

Research Article

In Vitro Pharmacological Activities and GC-MS Analysis of Different Solvent Extracts of *Lantana camara* Leaves Collected from Tropical Region of Malaysia

Mallappa Kumara Swamy,¹ Uma Rani Sinniah,¹ and Mohd. Sayeed Akhtar²

¹Department of Crop Science, Faculty of Agriculture, Universiti Putra Malaysia (UPM), 43400 Serdang, Selangor, Malaysia

²Institute of Tropical Agriculture, Universiti Putra Malaysia (UPM), 43400 Serdang, Selangor, Malaysia

Correspondence should be addressed to Mallappa Kumara Swamy; swamy.bio@gmail.com
and Uma Rani Sinniah; umarani@upm.edu.my

Received 18 September 2015; Revised 19 November 2015; Accepted 22 November 2015

Academic Editor: Rahmatullah Qureshi

Copyright © 2015 Mallappa Kumara Swamy et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

We investigated the effect of different solvents (ethyl acetate, methanol, acetone, and chloroform) on the extraction of phytoconstituents from *Lantana camara* leaves and their antioxidant and antibacterial activities. Further, GC-MS analysis was carried out to identify the bioactive chemical constituents occurring in the active extract. The results revealed the presence of various phytocompounds in the extracts. The methanol solvent recovered higher extractable compounds (14.4% of yield) and contained the highest phenolic (92.8 mg GAE/g) and flavonoid (26.5 mg RE/g) content. DPPH radical scavenging assay showed the IC₅₀ value of 165, 200, 245, and 440 µg/mL for methanol, ethyl acetate, acetone, and chloroform extracts, respectively. The hydroxyl scavenging activity test showed the IC₅₀ value of 110, 240, 300, and 510 µg/mL for methanol, ethyl acetate, acetone, and chloroform extracts, respectively. Gram negative bacterial pathogens (*E. coli* and *K. pneumoniae*) were more susceptible to all extracts compared to Gram positive bacteria (*M. luteus*, *B. subtilis*, and *S. aureus*). Methanol extract had the highest inhibition activity against all the tested microbes. Moreover, methanolic extract of *L. camara* contained 32 bioactive components as revealed by GC-MS study. The identified major compounds included hexadecanoic acid (5.197%), phytol (4.528%), caryophyllene oxide (4.605%), and 9,12,15-octadecatrienoic acid, methyl ester, (Z,Z,Z)- (3.751%).

1. Introduction

Nature has existed as a source of almost all drugs for many years and natural products were the only source of medicine for mankind ever since the ancient period. Herb based products play an important role in primary human health care as the majority (80%) of the global population rely on traditional medical practices [1, 2]. Most of the modern drugs are derived either from plant sources or from their derivatives for various medicaments and are extensively used in the pharma industry [2]. In addition to the prevailing health problems, emerging infectious diseases and disorders have seriously caused the world population to suffer with a high mortality rate. It is reported that about 50% of all fatality occurring in tropic countries is mainly due to the current

infectious diseases [3]. Also, increase of antimicrobial resistance among the pathogens is a rising problem which is challenging the scientific advancement of the medical world [4]. This situation has prompted researchers to develop efficient new antimicrobial agents. Therefore, exploration of natural products as leads to discover new drug molecules is continuously made to understand their therapeutic potential with special reference to biological activities, efficiency, and safety aspects. Exploration of medicinal plants for curative purposes is mainly based on the available traditional information from the experts and local population [5, 6].

Lantana camara L. is a medicinal aromatic plant that belongs to the family Verbenaceae and occurs in most parts of the world as an evergreen notorious weed species. It is also considered as an ornamental garden plant. It is widely used

in different traditional medical practices for treating various health problems. Different parts of the plant are used in treating various human ailments such as measles, chicken pox, tetanus, malaria, cancers, asthma, ulcers, fevers, eczema, skin rashes, cardiac disorders, and rheumatism [7, 8]. Also leaf extracts and essential oil of *L. camara* leaves possess larvicidal activities, antioxidant, anti-inflammatory, analgesic, antidiabetic, hypolipidemic, anthelmintic, wound healing, and antipyretic properties [9, 10]. The therapeutic potential of the plant is due to the occurrence of many bioactive phytochemicals such as terpenoids, alkaloids, flavonoids, phenolics, glycosides, and steroids as major phytoconstituents [11]. Some of the important bioactive compounds include quercetin, isorhamnetin, oleanolic acid, lantadene A, β -sitosterol pomonic acid, camaric acid, verbacosides, lantanoside, linaroside, octadecanoic acid, palmitic acid, and docosanoic acid. The essential oil from the leaves is rich in monoterpenes and sesquiterpenes [11–13]. Various factors including genetic, geographical location, plant parts, and environmental factors have been shown to influence the accumulation of phytochemical contents in different parts of *L. camara* and its essential oil composition [14–16]. Also, occurrence of varietal differences in phytoconstituents of *L. camara* has been documented by Sharma et al. [17]. More recently, *in vitro* study indicated the existence of chemical differences in methanol leaf extract of four varieties of *L. camara* collected from India and their antioxidant property was found to differ [8]. Certainly, more research efforts should be carried out to explore the potential benefits of *L. camara* for treating various health problems. Therefore, the present investigation was aimed at determining the phytochemical constituents and antioxidant and antimicrobial activities of different solvent extracts of *L. camara* leaves collected from the tropical region of Malaysia. Moreover, the bioactive components of the extracts were also identified using GC-MS analysis.

2. Materials and Methods

2.1. Plant Collection. *L. camara* plant material was collected from the forest area near Universiti Putra Malaysia, Serdang, Selangor, Malaysia, during the month of May 2015. Plant material was authenticated by N. A. P. Abdullah, Department of Crop Science, Universiti Putra Malaysia, Malaysia, and the voucher specimen (LC-102015) was deposited at the department. The leaves were detached from the collected materials, washed with water and dried under shade for 1 week, and finely powdered using electric blender. The powdered material was kept at room temperature for future use.

2.2. Preparation of Extracts. Five grams of powdered leaves was kept in a beaker to which 100 mL of various organic solvents (ethyl acetate, methanol, acetone, and chloroform) was added and thoroughly shaken. Later the mixture was placed at room temperature for 48 hrs and stirred 2-3 times a day. After filtering the mixture, the filtrate was evaporated to dryness using Rotavapor. The final extracts were weighed to determine the yield (%) and the dried extracts were stored at 4°C in a refrigerator for further studies.

2.3. Phytochemical Screening. The presence of various phytochemical constituents such as alkaloids, saponins, flavonoids, phenolics, anthraquinones, tannins, cardiac glycosides, steroids, and terpenoids was screened qualitatively by using standard procedures [18–20].

2.4. Determination of Total Phenolic and Flavonoid Contents. Total phenolic contents present in different solvent extracts were determined by using FC (Folin-Ciocalteu) colorimetric method as detailed by Salar and Seasotiya [21]. About 0.1 g of dried extract was suspended in 1 mL of the respective solvents and 0.1 mL of this solution was mixed thoroughly with 1 mL of sodium carbonate (20%) solution and 0.5 mL of 50% FC reagent. Thereafter, the solution was allowed to remain at room temperature for about 20 min to observe the color change. The absorbance was taken against the blank (water) at 730 nm. Using different concentrations of gallic acid, a standard calibration curve was generated. Total phenolic content was represented as mg of gallic acid equivalents (GAE) per gram of dried extract.

Total flavonoids content present in different solvent extracts was verified by the modified procedure of Zhishen et al. [22] using colorimeter. Briefly, 2 mL of distilled water was mixed well with 0.5 mL of solvent extract. Thereafter, 150 μ L of NaNO₂ solution (5%) was added and kept for 5 min at room temperature. It was followed by adding 600 μ L of AlCl₃ (10%) and 2 mL of NaOH (4%). After mixing thoroughly, the solution was made up to 5 mL with distilled water and set aside for 15 min at room temperature. By using water as blank, absorbance was measured at 510 nm. A standard calibration curve was generated using different concentrations of rutin. Total flavonoid content was articulated as mg of rutin equivalent (RE) per gram of dried extract.

2.5. Antioxidant Activity

2.5.1. Free Radical Scavenging Activity. The antioxidant activity of each extract was assessed using DPPH (1,1-diphenyl-2-picrylhydrazyl) free radical scavenging assay as explained by Mohanty et al. [23] with little modifications. In short, 0.3 mL of different extract at varied concentrations (100–1000 μ g/mL) was mixed with 2 mL of 0.1 mM DPPH solution and incubated for 30 min under dark conditions at room temperature. Afterwards, using UV-visible spectrophotometer, the absorbance was taken at 517 nm against methanol (blank) while ascorbic acid served as a standard sample. The reaction mixture showing lower absorbance is an indication of higher activity of radical scavenging. The inhibition percentage of free radical scavenging was calculated based on the following formula:

$$\text{DPPH scavenging activity (\%)} = \frac{A_{\text{control}} - A_{\text{sample}}}{A_{\text{control}}} \times 100. \quad (1)$$

For each extract, IC₅₀ values (the minimum quantity of extract necessary for scavenging free radicals up to 50%) were calculated from the standard plot.

TABLE 1: Qualitative screening of phytochemicals present in different solvent extracts of *L. camara* leaves.

Phytochemicals	Ethyl acetate	Methanol	Chloroform	Acetone
Alkaloids	–	+	–	+
Flavonoids	+	+	+	+
Saponins	+	+	–	+
Phenolics	+	+	+	+
Tannins	–	–	+	–
Anthraquinones	+	–	–	+
Cardiac glycosides	+	+	+	+
Terpenoids	–	+	+	–
Steroids	–	+	–	–

Note: + = present, – = absent.

2.5.2. Hydrogen Peroxide (H_2O_2) Scavenging Activity. The potential capacity of different solvent extracts to scavenge H_2O_2 was carried out by the method outlined by Mohanty et al. [23]. In brief, various quantities of plant extract (100–1000 $\mu\text{g/mL}$) were prepared as detailed in previous experiment. By using phosphate buffer (pH 7.4), H_2O_2 solution (4 mM/L) was prepared. Varied amounts of plant extracts were mixed with 0.6 mL of H_2O_2 solution and kept for 30 min incubation at room temperature. The activity of H_2O_2 was analyzed by taking absorbance at 230 nm using phosphate buffer without H_2O_2 as blank solution. H_2O_2 scavenging activity (%) was calculated by using the following equation:

$$H_2O_2 \text{ scavenging activity (\%)} = \frac{A_{\text{control}} - A_{\text{sample}}}{A_{\text{control}}} \times 100. \quad (2)$$

For each extract, IC_{50} value (the minimum quantity of extract necessary for scavenging free radicals up to 50%) was calculated from the standard plot.

2.5.3. Determination of Antibacterial Activity. The antibacterial activities for the extracts were evaluated by using disc diffusion method. The experiment included both Gram positive (*Micrococcus luteus*, *Bacillus subtilis*, and *Staphylococcus aureus*) and Gram negative (*Escherichia coli* and *Klebsiella pneumoniae*) pathogenic strains of bacteria. One mg of each extract was dissolved in 1 mL of DMSO (dimethyl sulfoxide) and about 2.5, 5.0, and 10 μL of this solution were impregnated on sterilized filter paper discs (6 mm size). The discs were kept on the nutrient agar medium preinoculated uniformly with the known bacterial culture. The discs soaked with chloramphenicol (50 μL of 50 $\mu\text{g/mL}$) served as positive control while discs soaked with 50 μL DMSO served as negative control. All culture plates were kept in an incubator at 37°C for 24 h and the bacterial inhibition zone was recorded and expressed in mm. For each bacterial strain, the test was repeated 3 times.

2.6. GC-MS Analysis. GC-MS analysis of the active methanol extract of *L. camara* was carried out by using the GC-MS instrument (Model GCMS-QP2010 Ultra, Shimadzu Co., Japan) equipped with a capillary column DB-1 (0.25 μm film

$\times 0.25$ mm i.d. $\times 30$ m length). The instrument was operated in electron impact mode at ionization voltage (70 eV), injector temperature (230°C), and detector temperature (280°C). The carrier gas used was helium (99.9% purity) at a flow rate of 1 mL/min and about 1 μL of the sample was injected. The oven temperature was initially programmed at 80°C (isothermal for 5 min.) and then increased to 200°C at 5°C/min and finally to 280°C at 5°C/min (isothermal for 16 min). The identification of compounds from the spectral data was based on the available mass spectral records (NIST and WILEY libraries).

2.7. Statistical Analysis. All the data measured in each experiment included 3 replications ($n = 3$) and the results were represented as mean \pm SD. The one-way analysis of variance (ANOVA) was performed to compare the data and Tukey's test was used to find out the statistically significant differences at $p < 0.05$ using statistical software, GraphPad Prism version 5.0.

3. Results and Discussion

The results of preliminary investigation on the phytochemicals present in different solvent extracts of *L. camara* are presented in Table 1. Different phytochemicals such as alkaloids, saponins, flavonoids, phenolics, anthraquinones, tannins, cardiac glycosides, steroids, and terpenoids were detected in the crude extracts. Flavonoids, phenolics, and cardiac glycosides were noticed in all solvent extracts used. Methanol extracts with 7 phytoconstituents were the best among the organic solvents evaluated in our study. Acetone extract showed the presence of 6 phytoconstituents while ethyl acetate and chloroform extracts contained 5 phytochemicals. All these identified phytochemicals are known to have a wide range of biological activities including antibacterial, antifungal, antiviral, antioxidant, and cytotoxic properties [19]. Understanding the occurrence of phytochemicals in medicinal plants is advantageous and presently, the discovery of new drug compounds or lead molecules from plants is mainly based on the systematic examination of different plant extracts or plant based products. Also, this preliminary knowledge can decipher a new source for economically valued chemical compounds [1, 23].

TABLE 2: Dry weight and total yield of different solvent extracts of *L. camara* leaves.

Solvent extracts	Weight of the extract ($\mu\text{g} \pm \text{SD}$)	Yield (%)
Ethyl acetate	$501.3 \pm 3.5^{\text{b}}$	10.0
Methanol	$721.3 \pm 1.5^{\text{a}}$	14.4
Acetone	$260.6 \pm 4.0^{\text{c}}$	5.2
Chloroform	$141.6 \pm 2.5^{\text{d}}$	2.8

Note: each value is expressed as mean \pm standard deviation (SD) ($n = 3$). Values in the column followed by a different letter superscript are significantly different ($p < 0.05$).

The weight of the leaf extracts and their yield obtained from different solvent extracts of *L. camara* are presented in Table 2. Different solvents showed a significant influence on the total dry weight and yield of the extracts. Relatively, the extract from methanol resulted in superior extraction yield (14.4%) with $721.3 \pm 1.5 \mu\text{g}$ of dry weight. The recovery of extractable constituents from different extracts remained in the following order of methanol > ethyl acetate > acetone > chloroform. Our results are in conformity with previous studies supporting the use of methanol as the best solvent to recover higher extractable compounds from various medicinal plants [23, 24]. Similarly, Anwar et al. [16] have stated that methanol as the best solvent for extraction from *L. camara*. Also, methanol was commonly employed by other researchers in *L. camara* for various biological studies [8, 25–27]. However, literature study shows that there are no available reports on the comparative yield analysis obtained from different solvent extracts till date. The existence of significant differences of dry weight of the extracts and yields between various organic solvents can be because of different polar nature of the solvents tested [24].

Polyphenols and flavonoids are the plant secondary metabolites occurring in several medicinal plants known to possess antimicrobial, antioxidant, antispasmodic, antidepressant, antitumor, antimutagenic, anti-inflammatory, and many other biological activities [27, 28]. In plants, these phenolic compounds provide defense against various pathogens, regulate cell division and growth, and help in pigmentation and many other metabolic pathways [29]. Therefore, we investigated the occurrence of total phenolic and flavonoid content in different organic solvent extracts (Table 3). The results clearly indicated the existence of statistically significant differences ($p < 0.05$) among the different extracts. The highest phenolic content ($92.8 \pm 1.7 \text{ mg GAE/g}$) and flavonoid content ($26.5 \pm 0.5 \text{ mg RE/g}$) were observed in the methanol leaf extract of *L. camara*. On the other hand, ethyl acetate, acetone, and chloroform extract contained phenolic content of 75.6 ± 0.9 , 62.9 ± 1.7 , and $33.7 \pm 0.5 \text{ mg GAE/g}$, respectively. Total flavonoid content in ethyl acetate, acetone, and chloroform extract was found to be 16.7 ± 0.6 , 20.6 ± 0.3 , and $21.2 \pm 0.8 \text{ mg RE/g}$, respectively. Similarly, previous studies have shown the occurrence of rosmarinic and caffeic acid as the major phenolic compounds and few flavonoids such as 3,7-dimethoxy-, 3-methoxy-, and 3,7,4'-trimethoxy-quercetin, hispidulin, 3, pectolinarigenin 7-O- β -D-glucoside, and camaraside glycoside were evident in the plant extract

TABLE 3: Total phenolics and flavonoids content of different solvent extracts of *L. camara* leaves.

Solvent extracts	Total phenolic content (mg GAE/g) \pm SD	Flavonoid content (mg RE/g) \pm SD
Ethyl acetate	$75.6 \pm 0.9^{\text{b}}$	$16.7 \pm 0.6^{\text{c}}$
Methanol	$92.8 \pm 1.7^{\text{a}}$	$26.5 \pm 0.5^{\text{a}}$
Acetone	$62.9 \pm 1.7^{\text{c}}$	$20.6 \pm 0.3^{\text{b}}$
Chloroform	$33.7 \pm 0.5^{\text{d}}$	$21.2 \pm 0.8^{\text{b}}$

Note: each value is expressed as mean \pm standard deviation (SD) ($n = 3$). Values in the column followed by a different letter superscript are significantly different ($p < 0.05$) and values having the same letters are not statistically significant ($p < 0.05$). GAE: gallic acid equivalent, RE: rutin equivalent.

of *L. camara* [7, 11, 26]. Due to high polarity, methanol was found to exhibit better efficiency in extracting various polar phytochemicals (phenolics and flavonoids) from the leaves of *L. camara*. Both total phenolic and flavonoid contents obtained in our study were relatively much higher than the quantity obtained by earlier researchers in the same species but collected from geographically distant places [8, 16, 26]. However, total phenolic content was much lesser than the quantity ($245.5 \pm 3.5 \text{ mg gallic acid/g}$) as reported by Mahdi-Pour et al. [15] from methanol leaf extract of *L. camara* located in Kedah, Malaysia, and this difference can be attributed the influence of environmental factors and geographical location.

The formation of increased free radicals in human body may cause cell damage and induces various disorders such as cancer, myocardial infarction, atherosclerosis, and neurodegenerative disorders. However, antioxidant compounds derived from natural source or plants can repair these free radicals formed in cells and thereby, antioxidants are very useful in preventing various disorders [2, 15, 23]. The antioxidant capacities of either natural products or crude plant extracts are usually evaluated by making use of DPPH radical scavenging test [23]. In principal, the test depends on the capacity of DPPH free radicals reacting with plant metabolites such as phenolic and flavonoid compounds (H^+ donors) occurring in the sample. After the reaction, DPPH solution turns from purple to yellow color due to acquiring of a proton from the donor species. The intensity of color change directly relates to the scavenging ability of the biological sample [27]. Figure 1 shows the DPPH scavenging activity of *L. camara* leaf extracts in comparison to positive control (ascorbic acid). In all the solvent extracts, radical scavenging activity was found to be concentration dependent. The highest percentage of scavenging activity ($86.4 \pm 0.2 \mu\text{g/mL}$) was observed in methanolic leaf extracts at $500 \mu\text{g/mL}$ concentration. The next best solvent extract was found to be acetone ($80.5 \pm 0.3 \mu\text{g/mL}$) followed by ethyl acetate ($72.4 \pm 0.3 \mu\text{g/mL}$) and chloroform ($526.1 \pm 0.3 \mu\text{g/mL}$). However, the activities of all extracts were inferior to that of the ascorbic acid standard. The IC_{50} values of DPPH free radical scavenging activity were found to be in the following order: ascorbic acid ($80 \mu\text{g/mL}$) > methanol extract ($165 \mu\text{g/mL}$) > ethyl acetate ($200 \mu\text{g/mL}$) > acetone ($245 \mu\text{g/mL}$) > chloroform ($440 \mu\text{g/mL}$). These results are on a par with or even superior to that of reports by other researchers on several other crude extracts of the plant

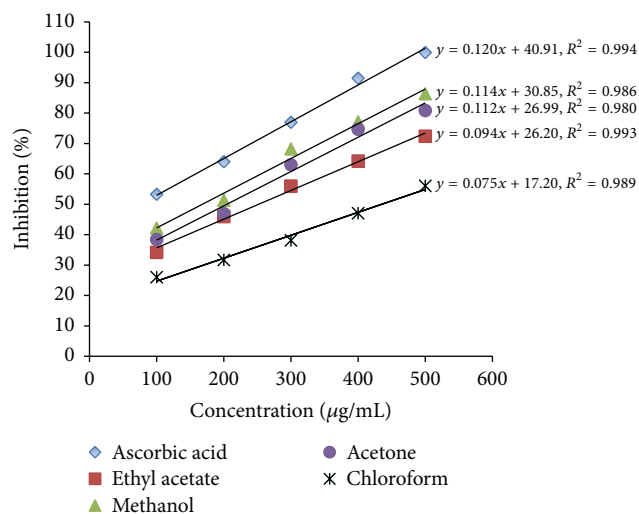


FIGURE 1: DPPH scavenging activities of various solvent extracts of *L. camara*.

evaluated under the same conditions using DPPH assay [27]. The higher antioxidant potential of plant extracts is correlated to the occurrence of many antioxidant compounds especially polyphenols [23]. Although the antioxidant activity of *L. camara* leaf extracts by using DPPH assay was reported by earlier researchers [15, 16, 26], none of them compared the effects of different solvents on antioxidant potential. In our study, due to higher solubility of antioxidant compounds, the methanol extract exhibited increased radical scavenging activity compared to other solvent extracts. Similarly, other researchers have stated that free radical scavenging potential of plant extracts depends mainly on the occurrence of bioactive compounds, particularly polyphenols [8, 30].

In biological system, a large quantity of hydroxyl radicals is formed due to activation of immune cells which are known to be highly toxic radicals and causes extreme damage to all molecules occurring in live cells. These radicals can trigger cell toxicity and mutagenesis by damaging DNA nucleotides [8, 31]. Therefore, measuring hydroxyl radical scavenging activity can provide good information on the antioxidant potential of *L. camara* leaf extracts obtained by different solvents. The results of hydroxyl radicals inhibition obtained from our study are depicted in Figure 2. It is evident that, irrespective of the organic solvents used for extraction, the percent inhibition of hydroxyl radicals increased with the concentration of the extracts. At higher concentration of 500 $\mu\text{g/mL}$, the percentage of hydroxyl scavenging activity for ascorbic acid, ethyl acetate, methanol, acetone, and chloroform extracts was found to be 88.1 ± 0.4 , 64.2 ± 1.0 , 76 ± 0.3 , 66.6 ± 0.5 , and $43.7 \pm 0.2\%$, respectively. The IC_{50} value was found to be in the following order of ascorbic acid (60 $\mu\text{g/mL}$) > methanol (110 $\mu\text{g/mL}$) > ethyl acetate (240 $\mu\text{g/mL}$) > acetone (300 $\mu\text{g/mL}$) > chloroform (510 $\mu\text{g/mL}$). The lower the IC_{50} value, the higher the scavenging activity and hence, methanol extract was found to possess superior antioxidant potential compared to other extracts tested. However, their effect was considerably lesser than the standard, ascorbic

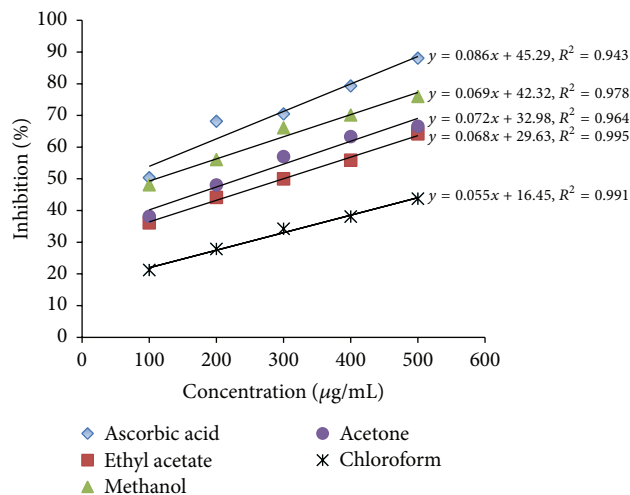


FIGURE 2: H_2O_2 scavenging activities of various solvent extracts of *L. camara*.

acid. These results completely corroborate the earlier reports that have proved the existence of correlation between the antioxidant property and the concentration and composition of different plant metabolites occurring in the extracts [23, 32]. In our study, methanol extracts contained significantly higher quantity of phenolics (92.8 ± 1.7 mg GAE/g) and flavonoids (26.5 ± 0.5 mg RE/g) and hence possessed superior antioxidant potential compared to other solvent extracts.

At present, the increased prevalence of deadly diseases and microbes adapting to antibiotic resistance is a great concern in the medical world [33]. Hence, more research interest is shown by medical community towards the development or discovery of novel antimicrobial agents. Due to the severe side effects of several synthetic antibiotics, research preference is mainly focused on discovering plant based natural drugs [5, 34]. Since *L. camara* possessed numerous secondary metabolites, we evaluated the effects of their different solvent extracts against some common human pathogenic bacterial strains. The results of our study revealed that all solvent extracts of *L. camara* were effective against both Gram positive and Gram negative bacterial strains tested, but their efficacy varied (Table 4). With the increase in the concentration, there was an enhanced antibacterial activity irrespective of the solvent used for extraction. Gram negative bacteria were more susceptible to all extracts compared to Gram positive bacteria. In general, Gram negative bacteria are known to exhibit high resistance towards wide range of chemical agents and antibiotics compared to Gram positive bacteria. Besides, Gram negative bacteria are reported to be the most prevailing pathogens causing a large number of deaths [34, 35]. Thus, *L. camara* leaf extracts can be more beneficial in treating most of these Gram negative disease causing pathogens. Methanol extract had the highest inhibition activity against all the tested microbes when compared to any other solvent extracts. The methanol leaf extract exhibited the highest activity against *E. coli* (24.1 ± 0.4 mm) followed by *K. pneumoniae* (18.1 ± 0.4 mm) at higher concentration (10 $\mu\text{L}/\text{disc}$). *S. aureus* was also more vulnerable to methanol extract with 18.0 ± 0.4 mm

TABLE 4: Antibacterial activity of different solvent extracts of *L. camara* at different concentrations.

Solvent extracts ($\mu\text{L}/\text{disc}$)	Zone of inhibition (mm)				
	<i>Escherichia coli</i>	<i>Klebsiella pneumoniae</i>	<i>Micrococcus luteus</i>	<i>Bacillus subtilis</i>	<i>Staphylococcus aureus</i>
Ethyl acetate					
2.5	08.2 \pm 0.8	06.9 \pm 0.5	06.1 \pm 0.7	10.0 \pm 0.5	—
5.0	13.0 \pm 0.9	08.3 \pm 0.5	07.1 \pm 0.3	13.8 \pm 0.5	06.1 \pm 0.1
10.0	12.0 \pm 0.5	10.5 \pm 0.8	07.0 \pm 0.5	13.9 \pm 0.4	10.6 \pm 0.4
Methanol					
2.5	14.6 \pm 0.5	08.2 \pm 0.3	08.1 \pm 0.7	10.0 \pm 0.3	14.1 \pm 0.2
5.0	18.2 \pm 0.2	14.5 \pm 0.5	12.2 \pm 0.3	14.0 \pm 0.6	16.1 \pm 0.6
10.0	24.1 \pm 0.4	18.1 \pm 0.4	18.0 \pm 0.5	16.1 \pm 0.2	18.0 \pm 0.4
Acetone					
2.5	16.1 \pm 0.4	06.1 \pm 0.6	07.9 \pm 0.2	14.2 \pm 0.3	05.8 \pm 0.6
5.0	24.0 \pm 0.4	10.1 \pm 0.6	12.3 \pm 0.2	15.9 \pm 0.4	10.2 \pm 0.2
10.0	28.2 \pm 0.6	16.2 \pm 0.2	12.2 \pm 0.2	16.3 \pm 0.4	12.2 \pm 0.4
Chloroform					
2.5	10.6 \pm 0.6	—	—	10.5 \pm 0.9	—
5.0	12.9 \pm 0.6	09.2 \pm 0.2	06.8 \pm 0.7	12.1 \pm 0.7	07.2 \pm 0.2
10.0	14.8 \pm 0.3	11.4 \pm 0.8	08.2 \pm 0.2	12.4 \pm 0.4	07.9 \pm 0.2

Note: the negative control discs were soaked with 50 μL DMSO and the positive control discs with 50 μL (50 $\mu\text{g}/\text{mL}$) chloramphenicol. Each value represents the mean \pm standard deviation (SD) of 3 replicates per treatment in 3 repeated experiments. “—” represents no activity.

of inhibition zone at 10 $\mu\text{L}/\text{disc}$ concentration. Among the tested bacterial strains, *E. coli* and *B. subtilis* showed increased zone of inhibition to all the solvent extracts and were the most susceptible strains. Ethyl extract was also most effective against *B. subtilis* while other bacterial strains were moderately inhibited. Likewise, acetone and chloroform extracts were also effective in inhibiting *E. coli* and *B. subtilis*. However, at lower concentrations, ethyl acetate extract showed no inhibition against *S. aureus*. In contrast, chloroform extract was less effective to all bacteria and at lower concentration, it failed to show antibacterial activity. *L. camara* finds its application in many parts of the world to cure various human ailments [7]. Previously, researchers have reported the varied antimicrobial potential of this plant and thus our reports support these findings [13, 14]. These results substantiate the findings of Naz and Bano [26] where methanol extract of *L. camara* leaves showed the highest antimicrobial activity. However, this is the first ever attempt which emphasizes the influence of different solvent extracts on human pathogenic bacterial strains. Our study clearly indicated the existence of considerable differences in the antibacterial activity among the various solvent extracts evaluated. This could be due to varied phytochemical constituents present in different solvent extracts.

Further, we used GC-MS analysis to identify the bioactive compounds occurring in the most competent solvent extract of *L. camara*. GC-MS profiling was performed only for methanolic leaf extract due to the fact that it contained many phytochemicals and exhibited superior biological activities. The distinctive chromatogram of the methanolic leaf extract of *L. camara* is shown in Figure 3. The analysis separated and identified a total of 32 known compounds belonging to different chemical classes (Table 5). The major compounds

included hexadecanoic acid (5.197%), phytol (4.528%), caryophyllene oxide (4.605%), 9,12,15-octadecatrienoic acid, methyl ester, (Z,Z,Z)- (3.751%), 2,3-dihydro-2,5-dihydroxy-6-methyl-4H-pyran-4-one (2.954%), α -D-galactopyranoside, methyl (2.790%), coumaran (2.288%), germacrene-D (2.185%), bicyclo[5.2.0]nonane, 2-methylene-4,8,8-trimethyl (2.065%), spathulenol (1.888%), 1,2,3-propanetriol, 1-acetate (1.689%), propane-1,2,3-triol (1.1615), and 2,4(1H,3H)-pyrimidinedione, 5-methyl- (1.180%). Few other compounds identified in the extract are α -elemol, myristic acid, neophytadiene, furfuryl alcohol, propargyl alcohol, and acetic acid, fluoro-, ethyl ester. Many of these identified constituents are known to possess several pharmacological activities. Hexadecanoic acid, a major phytoconstituent of *L. camara* methanolic leaf extract, is known to possess strong antimicrobial activity [36]. The diterpene, phytol, is an important compound reported with antioxidant, cytotoxic, and antimicrobial properties [37]. Similarly a conjugated saponin, 2,3-dihydro-2,5-dihydroxy-6-methyl-4H-pyran-4-one, is reported to possess strong antioxidant, anticancer, and anti-inflammatory properties [38, 39]. Caryophyllene oxide, spathulenol, and germacrene-D are known to possess anticarcinogenic, anti-inflammatory, and antibacterial properties [2, 40]. As a biofumigant, coumaran is reported to act against insect pests found in stored food grains [41]. The compound 2,3-dihydro-2,5-dihydroxy-6-methyl-4H-pyran-4-one has been reported in plant extracts exhibiting antioxidant, antiproliferative, and anti-inflammatory properties [40]. More recently, anti-inflammatory and cytotoxicity activities of hexadecanoic acid, methyl ester have been reported by Othman et al. [42]. However, pharmacological activities of other compounds of *L. camara* methanolic leaf extract are yet to be determined. Therefore, we assume that the strong bioactivities exhibited

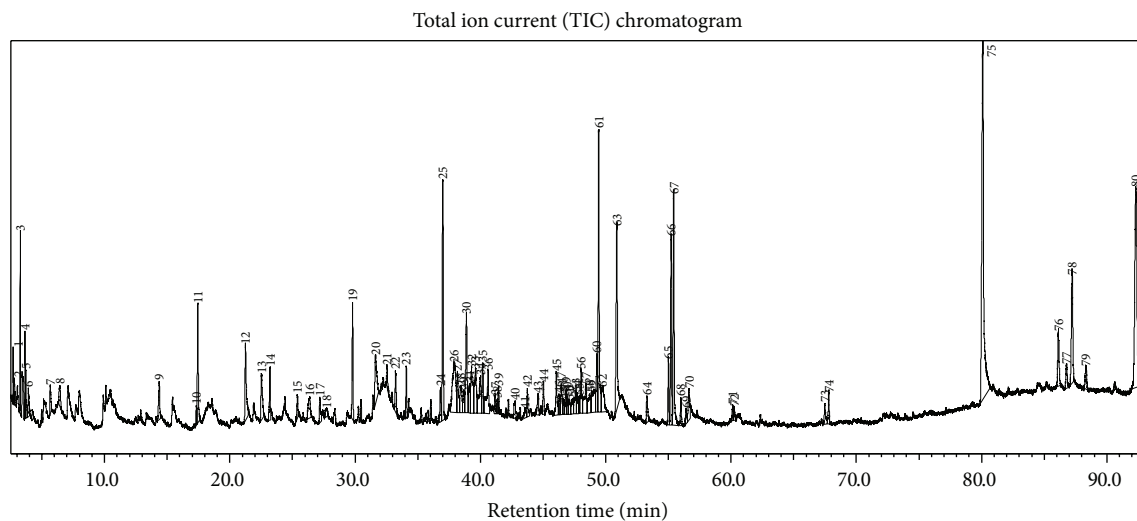


FIGURE 3: GC-MS chromatogram of methanolic leaf extract of *L. camara*.

TABLE 5: The major phytochemicals detected in the methanolic leaf extract of *L. camara* by GC-MS analysis.

S. number	Name of the compound	Peak number*	Retention time (min)	Area (%)
1	2-Propanone, 1-hydroxy-	1	2.703	0.481
2	Propane-1,2,3-triol	3	3.275	1.161
3	Propargyl alcohol	4	3.653	0.877
4	Acetic acid, fluoro-, ethyl ester	5	3.719	0.737
5	Furfuryl alcohol	7	5.672	0.750
6	2,4(1H,3H)-Pyrimidinedione, 5-methyl-	9	14.366	1.180
7	2,3-Dihydro-2,5-dihydroxy-6-methyl-4H-pyran-4-one	11	17.443	2.954
8	Coumaran	12	21.251	2.288
9	1,2,3-Propanetriol, 1-acetate	13	22.529	1.689
10	Cyclohexasiloxane, dodecamethyl-	14	23.200	0.978
11	4-Vinylguaiaicol	15	25.393	0.809
12	Bicyclo[5.2.0]nonane, 2-methylene-4,8,8-trimethyl	19	29.798	2.065
13	Germacrene-D	21	32.547	2.185
14	Longifolene	22	33.22	0.889
15	4-Epi-cubedol	23	34.084	0.817
16	Caryophyllene oxide	25	36.995	4.605
17	α -D-Galactopyranoside, methyl	26	37.909	2.790
18	Humulene epoxide II	27	38.173	0.696
19	Spathulenol	30	38.886	1.888
20	α -Elemol	34	39.997	0.507
21	Myristic acid	42	43.751	0.652
22	Neophytadiene	45	46.059	0.820
23	3-Eicosyne	53	47.594	0.211
24	9-Octadecenoic acid (Z)-, methyl ester	60	49.296	0.093
25	Hexadecanoic acid, methyl ester	61	49.438	0.696
26	Hexadecanoic acid	63	50.888	5.197
27	9,12-Octadecadienoic acid (Z,Z)-, methyl ester	65	55.007	1.037
28	9,12,15-Octadecatrienoic acid, methyl ester, (Z,Z,Z)-	66	55.225	3.751
29	Phytol	67	55.426	4.528
30	Octadecanoic acid, methyl ester	68	56.428	0.330
31	9,12,15-Octadecatrienoic acid, (Z,Z,Z)-	70	56.640	1.019
32	Phthalic acid, di(2-propylpentyl) ester	74	67.804	0.688

* Peak number is represented in Figure 3.

by *L. camara* in this study are correlated to the occurrence of these bioactive compounds in the methanol solvent extract. However, further studies on the isolation, characterization, and biological evaluation of these identified compounds are necessary to confirm their potential benefits.

4. Conclusion

In conclusion, the present investigation clearly revealed that phytochemical composition of *L. camara* leaf extract varied with respect to different solvents. Total phenolic and flavonoid content significantly varied among the different solvent extracts. Methanol solvent extract of *L. camara* leaves contained more extractable metabolites compared to any other solvents. Moreover, all solvent extracts of *L. camara* showed considerable antioxidant and antimicrobial activity with varying differences due to differences in their phytochemical composition. Thus, our study suggests that methanol leaf extract of *L. camara* containing many bioactive compounds may possibly be utilized as a therapeutical source for developing beneficial drugs to manage various human diseases and disorders.

Disclosure

Mohd. Sayeed Akhtar is a coauthor.

Conflict of Interests

The authors declare that there is no conflict of interests.

Acknowledgments

The authors are highly grateful to the Department of Crop Science, Universiti Putra Malaysia, Malaysia, for providing research facilities and the Department of Biosciences, Universiti Putra Malaysia, Malaysia, for GC-MS analysis.

References

- [1] Z. Gezahegn, M. S. Akhtar, D. Woyessa, and Y. Tariku, "Antibacterial potential of *Thevetia peruviana* leaf extracts against food associated bacterial pathogens," *Journal of Coastal Life Medicine*, vol. 3, no. 2, pp. 150–157, 2015.
- [2] M. K. Swamy and U. R. Sinniah, "A Comprehensive review on the phytochemical constituents and pharmacological activities of *Pogostemon cablin* Benth.: an Aromatic Medicinal Plant of Industrial Importance," *Molecules*, vol. 20, no. 5, pp. 8521–8547, 2015.
- [3] D. A. Lobo, R. Velayudhan, P. Chatterjee, H. Kohli, and P. J. Hotez, "The neglected tropical diseases of India and South Asia: review of their prevalence, distribution, and control or elimination," *PLoS Neglected Tropical Diseases*, vol. 5, no. 10, Article ID e1222, 2011.
- [4] R. C. Moellering Jr., J. R. Graybill, J. E. McGowan Jr., and L. Corey, "Antimicrobial resistance prevention initiative-an update: proceedings of an expert panel on resistance," *American Journal of Infection Control*, vol. 35, no. 9, p. -S23, 2007.
- [5] I. M. Ayoub, M. El-Shazly, M.-C. Lu, and A. N. B. Singab, "Antimicrobial and cytotoxic activities of the crude extracts of *Diets bicolor* leaves, flowers and rhizomes," *South African Journal of Botany*, vol. 95, pp. 97–101, 2014.
- [6] S. K. Mohanty, K. Malappa, A. Godavarthi, B. Subbanarasi-man, and A. Maniyam, "Evaluation of antioxidant, in vitro cytotoxicity of micropropagated and naturally grown plants of *Leptadenia reticulata* (Retz.) Wight & Arn.-an endangered medicinal plant," *Asian Pacific Journal of Tropical Medicine*, vol. 7, no. 1, pp. S267–S271, 2014.
- [7] E. L. Ghisalberty, "*Lantana camara* L. (Verbenaceae)," *Fitoterapia*, vol. 71, no. 5, pp. 467–486, 2000.
- [8] S. Kumar, R. Sandhir, and S. Ojha, "Evaluation of antioxidant activity and total phenol in different varieties of *Lantana camara* leaves," *BMC Research Notes*, vol. 7, no. 1, p. 560, 2014.
- [9] S. Patel, "A weed with multiple utility: *lantana camara*," *Reviews in Environmental Science and Biotechnology*, vol. 10, no. 4, pp. 341–351, 2011.
- [10] T. Venkatachalam, V. K. Kumar, P. K. Selvi, A. O. Maske, and N. S. Kumar, "Physicochemical and preliminary phytochemical studies on the *Lantana camara* (L.) fruits," *International Journal of Pharmacy and Pharmaceutical Sciences*, vol. 3, no. 1, pp. 52–54, 2011.
- [11] J. A. Parrotta, *Healing Plants of Peninsular India*, CABI International, New York, NY, USA, 2001.
- [12] S. B. Yadav and V. Tripathi, "A new triterpenoid from *Lantana camara*," *Fitoterapia*, vol. 74, no. 3, pp. 320–321, 2003.
- [13] A. Saraf, S. Quereshi, K. Sharma, and N. A. Khan, "Antimicrobial activity of *Lantana camara* L.," *Journal of Experimental Sciences*, vol. 2, no. 10, pp. 50–54, 2011.
- [14] D. Ganjewala, S. Sam, and K. Hayat Khan, "Biochemical compositions and antibacterial activities of *Lantana camara* plants with yellow, lavender, red and white flowers," *EurAsian Journal of Biosciences*, vol. 3, no. 10, pp. 69–77, 2009.
- [15] B. Mahdi-Pour, S. L. Jothy, L. Y. Latha, Y. Chen, and S. Sasidharan, "Antioxidant activity of methanol extracts of different parts of *Lantana camara*," *Asian Pacific Journal of Tropical Biomedicine*, vol. 2, no. 12, pp. 960–965, 2012.
- [16] F. Anwar, N. Shaheen, G. Shabir, M. Ashraf, K. M. Alkharfy, and A.-H. Gilani, "Variation in antioxidant activity and phenolic and flavonoid contents in the flowers and leaves of *Ghaneri* (*Lantana camara* L.) as affected by different extraction solven," *International Journal of Pharmacology*, vol. 9, no. 7, pp. 442–453, 2013.
- [17] O. P. Sharma, J. Vaid, and P. D. Sharma, "Comparison of lantadenes content and toxicity of different taxa of the *lantana* plant," *Journal of Chemical Ecology*, vol. 17, no. 11, pp. 2283–2291, 1991.
- [18] J. B. Harborne, *Phytochemical Methods*, Chapman & Hall, London, UK, 1998.
- [19] N. Pokharen, S. Dahal, and M. Anuradha, "Phytochemical and antimicrobial studies of leaf extract of *Euphorbia neriifolia*," *Journal of Medicinal Plants Research*, vol. 5, no. 24, pp. 5785–5788, 2011.
- [20] M. Kumara Swamy, K. M. Sudipta, P. Lokesh et al., "Phytochemical screening and in vitro antimicrobial activity of *Bougainvillea spectabilis* flower extracts," *International Journal of Phyto-medicine*, vol. 4, no. 3, pp. 375–379, 2012.
- [21] R. K. Salar and L. Seasotiya, "Free radical scavenging activity, phenolic contents and phytochemical evaluation of different extracts of stem bark of *Butea monosperma* (Lam.) kuntze," *Frontiers in Life Science*, vol. 5, no. 3-4, pp. 107–116, 2011.

- [22] J. Zhishen, T. Mengcheng, and W. Jianming, "The determination of flavonoid contents in mulberry and their scavenging effects on superoxide radicals," *Food Chemistry*, vol. 64, no. 4, pp. 555–559, 1999.
- [23] S. K. Mohanty, K. Malappa, A. Godavarthi, B. Subbanarasiman, and A. Maniyam, "Evaluation of antioxidant, in vitro cytotoxicity of micropropagated and naturally grown plants of *Leptadenia reticulata* (Retz.) Wight & Arn.-an endangered medicinal plant," *Asian Pacific Journal of Tropical Medicine*, vol. 7, supplement 1, pp. S267–S271, 2014.
- [24] B. Sultana, F. Anwar, and M. Ashraf, "Effect of extraction solvent/technique on the antioxidant activity of selected medicinal plant extracts," *Molecules*, vol. 14, no. 6, pp. 2167–2180, 2009.
- [25] D. Bhakta and D. Ganjewala, "Effect of leaf positions on total phenolics, flavonoids and proanthocyanidins content and antioxidant activities in *Lantana camara* (L)," *Journal of Scientific Research*, vol. 1, no. 2, pp. 363–369, 2009.
- [26] R. Naz and A. Bano, "Phytochemical screening, antioxidants and antimicrobial potential of *Lantana camara* in different solvents," *Asian Pacific Journal of Tropical Disease*, vol. 3, no. 6, pp. 480–486, 2013.
- [27] C. L. Apetrei, C. Tuchilus, A. C. Aprotosoia, A. Oprea, K. E. Malterud, and A. Miron, "Chemical, antioxidant and antimicrobial investigations of *Pinus cembra* L. bark and needles," *Molecules*, vol. 16, no. 9, pp. 7773–7788, 2011.
- [28] A. Ghasemzadeh and N. Ghasemzadeh, "Flavonoids and phenolic acids: role and biochemical activity in plants and human," *Journal of Medicinal Plant Research*, vol. 5, no. 31, pp. 6697–6703, 2011.
- [29] V. Lattanzio, V. M. Lattanzio, and A. Cardinali, "Role of phenolics in the resistance mechanisms of plants against fungal pathogens and insects," *Phytochemistry: Advances in Research*, vol. 661, pp. 23–67, 2006.
- [30] J. Dai and R. J. Mumper, "Plant phenolics: extraction, analysis and their antioxidant and anticancer properties," *Molecules*, vol. 15, no. 10, pp. 7313–7352, 2010.
- [31] R. Manian, N. Anusuya, P. Siddhuraju, and S. Manian, "The antioxidant activity and free radical scavenging potential of two different solvent extracts of *Camellia sinensis* (L.) O. Kuntz, *Ficus bengalensis* L. and *Ficus racemosa* L.," *Food Chemistry*, vol. 107, no. 3, pp. 1000–1007, 2008.
- [32] P. Kuppusamy, M. M. Yusoff, N. R. Parine, and N. Govindan, "Evaluation of in-vitro antioxidant and antibacterial properties of *Commelina nudiflora* L. extracts prepared by different polar solvents," *Saudi Journal of Biological Sciences*, vol. 22, no. 3, pp. 293–301, 2015.
- [33] E. M. Tekwu, A. C. Pieme, and V. P. Beng, "Investigations of antimicrobial activity of some Cameroonian medicinal plant extracts against bacteria and yeast with gastrointestinal relevance," *Journal of Ethnopharmacology*, vol. 142, no. 1, pp. 265–273, 2012.
- [34] K. Canli, E. M. Altuner, and I. Akata, "Antimicrobial screening of *Mnium stellare*," *Bangladesh Journal of Pharmacology*, vol. 10, no. 2, pp. 321–325, 2015.
- [35] M. V. Villegas and J. P. Quinn, "An update on antibiotic-resistant gram-negative bacteria," *Infections in Medicine*, vol. 21, no. 12, pp. 595–599, 2004.
- [36] R. Preethi, V. V. Devanathan, and M. Loganathan, "Antimicrobial and antioxidant efficacy of some medicinal plants against food borne pathogens," *Advances in Biological Research*, vol. 4, no. 2, pp. 122–125, 2010.
- [37] L. S. Wei, W. Wee, J. Y. F. Siong, and D. F. Syamsumir, "Characterization of anticancer, antimicrobial, antioxidant properties and chemical compositions of *Peperomia pellucida* leaf extract," *Acta Medica Iranica*, vol. 49, no. 10, pp. 670–674, 2011.
- [38] J. O. Ban, I. G. Hwang, T. M. Kim et al., "Anti-proliferate and pro-apoptotic effects of 2,3-Dihydro-3,5-dihydroxy-6-methyl-4Hpyranone through inactivation of NF- κ B in human colon cancer cells," *Archives of Pharmacol Research*, vol. 30, pp. 1455–1463, 2007.
- [39] N. Sharma, K. W. Samarakoon, R. Gyawali et al., "Evaluation of the antioxidant, anti-inflammatory, and anticancer activities of *Euphorbia hirta* ethanolic extract," *Molecules*, vol. 19, no. 9, pp. 14567–14581, 2014.
- [40] A. I. Hussain, F. Anwar, S. T. H. Sherazi, and R. Przybylski, "Chemical composition, antioxidant and antimicrobial activities of basil (*Ocimum basilicum*) essential oils depends on seasonal variations," *Food Chemistry*, vol. 108, no. 3, pp. 986–995, 2008.
- [41] Y. Rajashekar, A. Raghavendra, and N. Bakthavatsalam, "Acetylcholinesterase inhibition by biofumigant (Coumaran) from leaves of *Lantana camara* in stored grain and household insect pests," *BioMed Research International*, vol. 2014, Article ID 187019, 6 pages, 2014.
- [42] A. Othman, N. Abdullah, S. Ahmad, I. Ismail, and M. Zakaria, "Elucidation of in-vitro anti-inflammatory bioactive compounds isolated from *Jatropha curcas* L. plant root," *BMC Complementary and Alternative Medicine*, vol. 15, no. 1, p. 11, 2015.