, 178:1153–1166.
- [69] Shin EM, Zhou HY, Guo LY, Kim JA, Lee SH, Merfort I, et al.: Anti-Inflammatory Effects of Glycyrol Isolated from *Glycyrrhiza uralensis* in LPS-Stimulated RAW264.7 Macrophages. *International Immunopharmacology* 2008, 8:1524–1532.



## Review Article

- [70] Fan GW, Zhang Y, Jiang X, Zhu Y, Wang B, Su L, et al.: Anti-Inflammatory Activity of Baicalein in LPS-Stimulated RAW264.7 Macrophages via Estrogen Receptor and NF- $\kappa$ B-Dependent Pathways. *Inflammation* 2013, 36:1584–1591.
- [71] Xiao W, Li X, Li N, Bolati M, Wang XJ, Jia XG, et al.: Sesquiterpene Lactones from *Saussurea involucreta*. *Fitoterapia* 2011, 82:983–987.
- [72] Xue GM, Li XQ, Chen C, Chen K, Wang X-B, Gu YC, et al.: Highly Oxidized Guaianolide Sesquiterpenoids with Potential Anti-Inflammatory Activity from *Chrysanthemum indicum*. *Journal of Natural Products* 2018, 81:378–386.
- [73] Li D, Yang C, Zhu JZ, Lopez E, Zhang T, Tong Q, et al.: Berberine Remodels Adipose Tissue to Attenuate Metabolic Disorders by Activating Sirtuin 3. *Acta Pharmacologica Sinica* 2022, 43:1285–1298.
- [74] Rüngeler P, Lyss G, Castro V, Mora G, Pahl HL, Merfort I: Study of Three Sesquiterpene Lactones from *Tithonia diversifolia* on their Anti-Inflammatory Activity using the Transcription Factor NF-kappa B and Enzymes of the Arachidonic Acid Pathway as Targets. *Planta Medica* 1998, 64:588–593.
- [75] Sang N, Zhang J, Marcheselli V, Bazan NG, Chen C: Postsynaptically Synthesized Prostaglandin E2 (PGE2) Modulates Hippocampal Synaptic Transmission via a Presynaptic PGE2 EP2 Receptor. *Journal of Neurosciences* 2005, 25:9858–9870.
- [76] Ricciotti E, FitzGerald GA: Prostaglandins and Inflammation. *Arteriosclerosis, Thrombosis, and Vascular Biology* 2011, 31:986–1000.
- [77] Rao TS, Currie JL, Shaffer AF, Isakson PC: Comparative Evaluation of Arachidonic Acid (AA)- and Tetradecanoylphorbol Acetate (TPA)-Induced Dermal Inflammation. *Inflammation* 1993, 17:723–741.
- [78] Rodríguez-Chávez JL, Coballase-Urrutia E, Nieto-Camacho A, Delgado-Lamas G: Antioxidant Capacity of “Mexican arnica” *Heterotheca inuloides* Cass Natural Products and Some Derivatives: Their Anti-Inflammatory Evaluation and Effect on *C. elegans* Life Span. *Oxidative Medicine and Cellular Longevity* 2015, 2015:843237.
- [79] Daina A, Michielin O, Zoete V: SwissADME: A Free Web Tool to Evaluate Pharmacokinetics, Drug-Likeness and Medicinal Chemistry Friendliness of Small Molecules. *Scientific Reports* 2017, 7:42717.
- [80] Boulton DW, DeVane CL, Liston HL, Markowitz JS: *In vitro* P-glycoprotein affinity for atypical and conventional antipsychotics. *Life Sciences* 2002, 71:163–169.
- [81] Schroder A, Klein K, Winter S, Schwab M, Bonin M, Zell A, et al.: Genomics of ADME Gene Expression: Mapping Expression Quantitative Trait Loci Relevant for Absorption, Distribution, Metabolism and Excretion of Drugs In Human Liver. *Pharmacogenomics Journal* 2013, 13:12–20.
- [82] Hughes LD, Palmer DS, Nigsch F, Mitchell JBO: Why are Some Properties More Difficult to Predict Than Others? A Study of QSPR Models of Solubility, Melting Point, and Log P. *Journal of Chemical Information and Modeling* 2008, 48:220–232.
- [83] Ebeling H, Edge AC, Bohringer H, Allen SW, Crawford CS, Fabian AC, et al.: The ROSAT Brightest Cluster Sample - I. The Compilation of the Sample and the Cluster log N-log S Distribution. *Monthly Notices of the Royal Astronomical Society* 1998, 301:881–914.
- [84] Bade R, Chan H-F, Reynisson J: Characteristics of Known Drug Space. Natural Products, their Derivatives and Synthetic Drugs. *European Journal of Medicinal Chemistry* 2010, 45:5646–5652.
- [85] Camp D, Garavelas A, Campitelli M: Analysis of Physicochemical Properties for Drugs of Natural Origin. *Journal of Natural Products* 2015, 78:1370–1382.
- [86] Raymer B, Bhattacharya SK: Lead-Like Drugs: A Perspective. *Journal of Medicinal Chemistry* 2018, 61:10375–10384.
- [87] Agranat I, Caner H, Caldwell A: Putting Chirality to Work: The Strategy of Chiral Switches. *Nature Review Drug Discovery* 2002, 1:753–768.
- [88] Feng ZL, Lu XQ, Gan LS, Zhang QW, Lin LG: Xanthenes, a Promising Anti-Inflammatory Scaffold: Structure, Activity, and Drug Likeness Analysis. *Molecules* 2020, 25:598.
- [89] Feng Z, Cao J, Zhang Q, Lin L: The Drug Likeness Analysis of Anti-Inflammatory Clerodane Diterpenoids. *Chinese Medicine* 2020, 15:126.
- [90] Prasanna S, Doerksen RJ: Topological Polar Surface Area: A Useful Descriptor in 2D-QSAR. *Current Medicinal Chemistry* 2009, 16:21–41.
- [91] Tinworth CP, Young RJ: Facts, Patterns, and Principles in Drug Discovery: Appraising the Rule of 5 with Measured Physicochemical Data. *Journal of Medicinal Chemistry* 2020, 63:10091–10108.
- [92] Tarko L: Using the Bond Order Calculated by Quantum Mechanics to Identify the Rotatable Bonds. *Revista de Chimie* 2011, 62:135–138.
- [93] Veber DF, Johnson SR, Cheng HY, Smith BR, Ward KW, Kopple KD: Molecular Properties that Influence the Oral Bioavailability of Drug Candidates. *Journal of Medicinal Chemistry* 2002, 45:2615–2623.
- [94] Bhutani P, Joshi G, Raja N, Bachhav N, Rajanna PK, Bhutani H, et al.: US FDA Approved Drugs from 2015–June 2020: A Perspective. *Journal of Medicinal Chemistry* 2021, 64:2339–2381.
- [95] Lovering F, Bikker J, Humblet C: Escape from Flatland: Increasing Saturation as an Approach to Improving Clinical Success. *Journal of Medicinal Chemistry* 2009, 52:6752–6756.
- [96] Yan A, Gasteiger J: Prediction of Aqueous Solubility of Organic Compounds Based on a 3D structure representation. *Journal of Chemical Information and Computer Sciences* 2003, 43:429–434.
- [97] Meanwell NA: Improving Drug Candidates by Design: A Focus on Physicochemical Properties as a Means of Improving Compound Disposition and Safety. *Chemical Research in Toxicology* 2011, 24:1420–1456.
- [98] Daina A, Michielin O, Zoete V: iLOGP: A Simple, Robust, and Efficient Description of n-octanol/Water Partition Coefficient for Drug Design using the GB/SA Approach. *Journal of Chemical Information and Modeling* 2014, 54:3284–3301.
- [99] Box KJ, Comer JE: Using Measured pKa, LogP and Solubility to Investigate Supersaturation and Predict BCS Class. *Current Drug Metabolism* 2008, 9:869–878.
- [100] Cheng T, Zhao Y, Li X, Lin F, Xu Y, Zhang X, et al.: Computation of Octanol–Water Partition Coefficients by Guiding an Additive Model with Knowledge. *Journal of Chemical Information and Modeling* 2007, 47:2140–2148.
- [101] Wildman SA, Crippen GM: Prediction of Physicochemical Parameters by Atomic Contributions. *Journal of Chemical Information and Computer Sciences* 1999, 39:868–873.
- [102] Moriguchi I, Hirono S, Liu Q, Nakagome I, Matsushita Y: Simple Method of Calculating Octanol/Water Partition Coefficient. *Chemical and Pharmaceutical Bulletin* 1992, 40:127–130.

- [103] Jorgensen WL, Duffy EM: Prediction of Drug Solubility from Structure. *Advanced Drug Delivery Reviews* 2002, 54:355–366.
- [104] Xiong GL, Wu ZX, Yi JC, Fu L, Yang ZJ, Hsieh CY, et al.: ADMETlab 2.0: An Integrated Online Platform for Accurate and Comprehensive Predictions of ADMET Properties. *Nucleic Acids Research* 2021, 49:W5–W14.
- [105] Daina A, Zoete V: A BOILED-Egg to Predict Gastrointestinal Absorption and Brain Penetration of Small Molecules. *ChemMedChem* 2016, 11:1117–1121.
- [106] Bode H, Brendel E, Ahr G, Fuhr U, Harder S, Staib AH: Investigation of Nifedipine Absorption in Different Regions of the Human Gastrointestinal (GI) Tract After Simultaneous Administration of <sup>13</sup>C- and <sup>12</sup>C-Nifedipine. *European Journal of Clinical Pharmacology* 1996, 50:195–201.
- [107] Wilhelm I, Krizbai IA: In Vitro Models of the Blood–Brain Barrier for the Study of Drug Delivery to the Brain. *Molecular Pharmaceutics* 2014, 11:1949–1963.
- [108] Brewer CT, Chen T: Hepatotoxicity of Herbal Supplements Mediated by Modulation of Cytochrome P450. *International Journal of Molecular Sciences* 2017, 18:2353.
- [109] Corvi R, Spielmann H, Hartung T: Chapter 3.6 - Alternative Approaches for Carcinogenicity and Reproductive Toxicity. In: Balls M, Combes R, Worth A, (Eds), *The History of Alternative Test Methods in Toxicology*. Academic Press; 2019:209–217.
- [110] Garrido A, Lepailleur A, Mignani SM, Dallemagne P, Rochais C: hERG Toxicity Assessment: Useful Guidelines for Drug Design. *European Journal of Medicinal Chemistry* 2020, 195:112290.
- [111] Andrade RJ, Chalasani N, Bjornsson ES, Suzuki A, Kullak-Ublick GA, Watkins PB, et al.: Drug-Induced Liver Injury. *Nature Reviews Disease Primers* 2019, 5:58.