

Three-Dimensional Quantitative Magnetic Resonance Imaging of Epiphyseal Cartilage Vascularity Using Vessel Image Features

New Insights into Juvenile Osteochondritis Dissecans

Jutta M. Ellermann, MD, PhD, Kai D. Ludwig, PhD, Mikko J. Nissi, PhD, Casey P. Johnson, PhD, John P. Strupp, MSEE, Luning Wang, PhD, Štefan Zbýň, PhD, Ferenc Tóth, DVM, PhD, Elizabeth Arendt, MD, Marc Tompkins, MD, Kevin Shea, MD, and Cathy S. Carlson, DVM, PhD

Investigation performed at the University of Minnesota, Minneapolis, Minnesota

Background: We introduce a quantitative measure of epiphyseal cartilage vascularity and examine vessel networks during human skeletal maturation. Understanding early morphological changes in the distal femoral condyle is expected to provide information on the pathogenesis of developmental diseases such as juvenile osteochondritis dissecans.

Methods: Twenty-two cadaveric knees from donors ranging from 1 month to 10 years of age were included in the study. Images of bone, cartilage, and vascularity were acquired simultaneously with a 3-dimensional gradient-recalled-echo magnetic resonance imaging (MRI) sequence. The secondary ossification center volume and total epiphysis cartilage volume ratio and articular-epiphyseal cartilage complex and epiphyseal cartilage widths were measured. Epiphyseal cartilage vascularity was visualized for 9 data sets with quantitative susceptibility mapping and vessel filtering, resulting in 3-dimensional data to inform vessel network segmentation and to calculate vascular density.

Results: Three distinct, non-anastomosing vascular networks (2 peripheral and 1 central) supply the distal femoral epiphyseal cartilage. The central network begins regression as early as 3 months and is absent by 4 years. From 1 month to 3 years, the ratio of central to peripheral vascular area density decreased from 1.0 to 0.5, and the ratio of central to peripheral vascular skeletal density decreased from 0.9 to 0.6. A narrow, peripheral vascular rim was present at 8 years but had disappeared by 10 years. The secondary ossification center progressively acquires the shape of the articular-epiphyseal cartilage complex by 8 years of age, and the central areas of the medial and lateral femoral condyles are the last to ossify.

Conclusions: Using cadaveric pediatric knees, we provide quantitative, 3-dimensional measures of epiphyseal cartilage vascular regression during skeletal development using vessel image features. Central areas with both early vascular regression and delayed ossification correspond to predilection sites of juvenile osteochondritis dissecans in this limited case series. Our findings highlight specific vascular vulnerabilities that may lead to improved understanding of the pathogenesis and better-informed clinical management decisions in developmental skeletal diseases.

Clinical Relevance: This paradigm shift in understanding of juvenile osteochondritis dissecans etiology and disease progression may critically impact future patient management. Our findings highlight specific vascular vulnerabilities during skeletal maturation in a group of active young patients seen primarily by orthopaedic surgeons and sports medicine professionals.

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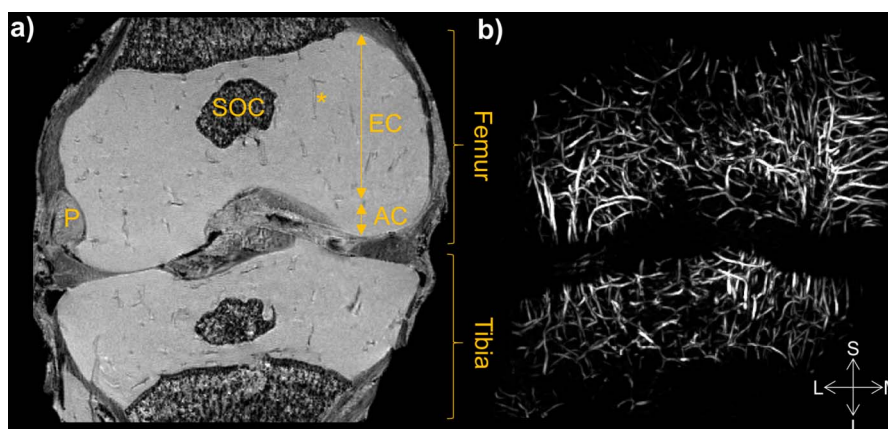


Fig. 1

Figs. 1-A and 1-B Anatomical and vascular MRI scans of a 1-month postnatal knee joint. **Fig. 1-A** A representative magnitude image from the 3-dimensional GRE sequence depicts the femur and tibia of a knee joint in the coronal orientation. Epiphyseal cartilage (EC) and articular cartilage (AC) are of high signal intensity and together encompass the articular-epiphyseal cartilage complex where low-intensity vascular canals (indicated by an asterisk) traverse the EC only. There is a small, low-intensity secondary ossification center (SOC). The popliteus tendon (P) marks the lateral aspect of the knee. **Fig. 1-B** The corresponding quantitative susceptibility mapping post-processed maximum intensity projection demonstrates the vascular network of the EC in both the femur and tibia. S = superior, L = lateral, M = medial, and I = inferior.

There is a growing clinical need to better understand pediatric joint diseases of developmental origin, such as juvenile osteochondritis dissecans¹⁻³. These diseases pose a serious burden on affected children, their families, and the health-care system. In the short term, they can negatively impact the self-esteem and social integration of otherwise healthy young patients, and later they can progress to premature osteoarthritis⁴. Unfortunately, research in this area has

been scarce, largely because of the lack of appropriate study methods.

Current clinical treatment of juvenile osteochondritis dissecans is hampered by a lack of clear, evidence-based management guidelines^{1,2}. Juvenile osteochondritis dissecans is diagnosed in its chronic stage with up to 75% of lesions involving the central aspect of the medial femoral condyle⁵⁻¹⁰. The pathogenesis remains poorly understood^{10,11}. A similar disease in animals, osteochondrosis, has

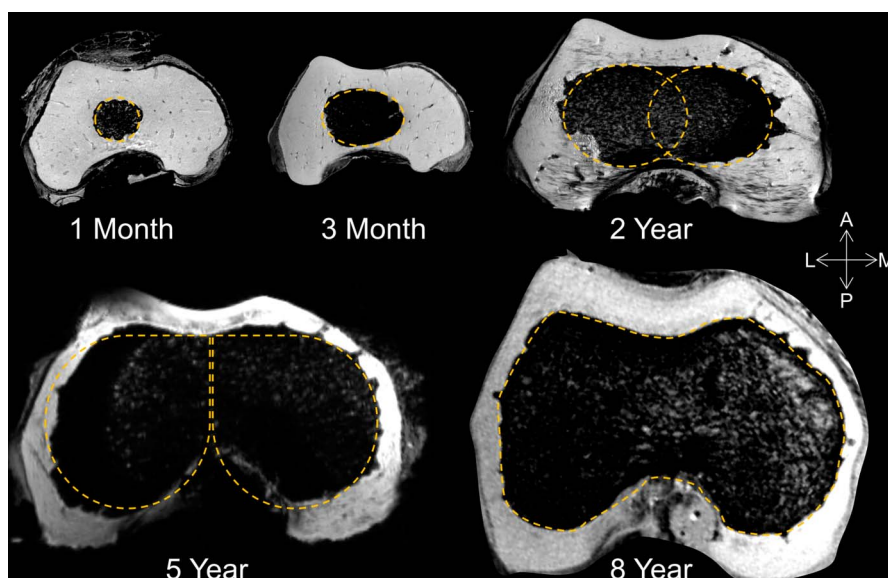


Fig. 2

Secondary ossification center development with age. Representative stages of the development of the secondary ossification center in the axial plane at the level of the maximal trochlear depth. The shape of the secondary ossification center changes from round (1 month), to oval (3 months), to congruent ovals (2 years), to aviator glasses (5 years), to finally assuming the asymmetric (about the midline) contour of the overlying cartilage template of the adult distal femur (8 years). A = anterior, L = lateral, M = medial, and P = posterior.

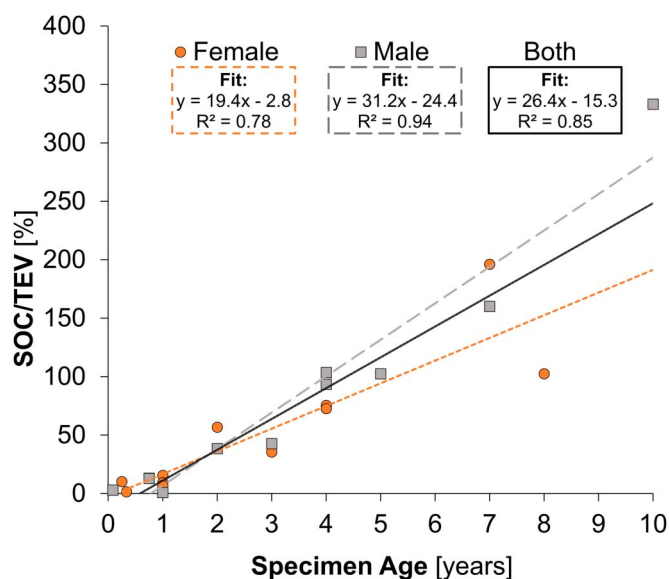


Fig. 3
The secondary ossification center (SOC) and total epiphysis volume (TEV) change with age. The percentage of volume ratio of the SOC (bone) to TEV (cartilage) increases linearly with age for both sexes.

been demonstrated to be caused by failure of the vascular supply to the epiphyseal cartilage¹²⁻¹⁷; work in cadaveric specimens has strongly suggested that the pathogenesis of the disease in humans is of similar origin¹⁸⁻²⁰. Recent clinical magnetic resonance imaging (MRI) studies provided evidence on epiphyseal cartilage origin and subsequent osseous manifestation²¹ of juvenile osteochondritis dissecans in pediatric patients and described the possible role of the secondary physis in the disease²².

Although the articular cartilage is avascular, the subarticular epiphyseal cartilage is highly vascular²³⁻²⁵. Growth and development during skeletal maturation lead to the gradual replacement of the vascular epiphyseal cartilage with bone via enchondral ossification²⁶. Traditionally, the vascular supply could only be studied using invasive techniques²⁷⁻²⁹. Recently, our group described noninvasive MRI approaches, including susceptibility-weighted imaging, to visualize epiphyseal cartilage vascularity³⁰⁻³², a method that has been already adapted to animal and human *in vivo* studies^{33,34}.

The purpose of this study was to introduce quantitative measures of the entire joint. Utilizing images of bone, cartilage, and vascular canals acquired simultaneously with a single 3-dimensional MRI sequence, we provide a comprehensive visualization of vessel network connectivity, quantification of epiphyseal vascular density, and progressive ossification during normal human skeletal maturation. These measurements constitute a necessary step toward improved clinical assessment tools for orthopaedic developmental diseases of children and young adults.

Materials and Methods

Subjects

Twenty-two cadaveric knee specimens from donors who were 1 month to 10 years of age, provided by AlloSource,

were included in this retrospective case series study. A waiver for institutional review board approval was applied. Eleven of the specimens were from female donors and 11 specimens were from male donors. For each sex, there were 7 unilateral specimens (right or left) and 2 bilateral specimens (same donor,

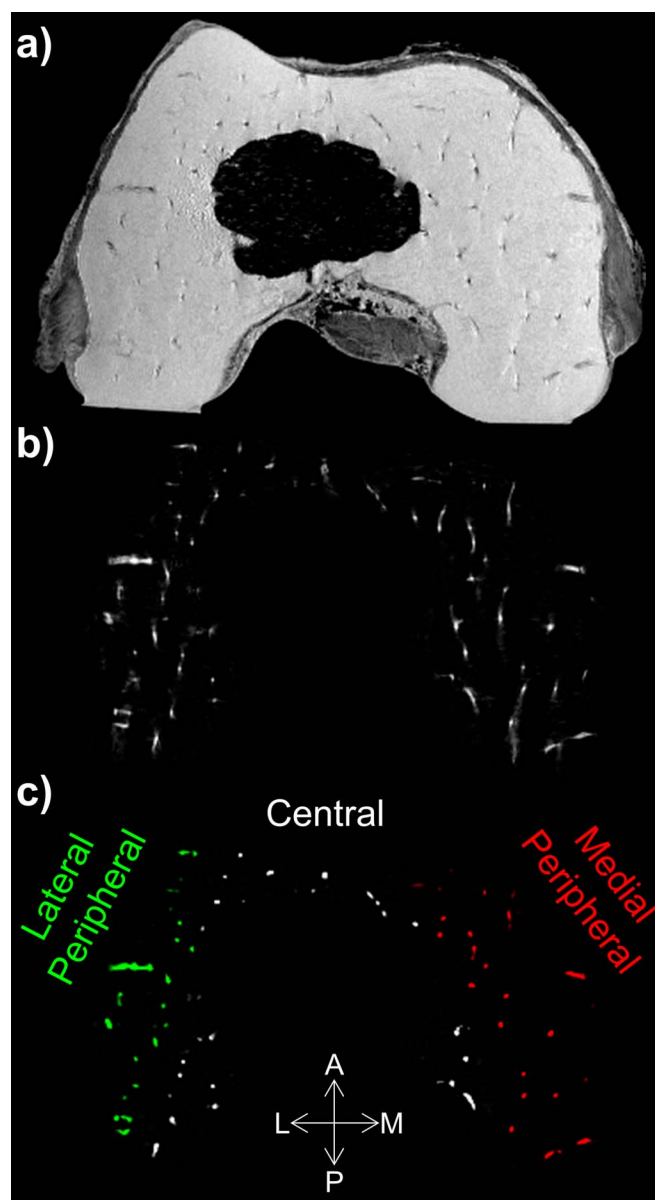


Fig. 4
Figs. 4-A, 4-B, and 4-C Epiphyseal cartilage vascular visualization with MRI. A = anterior, L = lateral, M = medial, and P = posterior. A representative slice of the 3-dimensional GRE MRI data set shows the magnitude anatomical image (**Fig. 4-A**) and vasculature (**Fig. 4-B**) after quantitative susceptibility mapping post-processing. Vessel-enhancement filters and semiautomatic segmentation allows for the vascular supply of the distal femoral epiphyseal cartilage to be visualized as a color-coded vascular network image (**Fig. 4-C**) (lateral peripheral = green vessels, central = white vessels, and medial peripheral = red vessels).

right and left). All knee specimens were presumed to be normal and without evidence of joint disease. Specimens were received frozen at -20°C and were thawed at room temperature immediately prior to MRI. For a description of the intra-epiphyseal vascular supply, we utilized a modification of a previously published nomenclature²⁷.

MRI

Images of bone, cartilage, and vascularity were acquired simultaneously with a 3-dimensional gradient-recalled-echo (GRE) MRI sequence on specimens between August 2013 and January 2016. The specimens were placed in test tubes immersed in perfluoropolyether to avoid image artifacts introduced by the susceptibility mismatch of tissue-air interfaces. The distal femoral specimens were oriented with the femoral diaphysis along the main magnetic field. Smaller specimens ($n = 7$) were imaged using a preclinical 9.4-T MRI system (31-cm bore; Agilent Technologies) and a quadrature volume transceiver coil (Millipede; Varian NMR Systems). The GRE sequence used a repetition time (TR)/echo time (TE) = 40/15 ms, flip angle = 15° , matrix = $384 \times 384 \times 384$, resolution = $\sim 100 \mu\text{m}$ isotropic, and acquisition bandwidth = 43 Hz/voxel. Larger specimens ($n = 15$) were imaged using a whole-body, 7.0-T MR system (MAGNETOM; Siemens Healthineers) with an 8-channel transmit/receive knee coil (Virtumed) driven by a B_1^+ shimming unit (CPC) with $8 \times 1 \text{ kW}$ amplifiers. At 7 T, the GRE sequence used a TR/TE = 45/29 ms, flip angle = 25° , matrix = up to $384 \times 384 \times 176$, resolution = $\sim 320 \mu\text{m}$ isotropic, and acquisition bandwidth = 60 Hz/voxel. Transmit B_1^+ shimming was performed to maximize excitation homogeneity using a fast estimation technique from B_1^+ maps as previously described^{30,35}.

Data Analysis

Three-dimensional visualization of the vascular structures in the epiphyseal cartilage was achieved using quantitative susceptibility mapping post-processing^{30,35}. Briefly, the GRE phase images were preprocessed to remove wrapping artifacts and background field contributions utilizing masks comprising the articular-epiphyseal cartilage complex³⁶⁻³⁹. A regularized quantitative susceptibility mapping method⁴⁰ was used to estimate the susceptibility distribution in the articular-epiphyseal cartilage complex using MATLAB (MathWorks)³⁰, which was then loaded into OsiriX (Pixmeo) for visualization of the intra-epiphyseal vascular supply. A board-certified musculoskeletal radiologist with >10 years of experience performed a qualitative evaluation of changes in the vascular supply of the distal femoral condyle and the morphological growth of the joint.

To pilot our vessel-filtering post-processing algorithm for 3-dimensional MRI data⁴¹, the vascular network of the epiphyseal cartilage was analyzed in a subset of 9 specimens, void of obvious degradation, at ages representing developmental milestones. A Hessian-based Frangi filter⁴² converted quantitative susceptibility mapping images to vessel-likelihood maps that were subsequently converted to binary vessel masks using a thresholding operation. The binary vessel masks were skeletonized using parallel medial axis thinning⁴³, and a network graph was calcu-

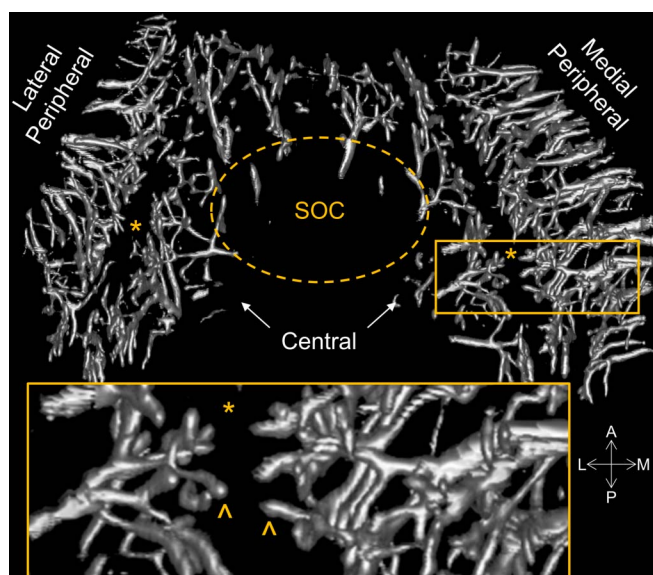


Fig. 5

Distal femoral epiphysis vascular canals at 3-month postnatal development. A volume rendering demonstrates the 2 distinct segmental distributions of the vascular canals with an intervening avascular zone (indicated by the asterisks) surrounding the small secondary ossification center (SOC). The peripheral vascular network consists of the lateral and medial vascular canals, and the central vascular network (arrows) consists of the intercondylar vessels. The inset of a higher-magnification image shows a group of regularly spaced vessels that extend from the periphery to the avascular zone (indicated by the asterisk) in a cluster of terminating small sinusoidal buds (carets). The avascular zone is interposed between the peripheral and central vascular networks; no anastomoses are observed between these 2 sources of blood supply. A = anterior, L = lateral, M = medial, and P = posterior.

lated⁴⁴. The 2 peripheral and single central vascular networks were segmented by manually selecting initial boundary seed points on the network graph for each region. Then an automatic region-growing algorithm enclosed the selected seeds and grew outwardly to the edges of the articular-epiphyseal cartilage complex mask, defining each vascular region simultaneously. The vascular area density was calculated using the binary vessel mask, and the vascular skeletal density (density measure independent of vessel thickness) was calculated using the skeletonized vessel mask⁴⁵. The vascular area density and vascular skeletal density for the peripheral network, both medial and lateral sides combined, were normalized to the central network to account for post-processing variations across specimens.

The secondary ossification center and articular-epiphyseal cartilage complex of the distal femoral epiphysis were manually segmented using OsiriX from 3-dimensional GRE images, and the ratio of the secondary ossification center volume to the total epiphysis volume, defined as the secondary ossification center volume plus the articular-epiphyseal cartilage complex volume, was calculated. The articular-epiphyseal cartilage complex and epiphyseal cartilage widths on the lateral and medial condyle were measured in the axial orientation for all specimens.

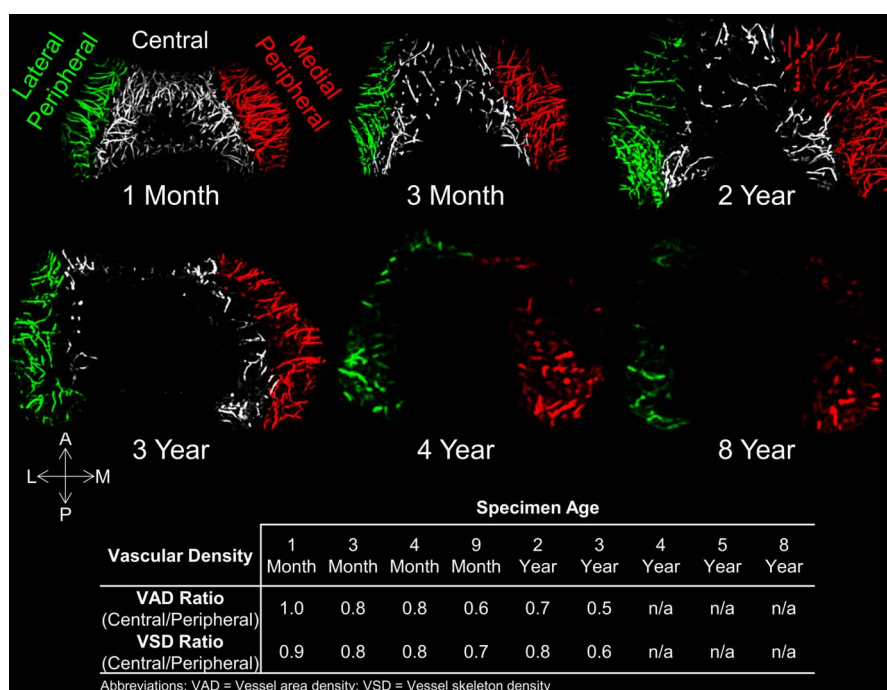


Fig. 6

Visualizations and density changes in vascular network with age. Maximum-intensity projections of the quantitative susceptibility mapping-post-processed, vessel-enhanced maps from 6 selected cadaveric specimens with increasing age are shown in the top 2 rows. The vasculature is color-coded according to the vascular network (lateral peripheral = green vessels, central = white, and medial peripheral = red). The embedded table summarizes the density ratio measurements (vascular area density and vascular skeletal density) of the central network to the peripheral (medial and lateral) network. No central network was observed in the specimens from donors who were ≥ 4 years of age. By 8 years of age, there was a small, residual peripheral vascular network. At 10 years of age, no epiphyseal cartilage vascularity could be detected. A = anterior, L = lateral, M = medial, P = posterior, and n/a = not available.

Statistics

A paired Student t test was performed between the medial and lateral epiphyseal cartilage widths, both of which were normalized to the articular-epiphyseal cartilage complex width, with significance set at $p < 0.05$.

Results

Advanced processing of 3-dimensional GRE images offers a high-resolution visualization of the vascular network within the epiphyseal cartilage of the knee. A representative GRE magnitude image from a cadaveric specimen from a 1-month-old donor is shown in Figure 1-A and demonstrates the anatomical locations of the articular cartilage, epiphyseal cartilage, and secondary ossification center within the femur of the right knee joint. The corresponding maximum intensity projection of quantitative susceptibility mapping images in Figure 1-B showed the vascular canals contained within both the femoral epiphyseal cartilage and the tibial epiphyseal cartilage, and no canals were observed in the articular cartilage.

The secondary ossification center volume increased and changed shape throughout development (Fig. 2). The contour of the secondary ossification center progressed from round (1 month), to oval (3 months), to congruent ovals (2 years), to the shape of aviator glasses (5 years). Between the ages of 2 and 5 years, the transverse (medial-lateral) dimension of the sec-

ondary ossification center appeared greater in width than its anterior-posterior dimension. The secondary ossification center changes were symmetric about the midline from 1 month up

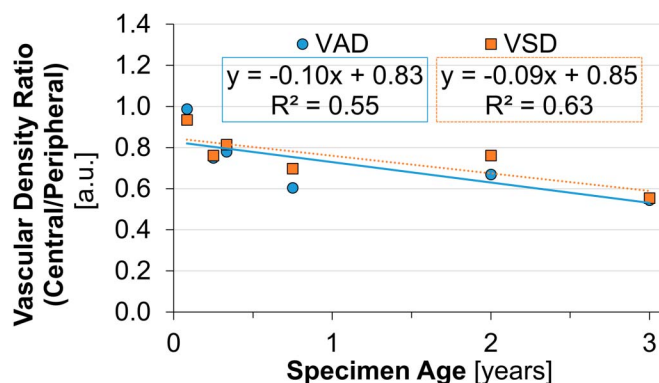


Fig. 7

Vascular density compared with specimen age. Quantitative vascular density measurements (displayed as a ratio of the central vascular network to the peripheral vascular network) for the selected 6 specimens containing a central vascular network (< 3 years of age). A linear fit showed a decreasing trend in the central and peripheral vascular area density (VAD) and vascular skeletal density (VSD) ratios with specimen age up to 3 years. a.u. = arbitrary units.

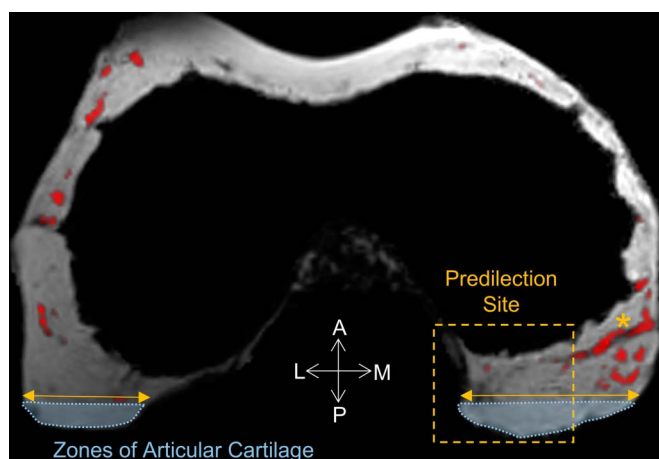


Fig. 8
MRI visualization of the vascular supply to the predilection site. A representative axial slice of the distal part of the femur from the 3-dimensional GRE MRI data set from a 5-year-old specimen shows the magnitude image (gray scale) overlaid with the vascular post-processed image (red scale). The central vascular supply has vanished. The ossification front has reached the central aspect of the intercondylar notch and grew wider in the transverse than the anteroposterior dimension leaving the thickest not-yet-ossified cartilage at the weight-bearing aspect of the joint. The articular cartilage (denoted by a light blue dotted line) is free of vessels; therefore, the subarticular epiphyseal cartilage vascular supply has to travel from the peripheral extra-articular perichondrium in parallel to the articular surface to reach the central aspect of the medial and lateral femoral condyles. This leaves these areas at the central aspect of the medial femoral condyle (wider), and to a lesser degree the lateral femoral condyle, most distant to the origin of their peripheral vascular supply. The predilection site of juvenile osteochondritis dissecans (highlighted) and a medial-lateral running vascular canal from the peripheral vascular supply is marked (denoted by an asterisk). A = anterior, L = lateral, M = medial, and P = posterior.

until approximately 5 years of age. During the early development, the secondary ossification center did not resemble the shape of the articular-epiphyseal cartilage complex template, and, by approximately 7 to 8 years of age, the secondary ossification center acquired the asymmetric shape of the articular-epiphyseal cartilage complex.

In contrast, the articular-epiphyseal cartilage complex assumed the adult shape at all stages of development, effectively providing a preformed cartilaginous template of the distal part of the femur as early as 1 month (Fig. 2). The articular-epiphyseal cartilage complex gradually increased in size with age (Appendix Fig. S1a) while qualitatively maintaining the asymmetric (lateral compared with medial) shape of the cartilaginous template of the condyles from 1 month to 10 years of age. The secondary ossification center in the center of the distal femoral epiphysis in the 1-month-old specimen occupied <3% of the epiphysis volume. Approximately 50% of the epiphysis volume was occupied by the secondary ossification center at 5 years of age. The ratio of the secondary ossification center to the total epiphysis volume increased approximately linearly

with age for both sexes (Fig. 3). The enchondral ossification in the epiphyseal cartilage appeared to occur at a faster pace in the male specimens than in the female specimens in our sample group. These ratios for the left and right distal femoral condyles of an individual subject were similar, as expected. The secondary ossification center volume and total epiphysis volume were measured for all specimens between the ages of 1 month and 10 years (Appendix Table S1).

A color-coded visualization of the epiphyseal cartilage vascular canals shows a highly ordered vascular network based on a segmental distribution (Fig. 4). A volume rendering for a 3-month-old specimen depicts the epiphyseal vascular canals penetrating the epiphyseal cartilage from the perichondrium and intercondylar notch in a radiating fashion, supplying the epiphyseal cartilage with a peripheral network, which consists of the lateral and medial vascular canals, and the central vascular network, which consists of the intercondylar vessels (Fig. 5). The vascular networks lack connecting anastomoses, leaving a narrow, linear watershed zone of avascular cartilage between vascular networks in both medial and lateral condyles. Each vascular canal terminated in a small sinusoidal bud that was visualized in detail using the 3-dimensional quantitative susceptibility mapping data (Fig. 5, inset).

The central network begins regression as early as 3 months and is absent by 4 years. Although the number of intra-epiphyseal vascular canals decreased with increasing age in all vascular networks, the relative decrease in number of vessels varied by location. Specifically, by 1 to 2 years of age, there was a substantial loss of central vessels in the intercondylar area of both the medial and lateral femoral condyles, as evidenced by a paucity of central epiphyseal vascular canals. The ratio of central to peripheral vascular area density decreased (from 1.0 to 0.5) and the central and peripheral vascular skeletal density ratio decreased (from 0.9 to 0.6) from 1 month to 3 years, showing a similar decreasing trend of vessel density across the analyzed specimens (Fig. 6). With the progression of the ossification front toward the intercondylar notch, the central vascular network had already markedly regressed by 3 years (Fig. 6) and was completely absent at approximately 4 years of age. A linear decrease in the central and peripheral vascular density ratios (vascular area density and vascular skeletal density ratios) was observed with increasing age (Fig. 7). The peripheral vascular network continued to regress with increasing age. The residual peripheral vascular network that was identified at 8 years of age had largely vanished at 10 years of age. The superficial weight-bearing articular cartilage was avascular at all observed time points (age, 1 month to 10 years).

During morphogenesis, the secondary ossification center first was observed at the central aspects of both femoral condyles, as expected. Subsequently, the secondary ossification center predominately grew to reach the extra-articular peripheral perichondrium. The weight-bearing aspects of both femoral condyles are last to be reached by the secondary ossification center as seen in the 5-year-old distal femoral specimen in Figure 8. The weight-bearing surface is covered with avascular articular

cartilage. The transverse epiphyseal cartilage width normalized to the articular-epiphyseal cartilage complex width was significantly greater ($p < 0.001$) in the medial compared with the lateral femoral condyle (Appendix Table S1), with a mean difference (and standard deviation) of $11\% \pm 6\%$. Epiphyseal cartilage width also increased with specimen age (Appendix Fig. S1b). This leaves areas at the central aspect of the wider medial femoral condyle and, to a lesser degree, the lateral femoral condyle most distant from the origin of their peripheral vascular supply.

Discussion

The most substantial finding from this study was that both early vascular regression and late enchondral ossification during skeletal maturation were demonstrated in areas that coincide with known predilection sites of juvenile osteochondritis dissecans. Our results demonstrate that the specific vascular architecture leaves the wider, last-to-ossify medial femoral condyle, where up to 75% of juvenile osteochondritis dissecans lesions occur⁵⁻⁹, most distant from the residual peripheral vascular supply.

Importantly, our observations on age-dependent milestones of vascular regression and juvenile osteochondritis dissecans were supported by a recent, very large retrospective demographic study with >1 million patients in this cohort¹⁰. No juvenile osteochondritis dissecans lesions of the knee were found in children who were 2 to 5 years of age, supporting the notion that the presence of both central and peripheral vascular beds up until 4 years of age as identified in our study might protect from clinically apparent lesions. The earliest manifestation at 6 years reported in this cohort coincides with our findings on regression of the peripheral vascularity between 4 and 10 years of age after the central vascular bed had already largely regressed at 4 years of age. The incidence of juvenile osteochondritis dissecans in individuals who were 6 to 19 years of age was low overall and was reported to be 9.5 per 100,000, with 6.8 cases per 100,000 for those who were between 6 and 11 years of age compared with 11.2 cases per 100,000 for those who were between 12 and 19 years of age¹⁰. It seems plausible that early lesions may present in the presence of residual peripheral vascularity between the ages of 6 and 11 years, but it takes progression to later stages between the ages of 12 and 19 years for them to become clinically symptomatic at a slightly higher rate. These findings are also consistent with a recent clinical MRI study²¹ that found an age-dependence depiction of epiphyseal cartilage lesion progression.

Shared architecture of vascularity and proposed similar pathogenesis of juvenile osteochondritis dissecans between pigs and humans were previously reported by our group¹⁸ based on 5 human specimens and 3 pig specimens. The current study provides a much larger sample size that enabled, for the first time, quantitative measures of the relative vascular density within the epiphyseal cartilage using a vessel-filtering algorithm, which has not previously been done. The new quantitative data add important new knowledge that is particularly relevant for accuracy in future patient studies. When putting our findings into the context of large

population cohorts, there is overwhelming evidence that, although pigs have a rampant prevalence of the disease⁴⁶, juvenile osteochondritis dissecans is rare in children and adolescents, despite evidence of shared vulnerabilities. That allows for promising conclusions that lesions might be clinically unimportant in humans, unless cofounding independent risk factors⁶ come into play.


Overuse in active children is an independent risk factor for developmental diseases such as juvenile osteochondritis dissecans^{22,47-49}. Approximately 60% to 75% of weight-bearing forces through the knee are normally transmitted through the medial compartment⁵⁰⁻⁵³. An association between the biomechanical loading including overweight and juvenile osteochondritis dissecans location exists^{9,54,55}. This implies that, once risk factors have been identified, preventive measures could be implemented. Furthermore, understanding the pathophysiology of juvenile osteochondritis dissecans as a delay in enchondral ossification due to the failure of vascular transfer from epiphyseal cartilage to the secondary ossification center with associated chondronecrosis in children²¹ would warrant new treatment approaches. A shift toward early detection and management is taking place in our clinical practice at the University of Minnesota, a referral center for juvenile osteochondritis dissecans; 2 to 3 years ago, the majority of patients referred for MRI were 13 to 20 years of age, and mostly osseous lesions were detected. Now, patients are younger, between 5 and 12 years of age, and increasingly cartilaginous lesions are identified (Tompkins M, University of Minnesota. Personal communication; 2019 Jun 4). Routinely, we utilize the clinical sequence described in this article to differentiate between cartilaginous and osseous lesions, information that is currently unattainable with clinical turbo-spin-echo sequences²¹. Epiphyseal cartilage vascular imaging in children has been shown to be feasible clinically at 3 T⁵⁶ and at 7 T³⁴; recently, the U.S. Food and Drug Administration (FDA) approved epiphyseal cartilage vascular imaging for clinical use in patients who weighed >30 kg. Alterations in epiphyseal cartilage vascularity are important in other diseases, such as infection, juvenile arthritis⁵⁷, Legg-Calvé-Perthes disease⁵⁸, Osgood-Schlatter disease, mucopolysaccharidosis⁵⁹, and failure of osteogenesis⁶⁰ during skeletal development.

This study had several major limitations. First, the number of specimens and the age and sex distribution depended on their availability. Unfortunately, these specimens were few in number and the ages and sex distribution were unpredictable. This limitation precluded a prospective study based on power calculations. Furthermore, the small sample size limited a statistical comparison between male and female specimens to identify sex differences. Vascular variations within an age range or between left and right knees were not evaluated because we had limited specimen numbers and this would require larger, population-based studies. Second, the conclusions drawn in this report were based on high-resolution ex vivo results. Additional in vivo studies are warranted to prove technical translatability and clinical utility. The clinical impact will need to be determined in future studies. Lastly, our data did not prove a direct cause-and-effect relationship between vascular regression and occurrence of juvenile osteochondritis

dissecans. Instead, the data provided evidence of complex, multifactorial relationships between local age-dependent vulnerabilities at predilection sites of juvenile osteochondritis dissecans, biomechanical loading, shear forces, and unknown genetic predisposition. Future studies are needed to shed more light into these complex interactions.

In conclusion, this study provides simultaneously acquired, quantitative, 3-dimensional MRI measures of the regressing epiphyseal cartilage vascularity during skeletal development using vessel image features, as well as bone and cartilage volumes. In pursuit of a paradigm shift toward precision medicine, the introduced quantitative approach can serve as a tool for necessary further clinical studies.

Appendix

 Supporting material provided by the authors is posted with the online version of this article as a data supplement at [jbjs.org \(http://links.lww.com/JBJSOA/A131\)](http://links.lww.com/JBJSOA/A131). ■

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Jutta M. Ellermann, MD, PhD¹
 Kai D. Ludwig, PhD¹
 Mikko J. Nissi, PhD^{1,2}
 Casey P. Johnson, PhD^{1,3}
 John P. Strupp, MSEE¹
 Luning Wang, PhD¹
 Štefan Zbýň, PhD¹
 Ferenc Tóth, DVM, PhD³

Elizabeth Arendt, MD¹
 Marc Tompkins, MD¹
 Kevin Shea, MD⁴
 Cathy S. Carlson, DVM, PhD³

¹Center for Magnetic Resonance Research (CMRR) (J.M.E., K.D.L., M.J.N., C.P.J., J.P.S., L.W., and S.Z.), Department of Radiology, and Department of Orthopaedic Surgery (E.A. and M.T.), Medical School, University of Minnesota, Minneapolis, Minnesota

²Department of Applied Physics, University of Eastern Finland, Kuopio, Finland

³Departments of Veterinary Population Medicine (F.T.) and Veterinary Clinical Sciences (C.P.J., C.S.C.), College of Veterinary Medicine, University of Minnesota, St. Paul, Minnesota

⁴Department of Orthopaedic Surgery, Stanford University, Stanford, California

Email address for J.M. Ellermann: eller001@umn.edu

ORCID iD for J.M. Ellermann: [0000-0002-7109-9656](https://orcid.org/0000-0002-7109-9656)

ORCID iD for K.D. Ludwig: [0000-0001-9867-9346](https://orcid.org/0000-0001-9867-9346)

ORCID iD for M.J. Nissi: [0000-0002-5678-0689](https://orcid.org/0000-0002-5678-0689)

ORCID iD for C.P. Johnson: [0000-0001-9826-7296](https://orcid.org/0000-0001-9826-7296)

ORCID iD for J.P. Strupp: [0000-0002-1728-3380](https://orcid.org/0000-0002-1728-3380)

ORCID iD for L. Wang: [0000-0003-0799-4469](https://orcid.org/0000-0003-0799-4469)

ORCID iD for Š. Zbýň: [0000-0002-8211-2828](https://orcid.org/0000-0002-8211-2828)

ORCID iD for F. Tóth: [0000-0002-6615-389X](https://orcid.org/0000-0002-6615-389X)

ORCID iD for E. Arendt: [0000-0002-6944-7332](https://orcid.org/0000-0002-6944-7332)

ORCID iD for M. Tompkins: [0000-0002-8641-3150](https://orcid.org/0000-0002-8641-3150)

ORCID iD for K. Shea: [0000-0002-4160-0163](https://orcid.org/0000-0002-4160-0163)

ORCID iD for C.S. Carlson: [0000-0002-0355-5525](https://orcid.org/0000-0002-0355-5525)

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