

Protective effect of delayed remote limb ischemic postconditioning: role of mitochondrial K_{ATP} channels in a rat model of focal cerebral ischemic reperfusion injury

Jing Sun^{1,3}, Tong Li^{1,3}, Qi Luan^{2,3}, Jiao Deng¹, Yan Li¹, Zhaoju Li¹, Hailong Dong¹ and Lize Xiong¹

¹Department of Anesthesiology, Xijing Hospital, Fourth Military Medical University, Shaanxi Province, People's Republic of China; ²Department of Dermatology, Xijing Hospital, Fourth Military Medical University, Shaanxi Province, People's Republic of China

Delayed remote ischemic postconditioning (DRIPost) has been shown to protect the rat brain from ischemic injury. However, extremely short therapeutic time windows hinder its translational use and the mechanism of action remains elusive. Because opening of the mitochondria K_{ATP} channel is crucial for cell apoptosis, we hypothesized that the neuroprotective effect of DRIPost may be associated with K_{ATP} channels. In the present study, the neuroprotective effects of DRIPost were investigated using adult male Sprague-Dawley rats. Rats were exposed to 90 minutes of middle cerebral artery occlusion followed by 72 hours of reperfusion. Delayed remote ischemic postconditioning was performed with three cycles of bilateral femoral artery occlusion/reperfusion for 5 minutes at 3 or 6 hours after reperfusion. Neurologic deficit scores and infarct volumes were assessed, and cellular apoptosis was monitored by terminal deoxynucleotidyl transferase nick-end labeling. Our results showed that DRIPost applied at 6 hours after reperfusion exerted neuroprotective effects. The K_{ATP} opener, diazoxide, protected rat brains from ischemic injury, while the K_{ATP} blocker, 5-hydroxydecanote, reversed the neuroprotective effects of DRIPost. These findings indicate that DRIPost reduces focal cerebral ischemic injury and that the neuroprotective effects of DRIPost may be achieved through opening of K_{ATP} channels.

Journal of Cerebral Blood Flow & Metabolism (2012) 32, 851–859; doi:10.1038/jcbfm.2011.199; published online 25 January 2012

Keywords: brain ischemia; K_{ATP} ; remote ischemic postconditioning; reperfusion injury

Introduction

Stroke is the third leading cause of death in the United States. Approximately 795,000 people experience a stroke and >143,579 people die each year. Of all strokes, 87% are ischemic (Lloyd-Jones *et al*, 2010). The current lack of clinical treatment for acute stroke necessitates the exploration of novel concepts that may eventually lead to clinical application.

One of these concepts is ischemic postconditioning (IPost) (Pignataro *et al*, 2008a; Zhao *et al*, 2006), which refers to interference of blood flow by a series of brief, repetitive occlusion and release of cerebral blood vessels after reperfusion. Yang *et al* (2004) reported that reduction to infarct size by IPost was dependent on opening of K_{ATP} channels. However, the extremely short therapeutic time windows may hinder its clinical translation. This specific limitation may prevent its application to those patients in whom reperfusion cannot be immediately and accurately identified.

However, it has been reported that delayed postconditioning conducted 2 days after transient global ischemia attenuates hippocampal injury in gerbils (Burda *et al*, 2006). In addition, Ren *et al* (2008) reported that delayed postconditioning reduced ischemic injury after focal ischemia. Recently, a new phenomenon, known as remote IPost (RIPost), was found to induce ischemic tolerance not only within the same piece of tissue, but also in distant tissues as well as

Correspondence: Dr L Xiong, Department of Anesthesiology, Xijing Hospital, Fourth Military Medical University, Xi'an, Shaanxi Province, People's Republic of China. E-mail: mzklz@126.com

³These authors contributed equally to this work.

This study was supported by the National Natural Science Foundation of China (Beijing, China, grants 30772059, 30972853, and 81128005 to HD), the National Science Foundation for Distinguished Young Scholars (Beijing, China, grant 30725039 to LX), and the Major Program of National Natural Science Foundation of China (Beijing, China, grant 30930091 to LX).

Received 21 August 2011; revised 6 December 2011; accepted 8 December 2011; published online 25 January 2012

in distant organs (Tsubota *et al*, 2010). Loukogeorgakis *et al* (2007) revealed that transient limb ischemia induces RIpost in humans by a K_{ATP} channel-dependent mechanism. This result suggested that mitochondrial K_{ATP} channel activation has a key role in the development of a protective effect during IPost and RIpost. RIpost has greater potential for clinical application than classic IPost as it can be performed in a nonvital organ, avoiding the high risk of IPost in the vital organ. However, all reported IPost procedures have been applied either at the onset of reperfusion or during the ischemic phase. For patients with transient ischemic reperfusion brain injury, pharmacologic and physiologic interventions influence the effective time window of treatment. However, whether delayed remote IPost (DRIPost) attenuates brain injury after cerebral ischemia is unknown. In this study, we tested whether DRIPost, conducted by repetitive occlusion and release of the bilateral femoral arteries, reduces the infarcted area in focal ischemic rats. Accumulating evidence suggests that K_{ATP} channel activation is involved in the protective effect of IPost. Thus, we further tested whether DRIPost protects the brain from stroke and the potential protective mechanisms related with K_{ATP} channel activation.

Materials and methods

Animals

Male Sprague-Dawley rats weighing 290 to 310 g were provided by the Experimental Animal Centre of the Fourth Military Medical University and housed under diurnal lighting conditions (12 hours darkness/light). All experimental protocols and animal handling procedures were performed in accordance with the National Institutes of Health (NIH, USA) guidelines for the use of experimental animals and the experimental protocols were approved by the Institutional Animal Care and Use Committee of the Fourth Military Medical University. Diazoxide (DIAZ) and 5-hydroxydecanote (5-HD) were purchased from Sigma-Aldrich (St Louis, MO, USA).

Focal Cerebral Ischemia

Sprague-Dawley rats were allowed free access to food and water but were fasted 12 hours before surgery. All animals were anesthetized by intraperitoneal injection of pentobarbital sodium (50 mg/kg in normal saline). The transient middle cerebral artery (MCA) occlusion model was performed as previously described (Xiong *et al*, 2003). Briefly, the right common carotid artery (CCA) and the right external carotid artery were exposed through a ventral midline neck incision, and were ligated proximally. A 3-0 nylon monofilament suture (Ethicon Inc., Osaka, Japan) with a blunt tip made by burning on a flame was inserted through the arterectomy in the CCA just below the carotid bifurcation, positioned into the internal carotid artery and advanced ~17 to 18 mm until a mild resistance was felt. Under these

conditions, the origin of the MCAs was occluded. Reperfusion was accomplished by withdrawing the suture after 90 minutes of ischemia. The incision sites were infiltrated with 0.25% (v/v) bupivacaine hydrochloride for post-operative analgesia.

Regional cerebral blood flow was monitored using a flexible optical fiber probe attached to the skull over the ipsilateral parietal cortex at one point (1 mm posterior and 5 mm lateral to bregma) by laser Doppler flowmetry (PeriFlux system 5000; Perimed AB, Stockholm, Sweden). Rats in which ipsilateral blood flow was not reduced to <20% of the baseline after placement of the intraluminal filament and whose cerebral blood flow signal was not rapidly restored during reperfusion were excluded from subsequent experiments. Cranial temperature was maintained at 36.8°C to 37.5°C by surface heating and cooling during surgery. In a separate experiment, physiologic parameters (cranial temperature, arterial pH, PaCO₂, PaO₂, and glucose) were monitored and analyzed in five additional rats. Arterial blood samples were taken 3 minutes before ischemia (baseline), 45 minutes after ischemia, and 30 minutes after reperfusion and DRIPost for gases and plasma glucose measurements.

Animal Recovery and Neurologic Evaluation

Rats were returned to their cages after the suture was withdrawn and were given free access to food and water. The neurologic behavior of rats was scored at 24, 48, and 72 hours after reperfusion by an investigator who was unaware of animal grouping. An 18-point scale of neurologic deficit scores (NDSs) was used for evaluation of neurologic behavior (Garcia *et al*, 1997). The scale was based on the following six tests: (1) spontaneous activity (0 to 3 points); (2) symmetry in the movement of four limbs (0 to 3 points); (3) forepaw outstretching (0 to 3 points); (4) climbing (1 to 3 points); (5) body proprioception (1 to 3 points); and (6) response to vibrissae touch (1 to 3 points). The six individual test scores were summed up at the end of the evaluation (minimum score, 3; maximum score, 18).

Infarct Volume Measurement

For measurements of infarct volume at 72 hours after cerebral artery reperfusion, rats were killed and the brains were rapidly removed and mildly frozen to keep the morphology intact during slicing. In brief, brains were cut into 2-mm-thick coronal sections in a brain matrix and stained with 2% (w/v) 2,3,5-triphenyltetrazolium chloride (Sigma-Aldrich) for 30 minutes at 37°C followed by overnight immersion in 4% (w/v) paraformaldehyde in phosphate buffer for after fixation. The infarct tissue area remained unstained (white), whereas normal tissue was stained red. Photographs were taken using a digital camera (SONY T9; Sony Corporation, Tokyo, Japan) The infarct area was demarcated and analyzed by Photoshop software (Adobe Photoshop 8.0, Adobe Systems Incorporated, San Jose, CA, USA). To compensate for the effect of brain edema, the corrected infarct volume was calculated as follows: percentage of corrected infarct volume = ((contralateral hemisphere

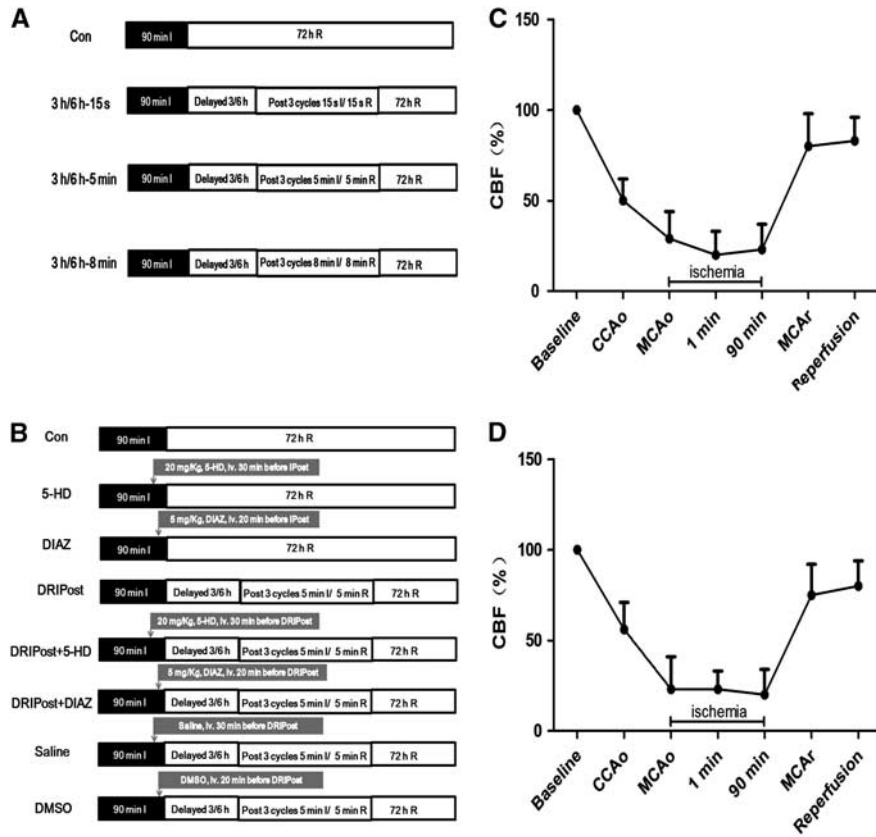


Figure 1 Schematic of the design and changes in cerebral blood flow (CBF) in animals for part 1 and part 2 of the experiment. All animals underwent 90 minutes of ischemia (dark bar) and 72 hours of reperfusion (open bar) ($n = 8$ for each). **(A)** In part 1, rats were subjected to delayed remote ischemic postconditioning (DRIPost) at 3 or 6 hours after reperfusion, and the DRIPost protocol was three cycles of bilateral femoral artery occlusion 15 seconds/5 minutes/8 minutes ischemia (I)/15 seconds/5 minutes/8 minutes reperfusion (R). **(B)** In part 2, except for delayed 6 hours/5 minutes DRIPost, the K_{ATP} channel blocker 5-hydroxydecanote (5-HD) or opener diazoxide (DIAZ), as well as the vehicle physiologic saline and dimethyl sulfoxide (DMSO), was injected through the caudal vein 20 minutes (for 5-HD and saline) or 30 minutes (for DIAZ and DMSO) before DRIPost to investigate the role of K_{ATP} channels in DRIPost-induced neuroprotection against focal cerebral ischemia–reperfusion injury. **(C, D)** Changes in CBF in animals subjected to 90 minutes ischemia followed by 72 hours reperfusion. Right common carotid artery (CCA) occlusion reduced CBF to $\sim 50\%$ of the baseline, and additional middle cerebral artery (MCA) occlusion further decreased CBF to $\sim 20\%$. CCAo, CCA occlusion; MCAo, MCA occlusion; MCAr, MCA release.

area–(ipsilateral hemisphere–measured infarct area)/contralateral hemisphere area) $\times 100\%$. After brain extraction, animals observed to have experienced a subarachnoid hemorrhage were excluded from the study.

Experimental Protocol

Experiments were composed of two parts. The first part was designed to prove the neuroprotective effect of DRIPost and explore the optimal method of DRIPost. The second part was designed to investigate the role of K_{ATP} channels in the neuroprotective effect of DRIPost *in vivo*.

Part 1: Neuroprotective Effect of Delayed Remote Ischemic Postconditioning Against Cerebral Ischemia–Reperfusion Injury

To explore the optimal parameters required to observe the protective effects of DRIPost, our pilot study tested 56 rats

with 7 different serials of DRIPost performed at 3 or 6 hours after reperfusion ($n = 8$ for each). The experimental protocols are shown in Figure 1A. Control (Con) group rats were subjected to 90 minutes of ischemia followed by 72 hours of reperfusion. For the DRIPost group, the bilateral femoral arteries were separated below the bilateral groin ligament for later induction of femoral artery occlusion after 90 minutes of ischemia. The occlusion duration varied from 15 seconds, 5 minutes to 8 minutes, and bilateral femoral arteries were released for the same duration and repeated for three cycles.

Part 2: Effect of K_{ATP} Channel Blocking/Opening on Delayed Remote Ischemic Postconditioning-Induced Neuroprotection

In part 2, we further studied the role of K_{ATP} channels in the neuroprotective effect of DRIPost. Sixty-four adult male Sprague-Dawley rats were randomly divided into eight

Table 1 Physiological parameters, mean \pm s.d. ($N = 6$)

Time point	Temperature ($^{\circ}C$)	Pao_2 (mm Hg)	$Paco_2$ (mm Hg)	Glucose (mmol/L)	Arterial pH
Baseline	37.1 \pm 0.2	99.8 \pm 7.2	35.6 \pm 4.5	5.0 \pm 0.2	7.38 \pm 0.03
Ischemia, 45 minutes	37.8 \pm 0.1	95.3 \pm 6.5	38.5 \pm 5.4	4.8 \pm 0.3	7.32 \pm 0.02
Reperfusion, 30 minutes	37.0 \pm 0.3	90.2 \pm 5.4	38.6 \pm 4.2	4.5 \pm 0.3	7.35 \pm 0.12
DRIPost, 10 minutes	37.2 \pm 0.2	89.3 \pm 6.2	39.8 \pm 5.7	4.2 \pm 0.4	7.32 \pm 0.21

DRIPost, delayed remote ischemic postconditioning; Pao_2 , arterial oxygen tension; $Paco_2$, arterial carbon dioxide tension. Values are expressed as mean \pm s.d.

groups. The experimental protocols are shown in Figure 1B. Delayed remote ischemic postconditioning (5 minutes ischemia/5 minutes reperfusion in bilateral femoral arteries, three cycles) was induced at 6 hours after reperfusion in the postconditioning groups. This protocol showed a superior outcome in part 1 of the study. To block K_{ATP} channels, 20 mg/kg of 5-HD, which was dissolved in normal saline to a concentration of 12 mg/mL, was injected through the caudal vein 30 minutes before DRIPost. To open K_{ATP} channels, 5 mg/kg of DIAZ, which was dissolved in dimethyl sulfoxide (DMSO) to a concentration of 3 mg/mL, was administered through the caudal vein 20 minutes before DRIPost. To exclude the effect of administered drugs on cerebral ischemia–reperfusion injury, two separate control groups were used, which were administered the same dosage of drugs without postconditioning. Vehicle groups (saline group and DMSO group) were also included to preclude any influence.

Terminal Deoxynucleotidyl Transferase Nick-End Labeling and Quantification of Apoptosis

Apoptosis was quantified using a commercially available fluorescent terminal deoxynucleotidyl transferase nick-end labeling (TUNEL) kit, in accordance with the manufacturer protocol (Roche Diagnostics Corporation, Indianapolis, IN, USA). The sections were mounted with 50% (v/v) glycerol for examination under a fluorescence microscope. The total number of TUNEL-positive neurons in the right hemisphere was counted in three different fields for each section in a blind manner by light microscopy at $\times 400$ magnification (BX51; Olympus, Tokyo, Japan), and data from five animals at each stage were averaged.

Statistical Analysis

All data, excepting NDSs, are expressed as the mean \pm s.d. Physiologic parameters were analyzed by repeated-measures analysis of variance. The NDSs were expressed as median (range). The NDSs among different groups were compared by Kruskal–Wallis test. When Kruskal–Wallis test showed significant difference, the Mann–Whitney U -tests with Bonferroni correction were applied. The infarct volumes and TUNEL-positive neurons were compared among groups by one factor analysis of variance. $P < 0.05$ was considered statistically significant.

Results

Physiologic Parameters and Regional Cerebral Blood Flow

No statistical significance was noted among different time points for any of the physiologic parameters including cranial temperature, blood gas, and glucose concentrations (Table 1). Physiologic parameters remained in the normal range during the experimental period. Monitoring of regional cerebral blood flow ensured successful MCA occlusion (Figures 1C and 1D).

Neurologic Deficit Scores

All rats survived until 72 hours after reperfusion. At 24, 48, and 72 hours after reperfusion, the NDS in the 3 h-5 min, 3 h-8 min, and 6 h-5 min groups was significantly higher than that in the Con group ($P < 0.05$). Furthermore, the scores of animals in all groups were significantly lower than that of the 6 h-5 min group at each time point, which highlighted the neuroprotective effect of the 6 h-5 min DRIPost protocol in experiment part 1 (Figures 2A–2C).

In part 2 of the experiment, at 24, 48, and 72 hours after reperfusion, the NDS in the DIAZ, DRIPost, DRIPost+DIAZ, and DRIPost+5-HD groups was significantly higher than that of the Con group ($P < 0.05$). There was no statistical difference in NDS among the DRIPost and DRIPost+DIAZ groups at 24 and 48 hours after reperfusion ($P = 0.328$ and 0.382 , respectively). In contrast, the NDS in the DRIPost+5-HD group was significantly lower than that of the DRIPost group at the three time points ($P < 0.05$; Figures 3A–3C).

Infarct Volume

In experiment part 1, the infarct volume results are shown in Figure 2. All DRIPost protocols, except 3 h-15 s (60.48 \pm 3.08%), significantly decreased infarct volumes at 72 hours after reperfusion compared with that of the Con group (63.88 \pm 3.66%, $P < 0.05$). Among the protocols implemented in this study, the neuroprotective effect of three cycles of 5 minutes occlusion/reperfusion at 6 hours after reperfusion was superior (34.64 \pm 4.00%, $P < 0.05$; Figures 2D and 2E).

In experiment part 2, the infarct volume results are shown in Figure 3. Infarct volume in the DRIPost

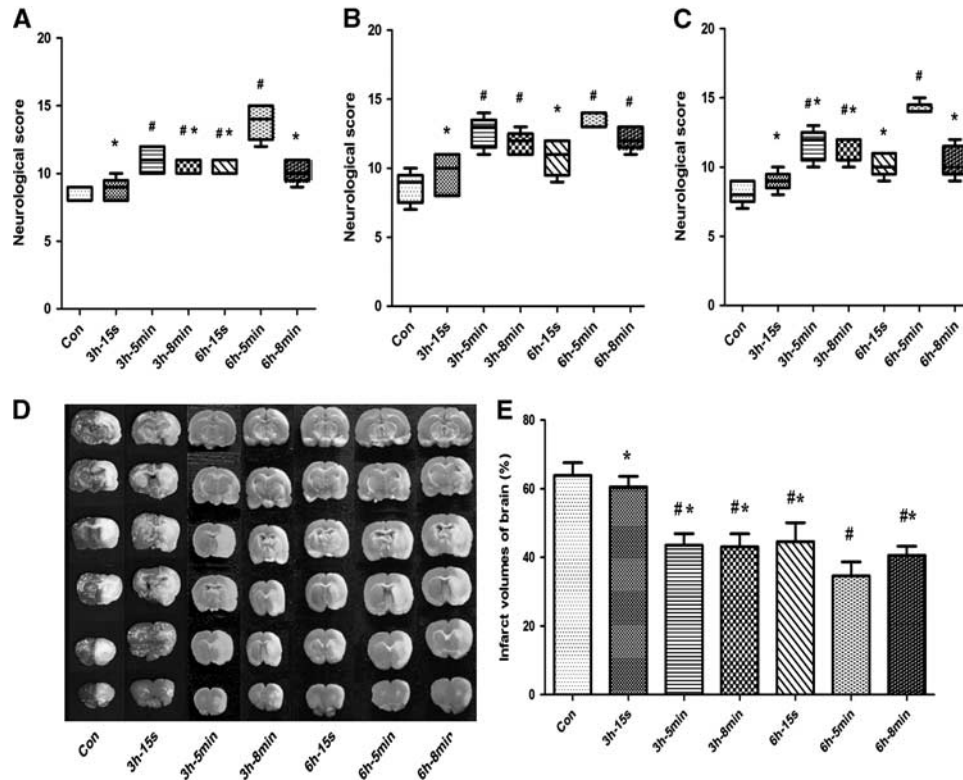


Figure 2 Neurologic scores and infarct volume after 90 minutes of transient middle cerebral artery occlusion in control (CON) and delayed remote limb ischemic postconditioning (DRIPost) groups in part 1 ($n = 8$ for each). DRIPost was performed with bilateral femoral artery occlusion/reperfusion at 3 or 6 hours after reperfusion. Neurologic scores are presented as the median (range); Data for infarct volumes are expressed as the mean \pm s.d. Neurologic scores were evaluated at 24 hours (A), 48 hours (B), and 72 hours (C) after reperfusion using the Garcia scoring system. (D) Representative 2,3,5-triphenyltetrazolium chloride staining of the cerebral infarct in the rat brain at 72 hours after reperfusion. (E) Statistical analysis of the percentage of infarct volume was determined for each study group. 3 h/6 h-15 s, 5 min and 8 min groups: DRIPost performed at 3 or 6 hours after reperfusion. The occlusion duration varied from 15 seconds, 5 minutes to 8 minutes, and bilateral femoral arteries were released for the same duration and repeated for three cycles. # $P < 0.05$ versus Con group. * $P < 0.05$ versus 6 h-5 min group.

group ($33.45 \pm 2.78\%$) was significantly smaller than that of the Con group ($65.58 \pm 3.30\%$) at 72 hours after reperfusion ($P = 0.000$). Administration of DIAZ (DIAZ group and DRIPost + DIAZ group) resulted in a reduction in infarct volume ($39.83 \pm 5.04\%$ and $34.36 \pm 2.56\%$, respectively) compared with the Con group ($P = 0.000$). Administration of 5-HD (DRIPost + 5-HD) eliminated the neuroprotective effect of DRIPost ($46.44 \pm 2.65\%$ versus $33.45 \pm 2.78\%$, $P = 0.000$). However, there was no significant difference in infarct volume between the DRIPost and DRIPost + DIAZ groups ($33.45 \pm 2.78\%$ versus $34.36 \pm 2.56\%$, $P = 0.602$; Figures 3D and 3E).

Terminal Deoxynucleotidyl Transferase Nick-End Labeling

The number of TUNEL-positive neurons in the CA₁ region significantly increased at 72 hours after ischemia-reperfusion injury in the Con group (Figure 4Aa). At the same observation time, the number of TUNEL-positive neurons in the CA₁ region of the

DRIPost group was significantly lower than that in the Con group ($P < 0.05$; Figure 4Ad). Injection of 5-HD, a K_{ATP} blocker, before DRIPost, increased the number of TUNEL-positive neurons in the CA₁ region after reperfusion (Figure 4Ae). However, there was no significant difference in the number of apoptotic neurons between the DRIPost and DRIPost + DIAZ groups 72 hours after reperfusion (Figure 4Ad and f).

Discussion

This study showed that RIPost with three cycles of 5 minutes limb ischemia/5 minutes reperfusion, which was conducted even 6 hours after focal cerebral ischemia-reperfusion injury, could reduce infarct size and improve neurologic deficits. Ischemia-reperfusion-induced neuronal apoptosis was also markedly attenuated by DRIPost. Activation of K_{ATP} channels is critical for endothelial protection by DRIPost because the conditioning stimuli were ineffective when applied in the presence of the nonselective K_{ATP} channel blocker 5-HD. Taken together, the

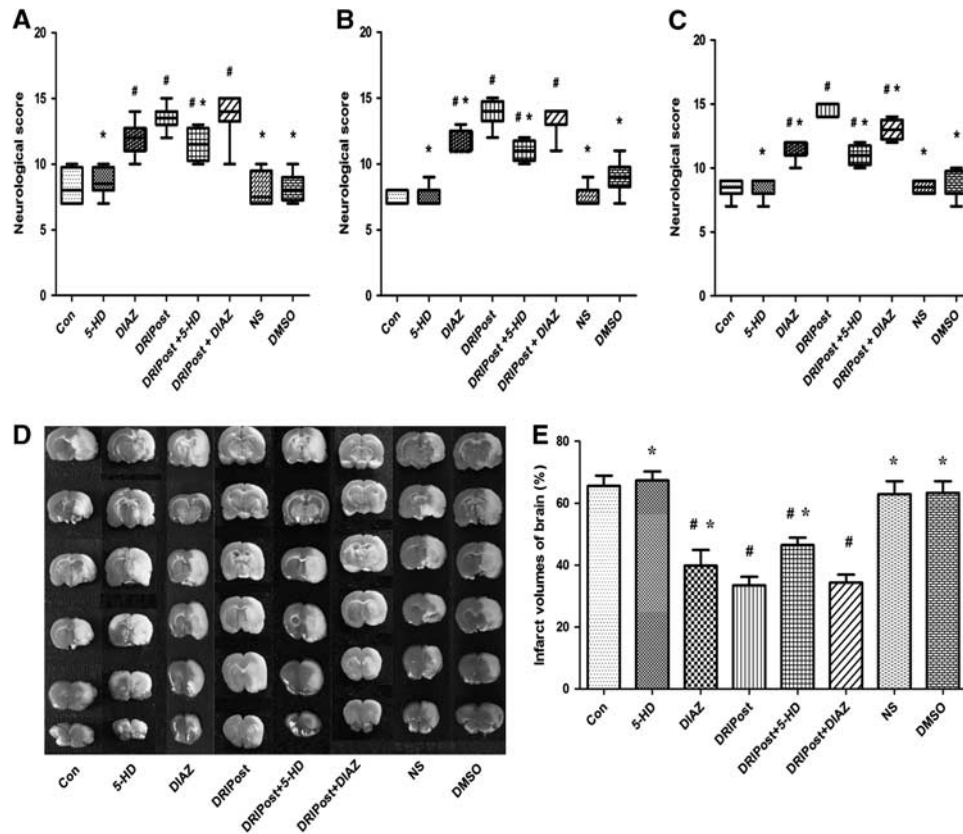


Figure 3 Neurologic scores and infarct volume after 90 minutes of transient middle cerebral artery occlusion in the control (CON), delayed remote limb ischemic postconditioning (DRIPost), 5-HD (K_{ATP} blocker, 20 mg/kg, intravenously), DIAZ (K_{ATP} opening, 5 mg/kg, intravenously), 0.9% (w/v) saline, dimethyl sulfoxide (DMSO) groups in part 2 ($n = 8$ for each). DRIPost was performed with bilateral femoral artery occlusion/reperfusion at 6 hours after reperfusion. Neurologic scores are presented as the median (range); Data for infarct volumes are expressed as the mean \pm s.d. Neurologic scores were evaluated at 24 hours (A), 48 hours (B), and 72 hours (C) after reperfusion using the Garcia scoring system. (D) Representative 2,3,5-triphenyltetrazolium chloride staining of the cerebral infarct in the rat brain at 72 hours after reperfusion. (E) Statistical analysis of the percentage of infarct volume was determined for each study group. # $P < 0.05$ versus Con group. * $P < 0.05$ versus DRIPost group. DIAZ, diazoxide; 5-HD, 5-hydroxydecanote; NS, normal saline.

present study provides evidence that the neuroprotective effect of DRIPost is associated with the activation of K_{ATP} channel opening.

We found that the optimal procedure for DRIPost, performed a few hours later, differed from that of rapid RIPost. Currently, there are at least two types of IPost, conventional IPost and RIPost. The conventional IPost refers to a series of brief, repetitive mechanical occlusions/reperfusion at the onset of reperfusion after long-term ischemia (Zhao *et al*, 2003). As for brain injury, conventional IPost was first reported by Zhao *et al* (2006) in a rat cerebral ischemia model. This protective concept of IPost has been confirmed by a number of groups using *in-vivo* global and focal ischemia models (Gao *et al*, 2008; Pignataro *et al*, 2008b) and *in-vitro* ischemic models (Pignataro *et al*, 2008b). Despite evidence from numerous animal studies, clinical application of *in-situ* ischemia for postconditioning of the brain is quite an unacceptable concept and impracticable. Recently, RIPost was primarily verified in studies of

myocardial injury. Both limb ischemic (Gritsopoulos *et al*, 2009; Tang *et al*, 2010) and renal IPost showed potential cardiac protective effects (Liu *et al*, 2007). A recent study reported that RIPost reduced cerebral infarction by 67% (Ren *et al*, 2009). In addition, DRIPost initiated as late as 3 hours, but not 6 hours, still robustly reduced brain infarction by 43%. In our study, we found that three cycles of postischemic reperfusion, either 5 minutes occlusion/5 minutes reperfusion or 8 minutes occlusion/8 minutes reperfusion for each cycle, were effective for inducing neuroprotection either applied at 3 hours or 6 hours after reperfusion. Among the effective procedures, 6 h-5 min group showed greater potential in reducing tissue damage after reperfusion. The different results between our group and Zhao's report may mainly reside in the difference of animal model. Taken together, both results suggest that early reperfusion after brain ischemic injury may broadly widen the therapeutic window for DRIPost, thus providing a better chance of neural salvage.

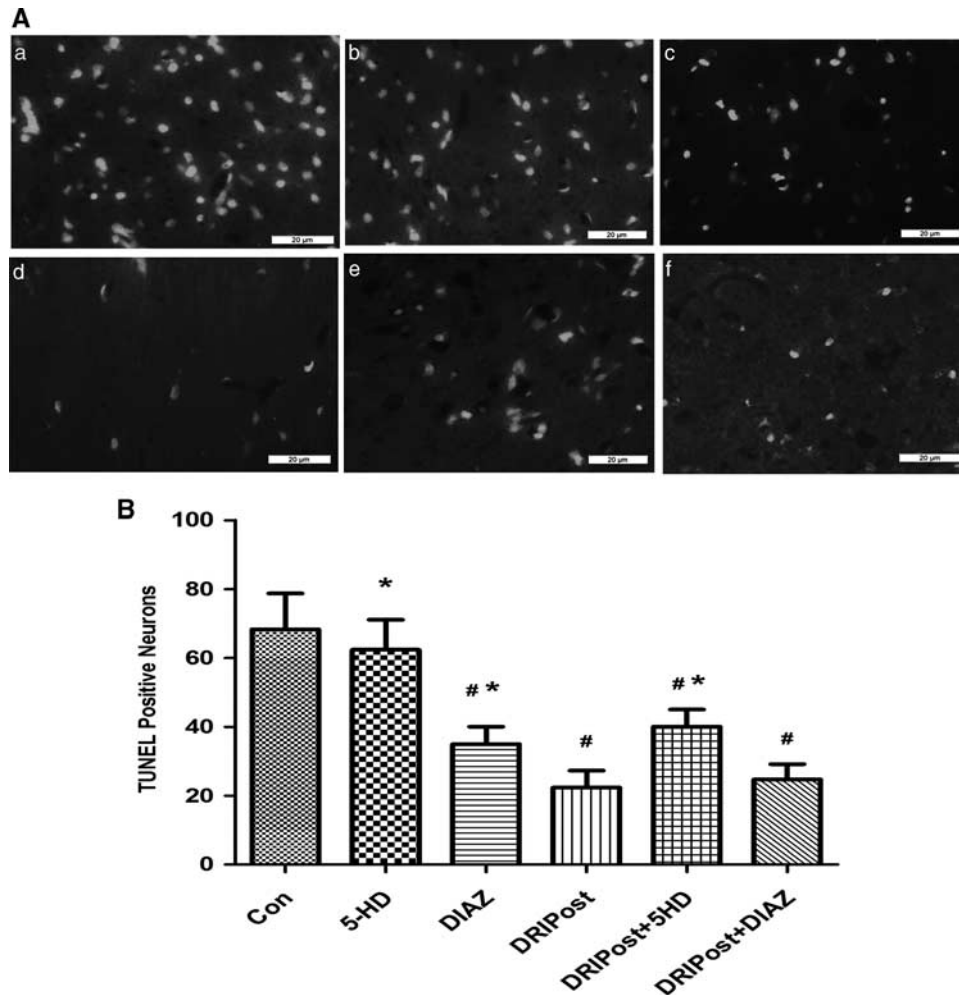


Figure 4 (A) Representative sections of nuclear DNA fragmentation staining performed by terminal deoxynucleotidyl transferase-mediated nick-end labeling (TUNEL) in the CA₁ region of the hippocampus from rats that randomly received 5-HD (20 mg/kg, intravenously) or DIAZ (5 mg/kg, intravenously) after 90 minutes of ischemia and 72 hours of reperfusion (Con). In the presence and absence of 5-HD or DIAZ pretreatment, or with a combination of DRIPost (three cycles of 5 minutes occlusion/reperfusion at 6 hours after reperfusion) and 5-HD or DIAZ. (a) Control (Con); (b) 5-HD; (c) DIAZ; (d) DRIPost; (e) DRIPost + 5-HD; (f) DRIPost + DIAZ. (B) Quantitative analysis of the number of TUNEL-positive neurons in the CA₁ region of the hippocampus. Data are presented as the mean ± s.d. Scale bar, 20 μm. **P* < 0.05 versus Con group. ***P* < 0.05, versus DRIPost group. DIAZ, diazoxide; 5-HD, 5-hydroxydecanote.

The underlying protective mechanisms of DRIPost are unknown. Nevertheless, our previous research from remote ischemic preconditioning (RIPC) against cerebral and spinal cord ischemia may shed light on our understanding of the protective mechanisms of DRIPost (Dong *et al*, 2010; Su *et al*, 2009). Emerging evidence from animal studies suggests that RIPC, IPost, and ischemic preconditioning share common signaling pathways, including triggers (adenosine receptor stimulation) (Kerendi *et al*, 2005; Steensrud *et al*, 2010), mediators (protein kinase C activation), and end effectors (opening of mitochondrial K_{ATP} channels, activation of prosurvival kinases, and inhibition of mitochondrial permeability transition pore opening; Hausenloy and Yellon, 2009; Meier *et al*, 2005; Yang *et al*, 2004). A popular theory has

proposed that a substance or humoral factor, such as oxygen free radicals, is carried in the blood stream from the preconditioning organ or tissue to the organ where it manifests its protective effect. After humoral factors arrive to the remote organs, the end effectors (such as K_{ATP} channels) appear to be activated and have an important role in the induction of ischemic tolerance by RIPC. The protective effect of K_{ATP} has been showed in several tissues, including the intestines, kidneys, liver, and brain (Hai *et al*, 2005; Tawfik *et al*, 2009). Opening of K_{ATP} channels has been shown to be a prerequisite event for the induction of protection against ischemia–reperfusion injury by RIPC, IPost, and RIPost (Schmidt *et al*, 2007; Steensrud *et al*, 2010; Yang *et al*, 2004). Zhang *et al* (2010) used an *in-vitro* model of PC12 cells

undergoing oxygen glucose deprivation to reveal that activation of K_{ATP} channels elicits protective effects against oxygen glucose deprivation-induced cell apoptosis by caspase-dependent and -independent mitochondrial pathways. In addition, Loukogeorgakis *et al* (2007) revealed that transient limb ischemia induces RPost in humans by a K_{ATP} channel-dependent mechanism. Thus, we speculate that K_{ATP} may also be involved in the neuroprotective effect of DRIPost. In our study, DIAZ, an activator of K_{ATP} channels, significantly reduced morphological injury, TUNEL-positive cells, and neurologic deficiency after cerebral ischemia–reperfusion injury. However, the K_{ATP} blocker 5-HD reversed the neuroprotective effect of DRIPost. These results indicate that the activation of K_{ATP} is involved in the neuroprotective effect of DRIPost.

Several limitations of this study merit comments. First, only six different protocols were designed in this study. The results showed that DRIPost is still effective when applied at 6 hours after reperfusion. Whether the same results can be drawn at even later phases after injury or protocols where other parameters are altered (i.e., changes in the frequency of I/R cycle, application time or duration of occlusion and reperfusion of limb postconditioning) show more potent neuroprotection deserves further elucidation. Our results only exhibited a partial reversal of neuroprotection induced by DRIPost after 5-HD injection, suggesting that opening of K_{ATP} channels is not the only cause for the protective effect. In addition, differences could be due to different animal models (heart or brain ischemia–reperfusion injuries) used. Both timing and length of IPost have been confirmed to be important in determining the magnitude of its protective effect. Ren *et al* (2008) also found that the protective effect of limb RIPost depends on the number and duration of the limb ischemic stimulus. Previous studies have shown that endogenous activation of adenosine receptors, especially the A_{2A} and A_{3A} subtypes, is involved in IPost- and RPost-mediated cardiac protection (Kerendi *et al*, 2005; Tsubota *et al*, 2010). Wang *et al* (2011) revealed that RPost performed in one limb alleviated reperfusion injury after focal cerebral ischemia through reactive oxygen species-mediated inhibition of endogenous δ protein kinase C activation signaling cascade in an *in-vivo* rat model of focal cerebral ischemia. Whether other mediators and effectors such as protein kinase C- ϵ and adenosine A_1 receptors that mediate the protective effect of IPost in myocytes could also participate in the induction of ischemic tolerance by DRIPost in the brain remains to be clarified.

In conclusion, DRIPost, even when applied at 6 hours after reperfusion, can induce a potent neuroprotective effect against focal cerebral ischemic reperfusion injury and inhibit apoptotic injury in the ischemic brain. The protective effects may partially be due to opening of K_{ATP} channels. Delayed remote ischemic postconditioning may be established as a potential practicable treatment for comparatively

late-hour ischemic stroke patients who receive medical care hours after the onset of stroke.

Disclosure/conflict of interest

The authors declare no conflict of interest.

References

- Burda J, Danielisova V, Nemethova M, Gottlieb M, Matiasova M, Domorakova I, Mechirova E, Ferikova M, Salinas M, Burda R (2006) Delayed postconditioning initiates additive mechanism necessary for survival of selectively vulnerable neurons after transient ischemia in rat brain. *Cell Mol Neurobiol* 26:1141–51
- Dong HL, Zhang Y, Su BX, Zhu ZH, Gu QH, Sang HF, Xiong L (2010) Limb remote ischemic preconditioning protects the spinal cord from ischemia-reperfusion injury: a newly identified nonneuronal but reactive oxygen species-dependent pathway. *Anesthesiology* 112:881–91
- Gao X, Zhang H, Takahashi T, Hsieh J, Liao J, Steinberg GK, Zhao H (2008) The Akt signaling pathway contributes to postconditioning's protection against stroke; the protection is associated with the MAPK and PKC pathways. *J Neurochem* 105:943–55
- Garcia JH, Liu KF, Ye ZR, Gutierrez JA (1997) Incomplete infarct and delayed neuronal death after transient middle cerebral artery occlusion in rats. *Stroke* 28: 2303–2309, 2310
- Gritsopoulos G, Iliodromitis EK, Zoga A, Farmakis D, Demerouti E, Papalois A, Paraskevaidis IA, Kremastinos DT (2009) Remote postconditioning is more potent than classic postconditioning in reducing the infarct size in anesthetized rabbits. *Cardiovasc Drugs Ther* 23:193–8
- Hai S, Takemura S, Minamiyama Y, Yamasaki K, Yamamoto S, Kodai S, Tanaka S, Hirohashi K, Suehiro S (2005) Mitochondrial K(ATP) channel opener prevents ischemia-reperfusion injury in rat liver. *Transplant Proc* 37:428–31
- Hausenloy DJ, Yellon DM (2009) Preconditioning and postconditioning: underlying mechanisms and clinical application. *Atherosclerosis* 204:334–41
- Kerendi F, Kin H, Halkos ME, Jiang R, Zatta AJ, Zhao ZQ, Guyton RA, Vinten-Johansen J (2005) Remote postconditioning. Brief renal ischemia and reperfusion applied before coronary artery reperfusion reduces myocardial infarct size via endogenous activation of adenosine receptors. *Basic Res Cardiol* 100:404–12
- Liu S, Wu XF, Zhang WZ, Sun YX, Cai SL (2007) [Remote postconditioning by brief renal ischemia and reperfusion reduces acute myocardial ischemia and reperfusion induced myocardial apoptosis in rabbits]. *Zhonghua Xin Xue Guan Bing Za Zhi* 35:757–60
- Lloyd-Jones D, Adams RJ, Brown TM, Carnethon M, Dai S, De Simone G, Ferguson TB, Ford E, Furie K, Gillespie C, Go A, Greenlund K, Haase N, Hailpern S, Ho PM, Howard V, Kissela B, Kittner S, Lackland D, Lisabeth L, Marelli A, McDermott MM, Meigs J, Mozaffarian D, Mussolino M, Nichol G, Roger VL, Rosamond W, Sacco R, Sorlie P, Roger VL, Thom T, Wasserthiel-Smoller S, Wong ND, Wylie-Rosett J (2010) Heart disease and stroke statistics—2010 update: a report from the American Heart Association. *Circulation* 121:e46–215

- Loukogeorgakis SP, Williams R, Panagiotidou AT, Kolvekar SK, Donald A, Cole TJ, Yellon DM, Deanfield JE, MacAllister RJ (2007) Transient limb ischemia induces remote preconditioning and remote postconditioning in humans by a $K(ATP)$ -channel dependent mechanism. *Circulation* 116:1386–95
- Meier JM, Urban P, Goy JJ (2005) Postconditioning inhibits mitochondrial permeability transition. *Future Cardiol* 1:457–60
- Pignataro G, Meller R, Inoue K, Ordonez AN, Ashley MD, Xiong Z, Gala R, Simon RP (2008a) *In vivo* and *in vitro* characterization of a novel neuroprotective strategy for stroke: ischemic postconditioning. *J Cereb Blood Flow Metab* 28:232–41
- Pignataro G, Meller R, Inoue K, Ordonez AN, Ashley MD, Xiong Z, Gala R, Simon RP (2008b) *In vivo* and *in vitro* characterization of a novel neuroprotective strategy for stroke: ischemic postconditioning. *J Cereb Blood Flow Metab* 28:232–41
- Ren C, Gao X, Niu G, Yan Z, Chen X, Zhao H (2008) Delayed postconditioning protects against focal ischemic brain injury in rats. *PLoS One* 3:e3851
- Ren C, Yan Z, Wei D, Gao X, Chen X, Zhao H (2009) Limb remote ischemic postconditioning protects against focal ischemia in rats. *Brain Res* 1288:88–94
- Schmidt MR, Smerup M, Konstantinov IE, Shimizu M, Li J, Cheung M, White PA, Kristiansen SB, Sorensen K, Dzavik V, Redington AN, Kharbanda RK (2007) Intermittent peripheral tissue ischemia during coronary ischemia reduces myocardial infarction through a $KATP$ -dependent mechanism: first demonstration of remote ischemic preconditioning. *Am J Physiol Heart Circ Physiol* 292:H1883–90
- Steensrud T, Li J, Dai X, Manlhiot C, Kharbanda RK, Tropak M, Redington A (2010) Pretreatment with the nitric oxide donor SNAP or nerve transection blocks humoral preconditioning by remote limb ischemia or intra-arterial adenosine. *Am J Physiol Heart Circ Physiol* 299:H1598–603
- Su B, Dong H, Ma R, Zhang X, Ding Q, Xiong L (2009) Cannabinoid 1 receptor mediation of spinal cord ischemic tolerance induced by limb remote ischemia preconditioning in rats. *J Thorac Cardiovasc Surg* 138:1409–1416
- Tang Y, Mennander A, Oksala N, Atalay M, Syvala H, Lagerstedt A, Kuuslahti M, Nieminen R, Anas N, Moilanen E, Tarkka M (2010) Postconditioning and remote postconditioning of ischemic rat cardiac grafts. *Eur Surg Res* 45:1–8
- Tawfik MK, Abo-Elmatty DM, Ahmed AA (2009) The role of ATP-sensitive potassium channel blockers in ischemia-reperfusion-induced renal injury versus their effects on cardiac ischemia reperfusion in rats. *Eur Rev Med Pharmacol Sci* 13:81–93
- Tsubota H, Marui A, Esaki J, Bir SC, Ikeda T, Sakata R (2010) Remote postconditioning may attenuate ischaemia-reperfusion injury in the murine hindlimb through adenosine receptor activation. *Eur J Vasc Endovasc Surg* 40:804–9
- Wang Q, Zhang X, Ding Q, Hu B, Xie Y, Li X, Yang Q, Xiong L (2011) Limb remote postconditioning alleviates cerebral reperfusion injury through reactive oxygen species-mediated inhibition of delta protein kinase C in rats. *Anesth Analg* 113:1180–7
- Xiong L, Zheng Y, Wu M, Hou L, Zhu Z, Zhang X, Lu Z (2003) Preconditioning with isoflurane produces dose-dependent neuroprotection via activation of adenosine triphosphate-regulated potassium channels after focal cerebral ischemia in rats. *Anesth Analg* 96:233–7
- Yang XM, Proctor JB, Cui L, Krieg T, Downey JM, Cohen MV (2004) Multiple, brief coronary occlusions during early reperfusion protect rabbit hearts by targeting cell signaling pathways. *J Am Coll Cardiol* 44:1103–10
- Zhang H, Zhao D, Wang Z, Zheng D (2010) Diazoxide preconditioning alleviates caspase-dependent and caspase-independent apoptosis induced by anoxia-reoxygenation of PC12 cells. *J Biochem* 148:413–21
- Zhao H, Sapolsky RM, Steinberg GK (2006) Interrupting reperfusion as a stroke therapy: ischemic postconditioning reduces infarct size after focal ischemia in rats. *J Cereb Blood Flow Metab* 26:1114–21
- Zhao ZQ, Corvera JS, Halkos ME, Kerendi F, Wang NP, Guyton RA, Vinten-Johansen J (2003) Inhibition of myocardial injury by ischemic postconditioning during reperfusion: comparison with ischemic preconditioning. *Am J Physiol Heart Circ Physiol* 285:H579–88



This work is licensed under the Creative Commons Attribution-NonCommercial-No Derivative Works 3.0 Unported License. To view a copy of this license, visit <http://creativecommons.org/licenses/by-nc-nd/3.0/>