

Review



Cite this article: Llinàs-Arias P, Esteller M. 2017 Epigenetic inactivation of tumour suppressor coding and non-coding genes in human cancer: an update. *Open Biol.* **7**: 170152.
<http://dx.doi.org/10.1098/rsob.170152>

Received: 21 June 2017
Accepted: 2 August 2017

Subject Area:
biochemistry/genetics/molecular biology

Keywords:
epigenetics, inactivation, methylation, cancer

Author for correspondence:
Manel Esteller
e-mail: mesteller@idibell.cat

Epigenetic inactivation of tumour suppressor coding and non-coding genes in human cancer: an update

Pere Llinàs-Arias¹ and Manel Esteller^{1,2,3}

¹Cancer Epigenetics Group, Cancer Epigenetics and Biology Program (PEBC), Bellvitge Biomedical Research Institute (IDIBELL), Barcelona, Catalonia, Spain

²Physiological Sciences Department, School of Medicine and Health Sciences, University of Barcelona (UB), Carrer de la Feixa Llarga, s/n, 08908 L'Hospitalet, Barcelona, Catalonia, Spain

³Institució Catalana de Recerca i Estudis Avançats (ICREA), Barcelona, Catalonia, Spain

PL-A, 0000-0002-3666-0532; ME, 0000-0003-4490-6093

Cancer cells undergo many different alterations during their transformation, including genetic and epigenetic events. The controlled division of healthy cells can be impaired through the downregulation of tumour suppressor genes. Here, we provide an update of the mechanisms in which epigenetically altered coding and non-coding tumour suppressor genes are implicated. We will highlight the importance of epigenetics in the different molecular pathways that lead to enhanced and unlimited capacity of division, genomic instability, metabolic shift, acquisition of mesenchymal features that lead to metastasis, and tumour plasticity. We will briefly describe these pathways, focusing especially on genes whose epigenetic inactivation through DNA methylation has been recently described, as well as on those that are well established as being epigenetically silenced in cancer. A brief perspective of current clinical therapeutic approaches that can revert epigenetic inactivation of non-coding tumour suppressor genes will also be given.

1. Introduction

Cell division is the molecular mechanism that allows us to grow, adapt and recover from stress. Essentially, it is a controlled process that keeps us alive. Tumour cells are the result of distortion of these mechanisms. They can grow faster and adapt better, living at our expense. They are an improved version of ourselves [1]. Cancer cells result from a set of aberrant alterations of DNA that lead to uncontrolled cell division. For many years the nature of these alterations has been studied. In 1969, Todaro and co-workers demonstrated that the administration of viral DNA and RNA had the ability to generate tumours [2,3]. This finding led to the discovery of the oncogenes, in other words, genes involved in growth and proliferation, which when deregulated contribute to malignant transformation. Two years later, Knudson identified another type of cancer-related gene through his retinoblastoma cancer studies, the tumour suppressor genes [4]. The function of this class of gene was none other than counteracting oncogenes, inhibiting growth in the absence of stimuli. A decade later, cancer studies met epigenetics. Epigenetics was first defined by Waddington in 1939 as 'the causal interactions between genes and their products, which bring the phenotype into being' [5]. Later the term was used to describe the occurrence of heritable changes in gene expression for which the DNA sequence is not altered [6]. One of the ways in which epigenetics is manifested is through DNA methylation. It was during the 1980s that a decrease in overall methylation levels was reported in the genomes of tumour cells [7]. This discovery was accompanied by the finding of oncogene activation through hypomethylation [8]. Paradoxically, a few years later hypermethylation and silencing of tumour suppressor genes were observed [9]. Another level of epigenetic regulation is the influence

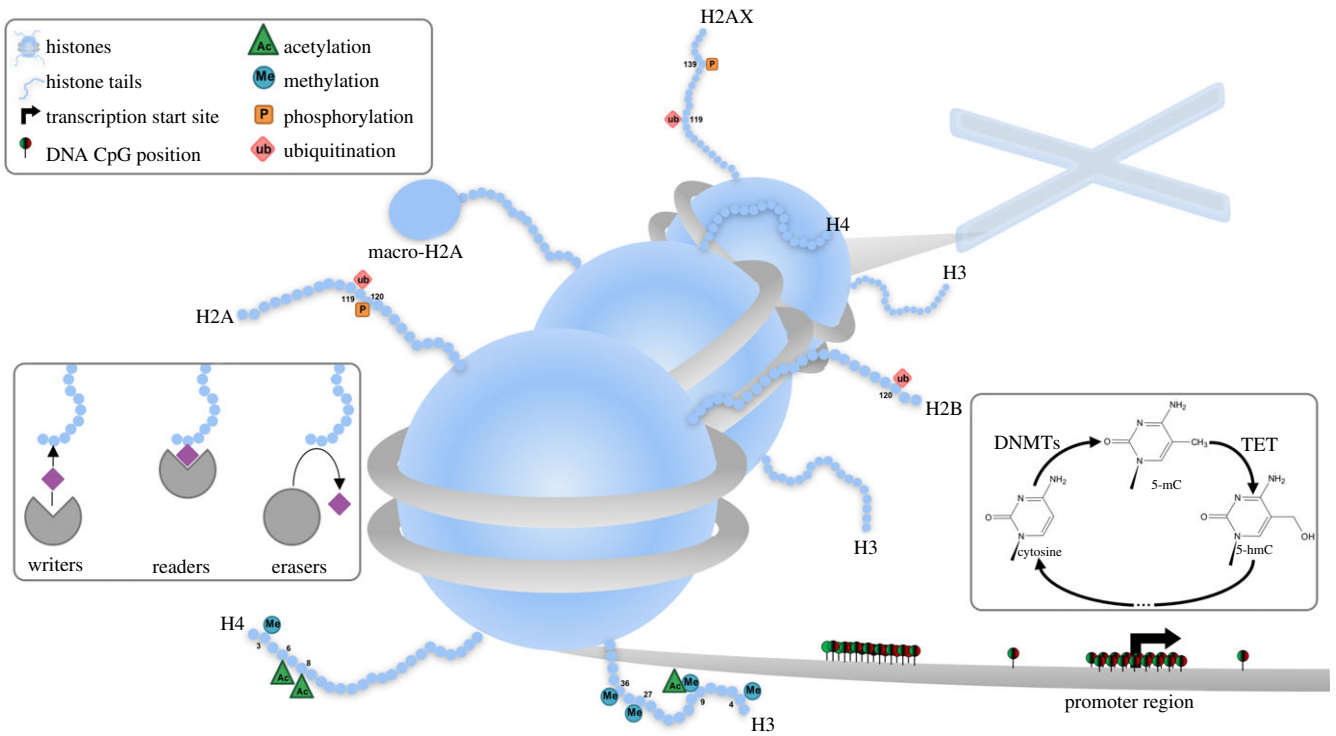


Figure 1. Covalent mechanisms of epigenetic regulation. DNA is bound to histone proteins forming the nucleosomes. Nucleosome compaction depends on histone tail modification, which is regulated by histone writers, readers and erasers. DNA methylation in CpG dinucleotides is regulated by different enzymes. DNMT1 and DNMT3a mediate the 5′mC synthesis. TET proteins catalyse DNA demethylation.

of non-coding genes on protein expression regulation. In relation to gene silencing, microRNAs and ncRNAs may act as effectors, decreasing mRNA and protein levels of their target genes, but they can also undergo epigenetic regulation by other mechanisms such as DNA promoter methylation.

This review will focus on how epigenetic alterations contribute to the silencing of tumour suppressor coding and non-coding genes with particular emphasis on recent discoveries. Molecular changes that are involved in carcinogenesis, such as aberrant division, immortality, genomic instability, metastasis, metabolic reprogramming and tumour plasticity will be analysed, highlighting genes silenced through epigenetic mechanisms.

2. Molecular epigenetics mechanisms

Various types of epigenetic mechanisms have been defined. The first group of these consist of covalent modifications of chromatin, affecting DNA and histones (figure 1). In mammals, DNA methylation occurs predominantly at the 5′ position of cytosine forming cytosine guanine dinucleotides (CpG). This modification is carried out by DNA methyltransferases (DNMTs), enzymes that use *S*-adenosylmethionine (SAM) as methyl group donor. In humans, DNMT1 is responsible for *de novo* methylation, whereas DNMT3a and DNMT3b are more related with methylation maintenance [10]. Demethylation can be carried out through 10–11-translocation proteins (TET). These enzymes can convert 5-methylcytosine (5mC) into 5-hydroxymethylcytosine (5hmC). The abundance of this hydroxylated nucleotide in promoter regions of genes seems to correlate with their active expression, while 5mC is generally related to transcription repression. 5hmC is finally converted back into cytosine by the action of TETs and other enzymes through the successive modification of the 5-residue, concluding

the demethylation reaction [11]. DNA is not naked in the nucleus; it is associated with histones. Histones are grouped together to form octamers around which DNA is wrapped. Histones have an intrinsically unfolded domain, known as a histone tail, which can be highly modified. These modifications influence chromatin compaction, and may affect the binding affinity of different proteins and complexes for chromatin. According to histone marks, three groups of proteins are defined. Writers are proteins responsible for histone marks deposition while erasers act by removing these marks. Readers recognize these marks, and upon mark recognition they recruit various proteins with different functions that depend on chromatin context. The nature and position of the mark determines its role [12]. Thus, the same modification in one position may play an activating role, such as trimethylation in lysine 4 of histone 3 (H3K4Me3), but in another position may mediate a repressive action, such as trimethylation of lysine 9 of histone 3 (H3K9Me). On the other hand, modification of the same position with different tags may give rise to opposite effects: whereas H3K9Me promotes repression, acetylation in the same position activates transcription. Beyond covalent modifications there are other epigenetic mechanisms to be considered. Among them there is the replacement of histones, such as macroH2A or H2AX [13,14]; ATP-dependent chromatin remodelling complexes, which are involved in nucleosome positioning; and non-coding RNAs (ncRNAs). ncRNAs participate in gene expression regulation in several ways. They can activate gene expression by forming DNA–RNA complexes, such as R-loops, but more often they interfere in mRNA translation by blocking ribosome binding or promoting mRNA degradation [15].

3. Aberrant division

The most prominent feature of tumour cells is their uncontrolled division. This ability comes from the deregulation of genes

responsible for cell-cycle control, as well as genes related to signal transduction pathways involved in detecting external stimuli such as nutrients or mitogens. While alterations in cell-cycle genes often result in a lack of checkpoints, changes in transduction pathways eventually lead to pervasive activation of them in a stimulus-independent manner. The cell cycle is broadly controlled by cyclin-dependent kinases (CDKs). These enzymes catalyse phosphorylation reactions of different substrates in association with their regulatory subunits, the cyclin proteins, which increase CDK activity and contribute to target recognition. A well-programmed fluctuation of cyclin expression entails progression through different phases on the cell cycle. Contrary to what one might think, the critical step in cancer cells is neither the S-phase nor mitosis itself, but rather G₀/G₁ transition, when a cell decides its fate. During this phase, CDK4/6 control G₀/G₁ transition, and are activated by D cyclins (D1, D2, D3) [16]. Moreover, CDK activity is controlled by CDK inhibitors (CKIs), which are also regulated by external stimuli. For instance, p15^{INK4B}/CDKN2B inhibits CDK4/6 activity induced by TGF- β [17]. Thus, CDK4/6 inhibition, either by the lack of cyclins or by the activation of CKIs, will result in a cessation of cellular proliferation. Cyclin expression is dependent on extracellular signal pathways that are often altered in cancer, such as Wnt/ β -Catenin, RANK receptor, Shh/Patched or tyrosine kinase receptors. p16^{INK4A}/CDKN2A and p15^{INK4B}/CDKN2B, which are CKIs for CDK4/6, are downregulated in several types of cancer [18]. In some, such as lymphoma, gastric cancer, head and neck squamous cell carcinoma (HNSC), liver and oesophageal cancers [18–22], p16^{INK4A}/CDKN2A downregulation is caused by promoter hypermethylation, but it can also be due to histone deacetylase (HDAC) activity [23]. Retinoblastoma protein (pRB) is the master regulator of G₀/G₁ to S transition, translating cyclin fluctuations into transcription factor activation. pRB is hypophosphorylated during early G₁ phase by CDK4/6 and hyperphosphorylated during late G₁ by E-CDK2 complex. Low levels of phosphorylation allow pRB binding with E2F transcription factors, whereas pRB hyperphosphorylation liberates E2Fs from pRB control, enabling cell-cycle progression. pRB, which is encoded by *RB1* gene, is one of the most commonly mutated genes in cancer [24]. Apart from genetic mutations, epigenetic silencing has also been observed. Cells without functional pRB are not able to maintain themselves in G₀/G₁, and so undergo aberrant division [25,26]. *miR-124a*, *miR-129* and *miR-137* are downregulated through promoter hypermethylation on colorectal cancer (CRC) and breast cancer cell lines. These microRNAs target CDK6, so their silencing promotes CDK6 activation, which triggers increased phosphorylation of pRb by promoting cell-cycle progression [27–29].

Receptor tyrosine kinases (RTKs) provide one way by which cells sense their environment. Their ligand binding activation promotes conformational changes in their cytoplasmic domain releasing signals into the cytoplasm. MET is a RTK which is targeted by *miR-1-1* and *miR-34a* microRNAs. All of them are downregulated through promoter hypermethylation resulting in MET activation in some cancers [30,31]. IGF-1R is another RTK that is also regulated by *miR-214* [32] and *miR-345* [33], which in turn are aberrantly methylated in cancer [34]. *FGFR* expression is negatively controlled by *miR-9* family, but these microRNAs are frequently silenced by promoter hypermethylation in cancer [35]. Post-translational modifications of RTKs recruit a number of proteins that can activate RAS, promoting its GDP release and preferential GTP binding.

RAS can activate the mitogen-activated protein kinase (MAPK) pathway by interacting with B-RAF, a MAPK pathway effector. B-RAF activates PERK, which in turn catalyses ERK phosphorylation. The MAPK pathway activates cytoplasmic and nuclear proteins involved in activating Jun and Fos transcription factors, as well as protein synthesis activation that contributes to cyclin D1 expression. Constitutive activation of the MAPK pathway is often observed in cancer cells due to epigenetic alterations of MAPK-related genes as well as mutations. For example, PTPRR, which dephosphorylates ERK, is methylated in cervical cancer [36]. DUSP1 and MKP1 are phosphatases which also target ERK, promoting MAPK pathway inactivation. The promoter regions of the *DUSP1* and *MKP1* genes have been found hypermethylated in oral squamous cell carcinoma and breast cancer, respectively [37,38]. RAS is directly regulated by *Let-7a* [39], a microRNA which is downregulated in different cancers such as head and neck cancer (HNC) through promoter hypermethylation [40].

The phosphatidylinositol 3-kinase (PI3K)/AKT pathway is also a RAS-dependent pathway. PI3K activation depends on its interaction with the GTP-bound form of RAS. Phosphatidylinositol (PI) is a component of the plasma membrane which can have a regulatory role that depends on its post-translational changes. After its phosphorylation by different kinases, such as PDK1, phosphatidylinositol-(4,5)-bisphosphate (PIP2) can be phosphorylated by PI3K, generating phosphatidylinositol-(3,4,5)-triphosphate (PIP3), which is recognized by AKT/PKB. PDK1 is targeted by *miR-375*, which is downregulated by promoter hypermethylation [41]. Active AKT/PKB contributes to cell proliferation by p21/CDKN1A, Tsc2 and GSK3- β inhibition [42]. The critical step for AKT/PKB activation is PIP3 formation. Alternatively, PIP2 can be cleaved by phospholipase C (PLC), generating diacylglycerol (DAG) and inositol-(1,4,5)-triphosphate (IP3). PLCD1 has been found silenced in breast and gastric cancers by promoter hypermethylation [43]. It can be considered a tumour suppressor gene, as its activity reduces AKT/PKB activation and decreases PIP2 levels, which are related to metastatic events [43–45]. There is another family of proteins which interact with RAS, the Ras-association domain family (RASSF). These proteins interact with the activated form of RAS, often playing a different role in tumorigenesis, because they behave as tumour suppressors in contrast with PI3K or B-RAF. RASSF1 is the most famous member of this family. Its promoter has been found methylated in more than 30 tumour types [46,47]. RASSF1 activation promotes cyclin D1 downregulation by its interaction with p120E4F, which in turn interacts with p14^{ARF}, pRB and p53 [48].

Apart from RTKs, the Wnt/ β -Catenin signalling pathway is another canonical pathway altered through promoter hypermethylation in cancer cells. In the absence of Wnt signalling, β -Catenin is degraded by the GSK3- β , Axin and APC destruction complex. GSK3- β phosphorylates β -Catenin leading to its degradation by the proteasome. When Wnt interacts with Frizzled, Disheveled (DSH) is released and displaces GSK3- β , promoting β -Catenin activation. This protein is then translocated to the nucleus, where it binds Lef/TCF, promoting transcription of its target genes including *MYC* and *CNND1* (cyclin D1), both considered proto-oncogenes due to their capacity to enhance cell growth and cell proliferation [49]. Epigenetic alterations of this pathway generate high β -Catenin levels. Secreted frizzled related proteins (SFRP1, SFRP2, SFRP3), which are Frizzled competitors for Wnt, are silenced by promoter hypermethylation in different cancers, including

hepatocellular carcinoma (HCC), lung adenocarcinoma, oesophageal squamous cell carcinoma (ESCC) and CRC [49–52]. NDRG2 and WIF-1, other Wnt regulators, are inactivated in pancreatic cancer and ESCC, respectively [53,54]. It is important to mention that some Wnt ligands cannot activate β -Catenin, and so act as Wnt/ β -Catenin antagonists, examples of which include WNT5a and WNT7a, both of which are commonly silenced in CRC, pancreatic and lung cancer [55,56]. APC is frequently mutated in CRC, but it is also inactivated by promoter hypermethylation in colon, breast and pancreatic cancer [57–59]. NKD2, DACT2 and CXXC4 inhibit DSH. However, during tumorigenesis they are silenced by promoter hypermethylation [60,61]. ROR2 is a Wnt5a receptor of Frizzled independent pathway, which inhibits the canonical Wnt/ β -Catenin pathway. This transmembrane protein is also downregulated in CRC through promoter hypermethylation [62].

Cancer cells often show myc activation through different mechanisms, such as myc amplification, but its activity can also be indirectly regulated through epigenetic events [63]. In acute myeloid leukaemia (AML), promoter hypermethylation of NUDT16, an RNA decapping enzyme, triggers an increase in c-myc half-life mRNA, contributing to its activation [64]. Extensive literature highlights the importance of epigenetic silencing events in tumour suppressor genes involved in cell-cycle control and mitogenic pathways.

4. Immortality

Tumour cells are considered immortal because they avoid cell death mechanisms that normally occur to avoid uncontrolled cell growth. This capacity can be given by telomerase activation through epigenetic deregulation or by the inhibition of pathways related to cell death. Tumour cells are subjected to different stress conditions, both extrinsic, such as oxygen and nutrient deprivation or death signals, and intrinsic, including DNA damage or ROS stress. These situations lead normal cells to die either by apoptosis or by other mechanisms. Therefore, tumour cells find ways to prevent cell death through inhibition of key genes involved in this programme. The crucial role of epigenetics in this reprogramming was observed by Kaminskyy *et al.* [65]. When they inhibited DNMTs and HDACs, apoptosis was reactivated. p53 is considered the master regulator of programmed cell death. Its activation, which is related to DNA damage or hypoxia, is highly regulated. One of these regulators is mouse double minute 2 (Mdm2), an E3-ubiquitin ligase which recognizes and mediates p53 degradation via the proteasome. Mdm2 transcription is upregulated by p53, generating a negative feedback, in order to avoid excessive p53 activity after its activation. Mdm2 activation depends on Akt/PKB phosphorylation, thereby connecting survival signals with the p53 regulation. Mdm2 is also regulated by p14/ARF, an E2F target gene that is often silenced in cancer through DNA hypermethylation of its promoter [66–68]. Thus, p53 deregulation can be explained in some cancers with p53 wild-type copies. When p53 is activated it promotes the transcription of growth arrest genes, such as p21/CDKN1A, a CDK inhibitor; 14-3-3 σ , which sequesters B-CDC2; and Reprimo, which promotes G2 arrest. 14-3-3 σ is silenced in a broad range of cancer types including nasopharyngeal carcinoma and breast cancer through promoter hypermethylation [69–71]. Meanwhile, Reprimo is also silenced by the same mechanism [72]. These epigenetic

alterations allow cell-cycle progression in spite of p53 activation. TP53TG1 is a long-non-coding RNA which is regulated by p53. This RNA contributes to the DNA damage response (DDR) through its interaction with YBX1 preventing its nuclear localization. TP53TG1 is often downregulated through promoter hypermethylation, which triggers a poor outcome in gastrointestinal cancer patients [73].

If the stimulus that activates p53 ceases, for instance DNA damage is repaired or survival signals bind membrane receptors, p53 levels decrease, promoting cell-cycle progression. However, if the DNA damage cannot be repaired or if there is a sustained lack of nutrients, cells can enter into apoptosis. In this case p53 promotes the transcriptional activation of Bcl-2 superfamily genes. Within this heterogeneous family there are antiapoptotic proteins, such as Bcl-L1 and Bcl-2 itself as well as proapoptotic genes, including BAX and Only-bh3 family proteins [74]. Proapoptotic proteins participate in the formation of mitochondrial channels, contributing to cytochrome C release. Some proapoptotic proteins including BAX, BIM, Bid, HRK and PUMA are silenced in cancer [75,76]. microRNA-7, which targets the antiapoptotic Bcl-2, is also downregulated in lung cancer cells [77]. Whereas most of them are silenced through DNA promoter hypermethylation, Bid inhibition is driven by SIN3a/HDAC1/2 corepressor complex [78]. Once cytochrome C is released it multimerizes with Apaf-1 and caspase 9, generating the apoptosome. Apaf-1 inhibition through DNA promoter hypermethylation allows apoptosis bypass in renal carcinomas [79]. Besides the intrinsic pathway, there is a p53-independent pathway, the extrinsic apoptosis pathway, that is activated by cytokines. When FAS receptor recognizes Fas ligand, or tumour necrosis factor-related apoptosis-inducing ligand (TRAIL) binds its receptor DR4 (death receptor), the apoptosis programme is initiated. Subsequently, the death-induced signalling complex (DISC) is formed by recruitment of FADD (Fas-interacting DD), pro-Casp8 and proCasp10 proteins. ProCasps are then activated and switch on the intrinsic pathway either through Bid activation by cleavage or through activating executioner caspases (Casp3,6,7). After its activation, Bid migrates to mitochondria where channels are opened and cytochrome C is released, becoming part of the apoptosome. During tumorigenesis death signals that can activate this pathway remain active, but the inactivation of the transduction proteins is a common feature among cancers. The DR4 receptor is inhibited by DNA methylation of its promoter region in different cancers [80,81]. Concerning Fas receptor, its regulation by DNA methylation is controversial. Butler *et al.* [82] described that Fas promoter hypermethylation did not explain Fas downregulation in CRC. Three years later, Petak *et al.* [83] demonstrated that methylation of the enhancer region leads to Fas downregulation in this type of cancer. In addition, Fas has been found methylated in bladder and oesophageal cancer [84,85]. Surprisingly, its promoter methylation status did not correlate with its expression in colorectal RKO cells, but its expression did correlate with the methylation status of the enhancer region. FADD and Casp8 were also downregulated by promoter hypermethylation [81,85].

TMS1/ASC is a bipartite protein that plays a role in apoptosis and in the NF- κ B pathway. It may serve as an adaptor for Casp8 and Casp10 caspases or it can interfere with IKK complex formation, contributing to apoptosis by activating the caspase cascade as well as blocking antiapoptotic and proliferative gene expression. The TMS1/ASC promoter

was described to be hypermethylated in several cancer subtypes [86,87]. HACE1 is a protein which mediates TNFR1 activation, promoting apoptosis or necroptosis activation. Necroptosis is another form of programmed cell death that triggers membrane rupture and inflammation without the need for caspase activation. HACE1 has been found methylated in HCC [88]. RIPK3 is also involved in this pathway, which in turn is also silenced through DNA promoter hypermethylation [89]. Epigenetic alterations related to the roles of p53 in mobilizing components of DNA repair machinery and inhibiting angiogenesis will be described in detail below in the Genomic instability and Tumour plasticity sections, respectively.

Overall, avoiding cell death is a key step for tumourigenesis, where epigenetic dysregulation plays an essential role, from p53 regulation to apoptosis and necroptosis effectors.

5. Genomic instability

In general, during cancer progression DNA methylation levels decrease. In some cancers, mutations in DNMTs and TET2 have been reported [90–92], but this is not a common feature. In most cases, the mechanisms that cause this phenomenon are unknown and do not seem to depend on any single pathway. Global DNA demethylation promotes genomic instability by various mechanisms. Eden *et al.* [93] demonstrated that mitotic recombination, which can lead to gene translocations and fusions, was increased in DNMT3a- and DNMT3b-deficient cells. In addition, DNA demethylation allows retrotransposon transcription, such as long interspaced nuclear elements (LINEs). LINE-1 transcription is activated after DNA demethylation, leading to the insertion of these elements into other genomic locations. During tumourigenesis, LINE-1 transcripts can cause gene disruption and can also function as alternative splicing sites, as novel promoters or as polyadenylation signals in a retrotransposition-independent manner [94]. LINE-1 internal bidirectional promoters can generate LINE-1 chimeric transcripts (LCTs) containing parts of genomic sequences surrounding the LINE-1 locus. Cruickshanks *et al.* [95] discovered that LCT13, a 300 kb LINE-1, was upregulated in cancer and contributed to TFPI-2 downregulation as an antisense gene, demonstrating an epigenetic inactivation caused by a LINE element.

DNA repair genes show a dual role during cancer progression. Its inactivation via mutations or epigenetic silencing promotes accumulation of repair errors, increasing tumour heterogeneity and provoking alterations which may confer different advantages to cancer cells [96]. Furthermore, a lack of DNA damage sensors or an efficient response to them can promote faster cell division and avoidance of apoptosis [97]. However, this damaged DNA becomes more fragile, so stimuli such as oxidative damage or certain drugs can be lethal for tumour cells, giving rise to its use as a therapeutic target [98].

The DDR begins with the recognition of damaged DNA by different sensors, which may recruit different mediators depending on the nature of the DNA damage detected. As well as activating DNA repair mechanisms, these mediators also participate in cell-cycle arrest, prioritizing DNA repair before replication or mitosis; if DNA damage cannot be removed, chronic DDR signalling may trigger cell death by apoptosis or cellular senescence. In humans, these DNA damage sensors are ataxia telangiectasia-mutated (ATM) and

ataxia telangiectasia and Rad3-related protein (ATR), which interact with double strand break (DSB) lesions [99]. On the other hand, following DNA lesions that generate ssDNA, replicative protein A (RPA) interacts with the DNA, positioning ATR interacting protein (ATRIP) close to the site of damage. ATM has been found hypermethylated in breast cancer [100], even before the appearance of palpable lesions [66]. The two best-studied ATM/ATR targets are CHK1 and CHK2, which stop cell-cycle progression by reducing CDK activity through different mechanisms. CHK2 has been found hypermethylated in gliomas [101]. ATM/ATR also catalyses Ser139- γ H2AX phosphorylation, located around DSB sites, promoting the recruitment of DDR factors.

DNA repair machinery is specific for each type of alteration. For example, direct DNA repair (DR) enzymes, base excision repair (BER), nuclear excision repair (NER) or mismatch repair (MMR) mechanisms are triggered when single nucleotide base damage occurs; by contrast, recombination repair mechanisms (non-homologous end joining (NHEJ) or homologous directed recombination (HDR)) are active when DSBs are induced. Deficiencies in DNA repair enzymes can be caused by somatic mutation, but are much more frequently caused by epigenetic alterations that reduce or silence their expression.

MGMT is an enzyme that directly repairs O-6-methylguanines. If it is inactive, unrepaired guanine will match with adenine instead of cytosine, giving rise to genomic instability. The MGMT promoter is often hypermethylated in oesophageal cancer and CRC [22,102], but its methylation status is especially taken into account in glioblastomas [103,104] because it is used as temozolomide response biomarker.

BER is a process which consists of the detection and correction of damaged bases in the DNA. This pathway is initiated by a number of glycosylases (e.g. Ogg1, MBD4, NEIL1), which differentially recognize and remove each aberrant base, generating an AP site (apurinic/apyrimidinic site). Ogg1 is a glycosylase that recognizes 8-oxoguanosine, being reported as inactivated by promoter DNA hypermethylation in breast cancer [105]. MBD4 recognizes thymidines or uracils produced by hydrolysis of cytosines in CpG sites, and it is methylated in CRC and ovarian cancer [106]. NEIL1 recognizes and removes oxidized pyrimidines. This gene is also methylated in certain types of cancer, such as HNC [107]. An AP endonuclease then cleaves the AP site, generating a single-strand break that can be processed by either a short-patch (just a nucleotide) or a long patch (from 5 to 10 nt). Finally, DNA replication machinery synthesis followed by XRCC1-ligase III α ligation ends the BER process.

NER is a similar process, which recognizes bulky DNA lesions produced by mutagens or UV irradiation. NER can be initiated by global genome NER (GG-NER) or transcription-coupled NER (TC-NER) on the transcribed strands of active genes [108]. In GG-NER, DNA lesions are mainly recognized by XPC-hHR23B-CETN2. After recognition, this complex degrades the DNA surrounding the lesion and recruits TFIIH. By contrast, TC-NER is activated when RNA polymerase II (RNAPII) is blocked by bulky adducts. Different factors including CSA, CSB and XAB2 interact with blocked RNAPII, promoting its backtracking and recruitment of TFIIH, the convergent step between GG-NER and TC-NER [108]. XPC and RAD23B are epigenetically silenced in cancer through DNA promoter hypermethylation in lung and multiple myeloma, respectively [109,110]. After recognition, TFIIH complex

proteins XPB and XPD unwind the DNA to create a 20- to 30-nt bubble. This event allows the recruitment of XPA, RPA, XPG and ERCC1–ERCC4/XPF. XPA binds the 5' side of the bubble and RPA interacts with the complementary ssDNA that does not have the lesion. Next, ERCC1–ERCC4/XPF cuts the 5' injured strand, and after patch synthesis by DNA replication machinery, XPG cuts the 3' strand. Epigenetic silencing of ERCC1 has been described in HNC through DNA promoter hypermethylation [111]. Finally, XRCC1-ligase III α seals the DNA [108].

DNA MMR machinery is responsible for replacing mismatched Watson–Crick nucleotides. Mismatches are recognized by the MutS heterodimer, which can consist of either MSH2–MSH6 (MutS α) or MSH2–MSH3 (MutS β). MutS is then stabilized by the MutL heterodimer, which in turn can be made of MLH1–PMS2 (MutL α), MLH1–PMS1 (MutL β) or MLH1–MLH3 (MutL γ) [112]. The MutS–MutL complex recruits different proteins that can generate an incision closer to the DNA injury and carry out the removal of the damaged strand. The DNA replicative machinery then resynthesizes the damaged strand anew. Genes involved in MMR are often altered in hereditary non-polyposis CRC syndrome (Lynch syndrome). For instance, MLH1 inactivation by promoter hypermethylation is a common alteration in this pathology [113].

HDR normally is found inactivated in cancer cells. When DNA damage occurs, H2A is replaced by γ H2AX in the nucleosomes near to the DNA lesion. After γ H2AX phosphorylation by ATM, MDC1 is recruited. MDC1 promotes RNF168/RNF8 binding and MRN complex interaction, formed by MRE11–RAD50–NBS1. The MRN complex stimulates the kinase activity of ATM [114] and participates in 5' \rightarrow 3' resection.

BRCA1 participates in HDR through RNF168/RNF8, a pair of E3-ubiquitin ligases that modify proteins at DSB sites. Next, BRCA1 mediates the 5' \rightarrow 3' resection. After 5' \rightarrow 3' resection by BRCA1, BRCA2 loads RAD-51 onto ssDNA, a necessary step for invading the homologous double helix. The homologous DNA is used as a template for damaged DNA strand resynthesis [115]. RAD51 is downregulated in renal cell carcinoma by histone methylation [116]. However, RAD51 seems to be overexpressed in many different types of cancer. RAD51 has five paralogues, which play essential roles in the HDR pathway [117]. Of these, RAD51B and XRCC3 are downregulated by DNA promoter hypermethylation in different cancer types, including HNSC, lung and cervix [118]. RNF168/RNF8 mediates BRCA1 or 53BP1 binding. BRCA1 is a key regulator of the HDR pathway and several mutations and epigenetic silencing events have been reported, especially in breast and ovarian cancers [119]. This inactivation leads to 53BP1 binding to DSB sites, activating NHEJ to the detriment of HDR [120]. BRCA1 can also be deregulated by epigenetic alterations of its partners, such as SRBC for which inactivation by promoter methylation has been described in CRC [121]. It has been also reported that some HDR-related helicases are silenced in different types of cancers, such as WRN, which is often methylated in cervical cancer [122]. DNA hypermethylation is also present on the SLFN11 promoter in different types of cancer [123]. SLFN11 is a putative helicase that interacts with DHX9, another BRCA1 interactor, and it has been postulated that SLFN11 destabilizes the DHX9 complex, thus impairing complete damage repair [124].

The alternative process to HDR following a DSB is NHEJ, which is more common during G₁ phase. When a DNA lesion occurs, BRCA1 and 53BP1 compete for binding to the

damage site. If 53BP1 interacts with phosphorylated γ H2AX or with the MRN complex, BRCA1-mediated resection is blocked, and NHEJ starts [125]. The Ku70/Ku80 heterodimer can bind DSB ends (non-resected ends). After its binding, these proteins recruit XRCC4, which may serve as scaffold for DNA end processing enzymes, such as ligase IV, WRN or a DNA polymerase [126]. Ku70/Ku80 also recruits DNA-PKcs, which can also process the damaged region. Ku80 and DNA-PKc (also called XRCC5 and XRCC7, respectively) have been found hypermethylated in glioma [127].

These data prompt us to think that DNA repair system inhibition is a double-edged sword for cancer cells, increasing their division rate and mutation accumulation but sensitizing them to certain types of damage inflicted by UV, ionizing radiation and chemotherapeutic agents.

6. Metastasis

Cancer cells can develop several strategies to adapt themselves to new environments. Epithelial-to-mesenchymal transition (EMT) is one of the most dramatic changes, whereby epithelial cells obtain mesenchymal features. These cells are poorly differentiated, and they can detach from the primary tumour, migrate through the stromal environment and reach the bloodstream. The reversibility of this process allows transformed cells to undergo mesenchymal-to-epithelial transition, and then continue proliferating at distant localizations. Carmona *et al.* [128] demonstrated common DNA methylation switches in MDCK and MDA-MB-468 associated with EMT phenotype acquisition after TGF- β treatment, supporting the importance of epigenetic alterations during this process. TGF- β activates several pathways, promoting the expression of transcription factors (TFs) that direct cell reprogramming, including FOXO1, TWIST and SNAIL [129]. Activation of these TFs positively correlates with the expression of vimentin, fibronectin and N-cadherin, all well-known mesenchymal markers. On the other hand, aberrant epigenetic silencing of TFs such as SOX1, KLF4, HIC1 and DACH1 through promoter hypermethylation has been reported in HCC, lung, gastric and urothelial cancer [130–133]. HOXA10, another epithelial-related TF, is silenced in breast cancer by CTCF insulator binding [134]. Several publications report that EMT reprogramming promotes epigenetic silencing of cell junction-related genes such as ITGA5, CDH11, CADM1 and OLFM4 [135–138]. Ectopic expression of these silenced genes decreased cell migration potential, highlighting their role in the EMT process. Post-translational modifications of membrane proteins may also contribute to cell adhesion. B3GNT7 is an O-glycosyltransferase that in CRC is downregulated by epigenetic silencing, and as in the cases listed above, its recovery decreases cells migration and invasion ability [139]. However, EMT is not a linear process and many different mechanisms are involved in its regulation. Thus, whereas ITGA2 is inhibited in some breast cancers by *miR-373* effect, *miR-373* silencing has been observed in lung cancer [140,141]. CDH1 is a cadherin protein involved in cell–cell adhesion. ZEB1 and ZEB2 are two transcriptional repressors of *CDH1*. These genes are regulated by *miR-200* microRNA family. Normal expression of *miR-200* may maintain *CDH1* expression, but aberrant promoter hypermethylation of *miR-200* results in a *CDH1* downregulation, which triggers cell migration [142]. In addition to cell detachment, another important step for cell invasion is extracellular matrix degradation, with activation

of proteases being a common feature in invasive front cells [143]. Epigenetic silencing of protease inhibitor proteins has also been reported. For instance, SPINT2 is often silenced in gastric cancer, ESCC and melanoma by DNA promoter hypermethylation [144–146]. MMP-9 is a well-characterized metalloprotease, which is overexpressed in different cancers. Its higher activity may be partially explained by RECK or KISS1 silencing due to promoter hypermethylation. Whereas RECK is an extracellular protein with a metalloprotease inhibitor domain [147], KISS1 seems to be involved in interfering with NF- κ B-mediated MMP-9 transcriptional activation [148]. *miR-145* targets MMP-11, and has been reported to be inactivated in different cancer malignancies. *miR-145* ectopic expression suppressed cell invasion and migration in renal cell carcinoma [149]. mTOR, a molecular sensor of cellular status that is also involved in metastasis, integrates several signals including PI3K/AKT, Wnt/ β -Catenin, Ras pathways and AMP/ATP ratio. Its activation is mainly produced by TSC1/TSC2 inhibition. mTOR can generate two different complexes, mTORC1 and mTORC2. mTORC1 is related to MMP-2 and MMP-9 activation, contributing to the invasiveness of cancer cells. mTORC1 is also related to anabolism, participating in protein synthesis and inhibiting autophagy. mTORC2 also seems to promote cell motility through activation of focal adhesion kinase (FAK) and Rho GTPases [150]. TSC1/TSC2, which partially contributes to mTOR regulation, is epigenetically silenced by metastasis associated 1 family, member 2 (MTA2) in association with EZH2, a component of polycomb repressor complex 2 [151]. PIP2 is a metabolite produced in the PI3K/AKT pathway, and plays an important role in cell motility. PIP2 participates in actin reorganization, becoming an interaction point for actin binding proteins. Actin reorganization is a necessary step for migration/invasion. PIP2 is hydrolysed by PLCD, a tumour suppressor gene methylated in cancer. MMP-7, another protease, also seems to be down-regulated by PLCD, thereby decreasing its activity [43]. *miR-345*, which is downregulated in non-small cell lung cancer through promoter hypermethylation [152], targets IRF1, a downstream factor of mTOR/AKT signalling involved in the EMT process [153]. Taken together, these data demonstrate the importance of epigenetic marks during EMT in modifying not only transcription factors but also cell adhesion and invasion genes.

7. Metabolic reprogramming

Metabolic reprogramming is one of the most universal features of cell transformation. Cancer cells tend to enhance glucose uptake through HIF1a stabilization, promoting a GLUT1 increase. In some cancers, HIF1a stabilization may be promoted by VHL promoter hypermethylation [154]. Paradoxically, this increase in glucose uptake is not translated into a huge increase at ATP levels, but instead the additional glucose is mostly metabolized by glycolysis. This phenomenon, known as the Warburg effect, is defined as the anaerobic use of glucose regardless of the presence of oxygen, conferring enormous advantages to cancer cells. Glycolysis results in lactate production, causing acidification of the extracellular space and activation of proteases involved in cell migration/invasion [155]. Enzymatic shift by alternative splicing of PKM gene between isoforms PKM1 or PKM2 promotes metabolite accumulation, activating the pentose phosphate pathway

(PPP) [156]. This pathway produces NADPH, an essential metabolite for glutathione and fatty acid synthesis, required for ROS protection and cell growth, respectively. The PPP also synthesizes ribonucleotides that are necessary for cell division [157]. Epigenetic silencing of certain genes may generate a glucose uptake increase. Lopez-Serra *et al.* [158] demonstrated that DERL3 targets GLUT1 in colorectal cells, leading to its proteasomal degradation. Ectopic DERL3 expression partially reverted the Warburg effect in HCT116 cells, decreasing the lactate production and increasing O₂ consumption, as well as decreasing metastasis in mice [158]. Tricarboxylic acid cycle is an anaplerotic mitochondrial pathway mainly involved in obtaining energy from acetyl-CoA. α -Ketoglutarate, a metabolite of this pathway, is necessary for TET and histone demethylase (HDM) activity. Mutations of the IDH1 and IDH2 enzymes (isocitrate dehydrogenases) and epigenetic inactivation of glutaminase (GS) can decrease α -ketoglutarate levels, promoting aberrant methylation marks on the epigenome [159–162]. Glucose uptake also increases serine glycine one-carbon metabolism (SGOC) pathway activity, connected by 3-phosphoglycerate dehydrogenase (PHGDH). This complex network connects the folate and methionine cycles. Folate is necessary for amino acid and nucleotide synthesis. Inhibition of this pathway, which is upregulated in cancer, has been used as a therapeutic strategy for several years due to its role in cell division [163]. Moreover, the methionine cycle is involved in SAM synthesis. SAM is a necessary substrate for DNMTs and histone methyltransferases (HMTs). Thus, this metabolite encompasses epigenetic modifications that are often related to gene silencing and metabolism. In some cancers, SAM synthesis is decreased by epigenetic silencing of SGOC genes. MAT1 and MTHFR protein levels are decreased in early pre-neoplastic rat livers by *miR-22* and *miR-29b* alterations and histone modifications, promoting a decrease in SAM synthesis, suggesting a rate-limiting substrate for epigenome maintenance [164]. The Met cycle is also connected with cysteine transsulfuration pathway. Two enzymes of this pathway, cystathionine beta synthase (CBS) and cysteine dioxygenase type 1 (CDO1), are often silenced by epigenetic inactivation in gastric and breast cancer, respectively [165,166]. Transsulfuration pathway products glutathione and taurine are redox controllers. These epigenetic alterations may lead to a decrease in redox-controlling molecules, facilitating tumourigenesis. Besides glutathione synthesis deregulation, antioxidant enzymes such as superoxide dismutase 2 (SOD2) and glutathione peroxidase (GP3) are frequently inactivated by promoter hypermethylation in lung and renal cancer, respectively [167,168]. Reviewed data suggest that epigenetic inactivation contributes to metabolic reprogramming, tumour progression through mutation accumulation by ROS, and dysregulation of epigenetic marks by rate-limiting substrates such as SAM and α -ketoglutarate. All of these metabolic alterations, acquired through epigenetic alterations, confer a selective advantage on cancer cells.

8. Tumour plasticity

The Sonic Hedgehog (Shh) pathway is aberrantly activated in different cancer types. This pathway can affect both tumour cells and stromal cells [169]. Within the tumour, it is important to highlight its importance in cancer stem cells (CSCs) [170]. It has been demonstrated that the Shh pathway is necessary for

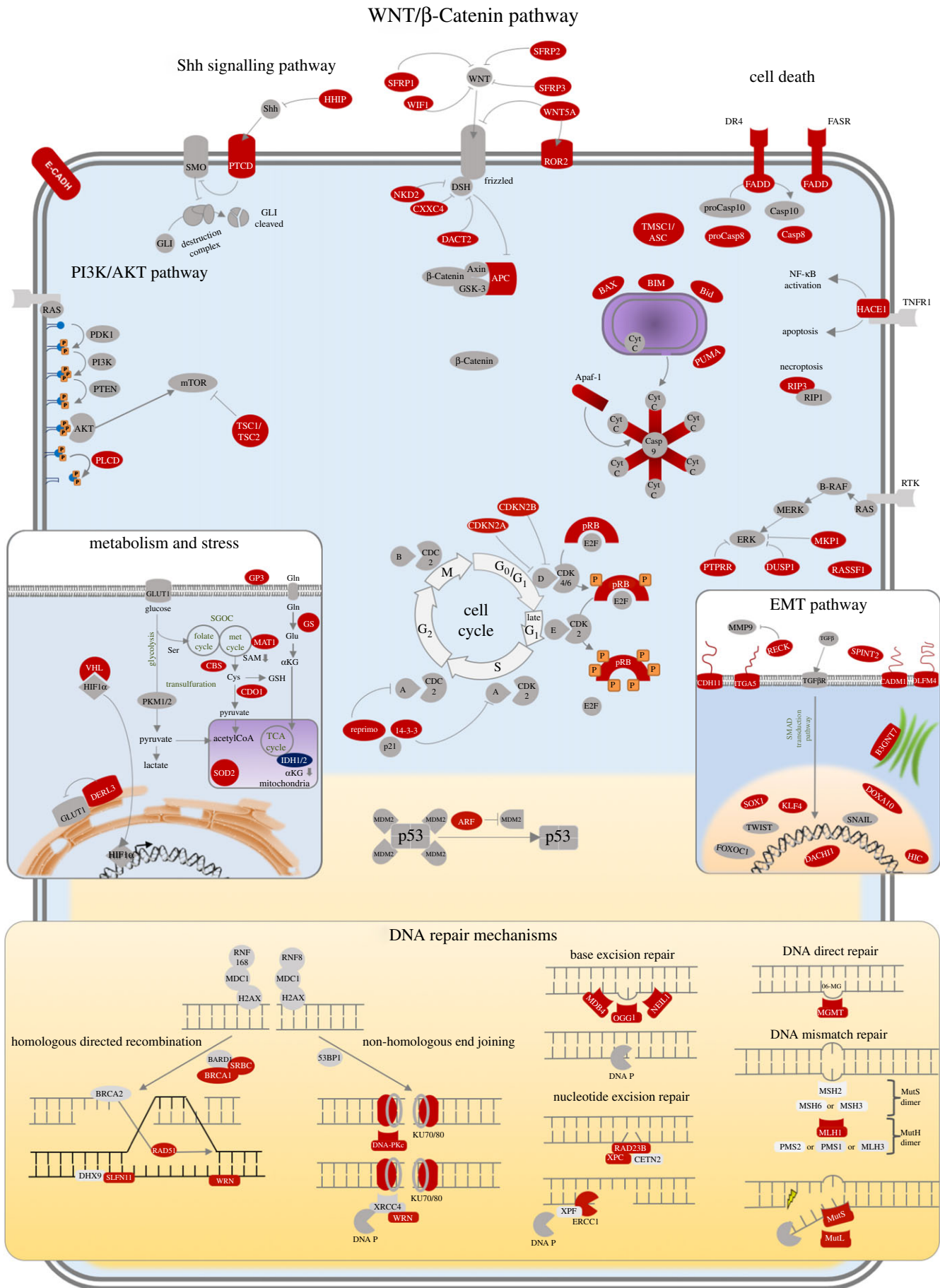


Figure 2. Molecular pathways altered by epigenetic inactivation in cancer. Represented pathways are altered in cancer. Red proteins correspond to epigenetically silenced genes.

Table 1. Coding and non-coding genes silenced in cancer.

gene	epigenetically inactivated	involved in	reference
14-3-3 σ	promoter hypermethylation	immortality	[70,71]
Apaf-1	promoter hypermethylation	immortality	[79]
APC	promoter hypermethylation	aberrant division	[57]
ATM	promoter hypermethylation	genomic instability	[100]
BAX	promoter hypermethylation	immortality	[75]
BIM	HDAC activity	immortality	[78]
BRCA1	promoter hypermethylation	genomic instability	[119]
CADM1	promoter hypermethylation	metastasis	[137]
Casp 8	promoter hypermethylation	immortality	[81]
CBS	promoter hypermethylation	metabolism and stress	[165]
CDH11	promoter hypermethylation	metastasis	[136]
CD01	promoter hypermethylation	metabolism and stress	[166]
CHK2	promoter hypermethylation	genomic instability	[101]
CXXC4	promoter hypermethylation	aberrant division	[61]
DACH1	promoter hypermethylation	metastasis	[133]
DACT2	promoter hypermethylation	aberrant division	[61]
DERL3	promoter hypermethylation	metabolism and stress	[158]
DNA-PKc	promoter hypermethylation	genomic instability	[127]
DR4	promoter hypermethylation	immortality	[80]
DUSP1	promoter hypermethylation	aberrant division	[37]
ERCC1	promoter hypermethylation	genomic instability	[111]
FADD	promoter hypermethylation	immortality	[85]
FASR	promoter hypermethylation	immortality	[83]
GP3	promoter hypermethylation	metabolism and stress	[168]
GS	promoter hypermethylation	metabolism and stress	[161]
HACE1	promoter hypermethylation	immortality	[88]
HHIP	promoter hypermethylation	tumour plasticity	[61]
HIC1	promoter hypermethylation	metastasis	[132]
HOXA10	CTCF binding	metastasis	[134]
ITGA2	miR-373	metastasis	[140]
ITGA5	promoter hypermethylation	metastasis	[135]
KISS1	promoter hypermethylation	metastasis	[148]
KLF4	promoter hypermethylation	metastasis	[131]
Ku80	promoter hypermethylation	genomic instability	[127]
Let-7a	promoter hypermethylation	aberrant division, tumour plasticity	[39,176]
MAT1	histone modification, miR	metabolism and stress	[164]
MBD4	promoter hypermethylation	genomic instability	[106]
MGMT	promoter hypermethylation	genomic instability	[22]
miR-124a	promoter hypermethylation	aberrant division	[27]
miR-129	promoter hypermethylation	aberrant division	[28]
miR-137	promoter hypermethylation	aberrant division	[29]
miR-145	promoter hypermethylation	metastasis	[149]
miR-200	promoter hypermethylation	metastasis	[142]
miR-214	promoter hypermethylation	aberrant division	[32]
miR-345	promoter hypermethylation	aberrant division	[33]
miR-34a	promoter hypermethylation	aberrant division	[31]

(Continued.)

Table 1. (Continued.)

gene	epigenetically inactivated	involved in	reference
miR-373	promoter hypermethylation	metastasis	[141]
miR-375	promoter hypermethylation	aberrant division	[41]
miR-7	promoter hypermethylation	immortality	[77]
miR-9	promoter hypermethylation	aberrant division	[35]
miR1-1	promoter hypermethylation	aberrant division	[30]
MLH1	promoter hypermethylation	genomic instability	[113]
MTHFR	histone modification, miR	metabolism and stress	[164]
NDRG2	promoter hypermethylation	aberrant division	[53]
NEIL1	promoter hypermethylation	genomic instability	[107]
NKD2	promoter hypermethylation	aberrant division	[60]
Notch L	promoter hypermethylation	tumour plasticity	[177]
NotchR	promoter hypermethylation	tumour plasticity	[177]
NUDT16	promoter hypermethylation	aberrant division	[64]
OGG1	promoter hypermethylation	genomic instability	[105]
OLFM4	promoter hypermethylation	metastasis	[138]
P14/ARF	promoter hypermethylation	immortality	[66]
P15INK4b/CDKN2B	promoter hypermethylation	aberrant division	[18]
P16INK4a/CDKN2A	promoter hypermethylation, HDAC activity	aberrant division	[18,23]
PLCD1	promoter hypermethylation	aberrant division	[43]
PTCD	promoter hypermethylation	tumour plasticity	[172]
PTPRR	promoter hypermethylation	aberrant division	[36]
RAD23B	promoter hypemethylation	genomic instability	[110]
RAD51	histone methylation	genomic instability	[116]
RAD51B	promoter hypermethylation	genomic instability	[118]
RASSF1	promoter hypermethylation	aberrant division	[46]
RB1	promoter hypermethylation	aberrant division	[24]
RECK	promoter hypermethylation	metastasis	[147]
Reprimo	promoter hypermethylation	immortality	[72]
RIPK3	promoter hypermethylation	immortality	[89]
ROR2	promoter hypermethylation	aberrant division	[62]
SFRP1	promoter hypermethylation	aberrant division	[50]
SFRP2	promoter hypermethylation	aberrant division	[51]
SFRP3	promoter hypermethylation	aberrant division	[52]
SLFN11	promoter hypermethylation	genomic instability	[123]
SOD2	promoter hypermethylation	metabolism and stress	[167]
SOX1	promoter hypermethylation	metastasis	[130]
SPINT2	promoter hypermethylation	metastasis	[146]
SRBC	promoter hypermethylation	genomic instability	[121]
TFPI-2	LCT activation	genomic instability	[95]
TMS1/ASC	promoter hypermethylation	immortality	[86,87]
TP53TG1	promoter hypermethylation	immortality	[73]
TSC1/TSC2	MTA2/EZH2	metastasis	[151]
VHL	promoter hypermethylation	metabolism and stress	[154]
WIF1	promoter hypermethylation	aberrant division	[54]
WNT5A	promoter hypermethylation	aberrant division	[55]

(Continued.)

Table 1. (Continued.)

gene	epigenetically inactivated	involved in	reference
WNT7A	promoter hypermethylation	aberrant division	[56]
WRN	promoter hypermethylation	genomic instability	[122]
XPC	promoter hypermethylation	genomic instability	[109]
XRCC3	promoter hypermethylation	genomic instability	[118]
ZIC1	promoter hypermethylation	tumour plasticity	[61]
ZIC4	promoter hypermethylation	tumour plasticity	[61]

renewal of CSCs [171]. In normal conditions, Shh ligand binds Patched (PTCD) transmembrane protein, allowing Smoothed (SMO) activation. HHIP is a Shh ligand antagonist that has been found silenced through promoter hypermethylation in HNSC [61]. PTCD can be epigenetically inactivated in cancer, promoting an Shh ligand-independent activity [172]. When SMO is activated, it inhibits GLI destruction complex, allowing its nucleus translocation. GLI activates the transcription of target genes involved in self renewal and EMT, such as SNAIL [173]. ZIC1 and ZIC4 are GLI antagonists, and they are also inhibited through promoter hypermethylation [61]. High-mobility-group proteins comprise a family of chromatin associated proteins that are involved in maintaining stem cell-like properties in CSCs. HMG2A is a member of this family, and has been associated with Wnt pathway activation [174] and EMT progression [175]. Overexpression of this gene in cancer cells in comparison with normal tissue can be partially explained because *Let-7a*, a microRNA that targets HMG2A, undergoes downregulation through promoter hypermethylation [176]. The Notch pathway is involved in cell–cell communication. Although its role in cancer is controversial due to its dual oncogenic and tumour suppressor properties, it has been demonstrated that epigenetics is involved in its regulation. Notch receptors recognize a number of ligands. This binding promotes migration of Notch intracellular domain into the nucleus and modification of gene expression. Four Notch receptors and five receptor ligands have been discovered. All receptors and four of the five ligands have been found methylated in different types of cancer [177].

9. Tumour suppressor genes epigenetic recovery in the fight against cancer

Cancer is not a single disease. There is no master alteration common to all cancer types. Epigenetic alterations may confer a selective advantage upon tumour cells, due to their potential reversibility. Many studies have been focused on cancer treatment by reversing these epigenetic events. DNA methylation inhibiting drugs were the first anti-epigenetic treatment, tested in leukaemias [178]. 5'-Aza-2'-deoxycytidine and decitabine are hypomethylating agents which can recover epigenetically silenced genes [179]. Alterations in histone modifications have also been therapeutic targets. Vorinostat was the first HDACi approved for clinical use, and several are currently undergoing clinical trials [180]. There also are pre-clinical trials with HATi, especially focused on Tip60 inhibition [181]. HMTs and HDMs have also been considered potential small molecule targets, including DOT1L, a HMT that has

been inhibited by different drugs in MLL (mixed lineage leukaemia) [182], and LSD1, which is a HDM studied as a target in AML [182]. Besides targeting writers and erasers, inhibition of readers is also an area of ongoing investigation. Several bromodomain inhibitors have been developed, especially against the BET family [183].

The major limitation of these treatments is their lack of specificity. Blocking the enzymes responsible for depositing, reading or removing epigenetic marks entails global genomic effects with counterproductive side effects. For instance, a hypomethylating agent could reactivate a hypermethylated tumour suppressor gene but may also promote the expression of an oncogene at the same time. In the near future, this limitation may be overcome by CRISPR-mediated epigenetic editing, allowing site-specific epigenetic modification. Indeed there are some interesting results in basic research, although they are still far from the clinic [184]. Despite the lack of widespread use of epigenetic drugs in cancer therapy, epigenetics has other clinical applications. DNA methylation, due its stability in comparison with mRNA, is used as a biomarker for diagnosis [185], cancer monitoring [186], cancer prediction [187], cancer prognosis [188] and treatment response [189].

10. Concluding remarks

'To defeat your enemy, you must know your enemy' [190]. This proverb, which derives from *The Art of War* by Sun-tzu, is applicable to one of the most important challenges of this century: to understand cancer at the molecular level. Since epigenetics was first described, many studies have contributed to decipher its implications in tumour transformation, particularly with regard to how the genes that are responsible for keeping cells under control are silenced. Tumour cells progressively acquire perturbations that allow them to divide without control, to adapt themselves to unfavourable conditions and even to abandon their niche and colonize other tissues (summarized in figure 2).

In this review, we have provided a global update on this knowledge, highlighting those coding and non-coding tumour suppressor genes whose epigenetic inactivation gives rise to proliferative advantages (listed in table 1), by rewiring the most important pathways by which cancer cells perpetuates themselves.

Data accessibility. This article has no additional data.

Authors' contributions. P.L.-A. wrote the manuscript and designed the figures. M.E. supervised and revised the manuscript and figures.

Competing interests. The authors have no relevant competing interests.

Funding. P.L.-A. was supported by Formación de Personal Investigador fellowship from the Spanish Ministry of Economy, Industry

References

1. Mukherjee S. 2011 *The emperor of all maladies*. New York, NY: Simon and Schuster.
2. Huebner RJ, Todaro GJ. 1969 Oncogenes of RNA tumor viruses as determinants of cancer. *Proc. Natl Acad. Sci. USA* **64**, 1087–1094. (doi:10.1073/pnas.64.3.1087)
3. Aaronson S, Todaro S. 1969 Human diploid cell transformation by DNA extracted from the tumor virus SV40. *Science* **166**, 390–391. (doi:10.1126/science.166.3903.390)
4. Knudson AG. 1971 Mutation and cancer: statistical study of retinoblastoma. *Proc. Natl Acad. Sci. USA* **68**, 820–823. (doi:10.1073/pnas.68.4.820)
5. Waddington CH. 1939 Preliminary notes on the development of the wings in normal and mutant strains of *Drosophila*. *Proc. Natl Acad. Sci. USA* **25**, 299–307. (doi:10.1073/pnas.25.7.299)
6. Esteller M. 2008 Epigenetics in cancer. *N. Engl. J. Med.* **358**, 1148–1159. (doi:10.1056/NEJMr072067)
7. Feinberg AP, Vogelstein B. 1983 Hypomethylation distinguishes genes of some human cancers from their normal counterparts. *Nature* **301**, 89–92. (doi:10.1038/301089a0)
8. Feinberg AP, Vogelstein B. 1983 Hypomethylation of ras oncogenes in primary human cancers. *Biochem. Biophys. Res. Commun.* **111**, 47–54. (doi:10.1016/S0006-291X(83)80115-6)
9. Greger V, Passarge E, Höpping W, Messmer E, Horsthemke B. 1989 Epigenetic changes may contribute to the formation and spontaneous regression of retinoblastoma. *Hum. Genet.* **83**, 155–158. (doi:10.1007/BF00286709)
10. Poh WJ, Wee CPP, Gao Z. 2016 DNA methyltransferase activity assays: advances and challenges. *Theranostics* **6**, 369–391. (doi:10.7150/thno.13438)
11. Huang Y, Rao A. 2014 Connections between TET proteins and aberrant DNA modification in cancer. *Trends Genet.* **30**, 464–474. (doi:10.1016/j.tig.2014.07.005)
12. Simó-Riudalbas L, Esteller M. 2015 Targeting the histone orthography of cancer: drugs for writers, erasers and readers. *Br. J. Pharmacol.* **172**, 2716–2732. (doi:10.1111/bph.12844)
13. Cantariño N, Douet J, Buschbeck M. 2013 MacroH2A—an epigenetic regulator of cancer. *Cancer Lett.* **336**, 247–252. (doi:10.1016/j.canlet.2013.03.022)
14. Gerić M, Gajski G, Garaj-Vrhovac V. 2014 γ -H2AX as a biomarker for DNA double-strand breaks in ecotoxicology. *Ecotoxicol. Environ. Saf.* **105**, 13–21. (doi:10.1016/j.ecoenv.2014.03.035)
15. Esteller M. 2011 Non-coding RNAs in human disease. *Nat. Rev. Genet.* **12**, 861–874. (doi:10.1038/nrg3074)
16. Fisher RP. 2016 Getting to S: CDK functions and targets on the path to cell-cycle commitment. *F1000Res* **5**, 2374. (doi:10.12688/f1000research.9463.1)
17. Liu F, Korc M. 2012 Cdk4/6 inhibition induces epithelial–mesenchymal transition and enhances invasiveness in pancreatic cancer cells. *Mol. Cancer Ther.* **11**, 2138–2148. (doi:10.1158/1535-7163.MCT-12-0562)
18. Krajnović M, Radoković M, Davidović R, Dimitrijević B, Krtolica K. 2013 Prognostic significance of epigenetic inactivation of p16, p15, MGMT and DAPK genes in follicular lymphoma. *Med. Oncol.* **30**, 441. (doi:10.1007/s12032-012-0441-3)
19. Peng D, Zhang H, Sun G. 2014 The relationship between P16 gene promoter methylation and gastric cancer: a meta-analysis based on Chinese patients. *J. Cancer Res. Ther.* **10**, 292–295. (doi:10.4103/0973-1482.151535)
20. El-Naggar AK, Lai S, Clayman G, Lee JK, Luna MA, Goepfert H, Batsakis JG. 1997 Methylation, a major mechanism of p16/CDKN2 gene inactivation in head and neck squamous carcinoma. *Am. J. Pathol.* **151**, 1767–1774.
21. Hinrichsen I, Kemp M, Peveling-Oberhag J, Passmann S, Plotz G, Zeuzem S, Brieger A. 2014 Promoter methylation of MLH1, PMS2, MSH2 and p16 is a phenomenon of advanced-stage HCCs. *PLoS ONE* **9**, e84453. (doi:10.1371/journal.pone.0084453)
22. Wang J, Sasco AJ, Fu C, Xue H, Guo G, Hua Z, Zhou Q, Jiang Q, Xu B. 2008 Aberrant DNA methylation of P16, MGMT, and hMLH1 genes in combination with MTHFR C677T genetic polymorphism in esophageal squamous cell carcinoma. *Cancer Epidemiol. Biomarkers Prev.* **17**, 118–125. (doi:10.1158/1055-9965.EPI-07-0733)
23. Kim JK *et al.* 2013 Targeted inactivation of HDAC2 restores p16INK4a activity and exerts antitumor effects on human gastric cancer. *Mol. Cancer Res.* **11**, 62–73. (doi:10.1158/1541-7786.MCR-12-0332)
24. Chinnam M, Goodrich DW. 2011 RB1, development, and cancer. *Curr. Top. Dev. Biol.* **94**, 129–169. (doi:10.1016/B978-0-12-380916-2.00005-X)
25. Quiñonez-Silva G, Dávalos-Salas M, Recillas-Targa F, Ostrosky-Wegman P, Aranda DA, Benítez-Bribiesca L. 2016 Monoallelic germline methylation and sequence variant in the promoter of the RB1 gene: a possible constitutive epimutation in hereditary retinoblastoma. *Clin. Epigenetics* **8**, 1. (doi:10.1186/s13148-015-0167-0)
26. McCormick TM, Canedo NHS, Furtado YL, Silveira FA, de Lima RJ, Rosman ADF, Almeida Filho GL, Carvalho MG. 2015 Association between human papillomavirus and Epstein-Barr virus DNA and gene promoter methylation of RB1 and CDH1 in the cervical lesions: a transversal study. *Diagn. Pathol.* **10**, 59. (doi:10.1186/s13000-015-0283-3)
27. Lujambio A *et al.* 2007 Genetic unmasking of an epigenetically silenced microRNA in human cancer cells. *Cancer Res.* **67**, 1424–1429. (doi:10.1158/0008-5472.CAN-06-4218)
28. Yu X, Song H, Xia T, Han S, Xiao B, Luo L, Xi Y, Guo J. 2013 Growth inhibitory effects of three miR-129 family members on gastric cancer. *Gene* **532**, 87–93. (doi:10.1016/j.gene.2013.09.048)
29. Zhu X, Li Y, Shen H, Li H, Long L, Hui L, Xu W. 2013 miR-137 inhibits the proliferation of lung cancer cells by targeting Cdc42 and Cdk6. *FEBS Lett.* **587**, 73–81. (doi:10.1016/j.febslet.2012.11.004)
30. Reid JF *et al.* 2012 miRNA profiling in colorectal cancer highlights miR-1 involvement in MET-dependent proliferation. *Mol. Cancer Res.* **10**, 504–515. (doi:10.1158/1541-7786.MCR-11-0342)
31. Li N, Fu H, Tie Y, Hu Z, Kong W, Wu Y, Zheng X. 2009 miR-34a inhibits migration and invasion by down-regulation of c-Met expression in human hepatocellular carcinoma cells. *Cancer Lett.* **275**, 44–53. (doi:10.1016/j.canlet.2008.09.035)
32. Das F, Dey N, Bera A, Kasinath BS, Ghosh-Choudhury N, Choudhury GG. 2016 MicroRNA-214 reduces Insulin-like Growth Factor-1 (IGF-1) receptor expression and downstream mTORC1 signaling in renal carcinoma cells. *J. Biol. Chem.* **291**, 14 662–14 676. (doi:10.1074/jbc.M115.694331)
33. Kong KL, Kwong DLW, Chan TH-M, Law SY-K, Chen L, Li Y, Qin Y-R, Guan X-Y. 2012 MicroRNA-375 inhibits tumour growth and metastasis in oesophageal squamous cell carcinoma through repressing insulin-like growth factor 1 receptor. *Gut* **61**, 33–42. (doi:10.1136/gutjnl-2011-300178)
34. Misiewicz-Krzeminska I *et al.* 2013 Restoration of microRNA-214 expression reduces growth of myeloma cells through positive regulation of P53 and inhibition of DNA replication. *Haematologica* **98**, 640–648. (doi:10.3324/haematol.2012.070011)
35. Rodriguez-Otero P *et al.* 2011 Dereglulation of FGFR1 and CDK6 oncogenic pathways in acute lymphoblastic leukaemia harbouring epigenetic modifications of the MIR9 family. *Br. J. Haematol.* **155**, 73–83. (doi:10.1111/j.1365-2141.2011.08812.x)
36. Su P-H *et al.* 2013 Epigenetic silencing of PTPRR activates MAPK signaling, promotes metastasis and serves as a biomarker of invasive cervical cancer. *Oncogene* **32**, 15–26. (doi:10.1038/onc.2012.29)
37. Khor GH, Froemming GRA, Zain RB, Abraham MT, Omar E, Tan SK, Tan AC, Vincent-Chong VK, Thong KL. 2013 DNA methylation profiling revealed promoter hypermethylation-induced silencing of p16, DDAH2 and DUSP1 in primary oral squamous cell carcinoma. *Int. J. Med. Sci.* **10**, 1727–1739. (doi:10.7150/ijms.6884)

38. Chen F-M, Chang H-W, Yang S-F, Huang Y-F, Nien P-Y, Yeh Y-T, Hou M-F. 2012 The mitogen-activated protein kinase phosphatase-1 (MKP-1) gene is a potential methylation biomarker for malignancy of breast cancer. *Exp. Mol. Med.* **44**, 356–362. (doi:10.3858/emm.2012.44.5.040)
39. Johnson SM *et al.* 2005 RAS is regulated by the let-7 microRNA family. *Cell* **120**, 635–647. (doi:10.1016/j.cell.2005.01.014)
40. Yu C-C *et al.* 2011 MicroRNA let-7a represses chemoresistance and tumorigenicity in head and neck cancer via stem-like properties ablation. *Oral Oncol.* **47**, 202–210. (doi:10.1016/j.oraloncology.2010.12.001)
41. Li X, Lin R, Li J. 2011 Epigenetic silencing of microRNA-375 regulates PDK1 expression in esophageal cancer. *Dig. Dis. Sci.* **56**, 2849–2856. (doi:10.1007/s10620-011-1711-1)
42. Carnero A. 2010 The PKB/AKT pathway in cancer. *Curr. Pharm. Des.* **16**, 34–44. (doi:10.2174/138161210789941865)
43. Song J-J, Liu Q, Li Y, Yang Z-S, Yang L, Xiang T-X, Ren G-S, Chen J-B. 2012 Epigenetic inactivation of PLCD1 in chronic myeloid leukemia. *Int. J. Mol. Med.* **30**, 179–184. (doi:10.3892/ijmm.2012.970)
44. Hu X-T *et al.* 2009 Phospholipase C delta 1 is a novel 3p22.3 tumor suppressor involved in cytoskeleton organization, with its epigenetic silencing correlated with high-stage gastric cancer. *Oncogene* **28**, 2466–2475. (doi:10.1038/onc.2009.92)
45. Xiang T, Li L, Fan Y, Jiang Y, Ying Y, Putti TC, Tao Q, Ren G. 2010 PLCD1 is a functional tumor suppressor inducing G(2)/M arrest and frequently methylated in breast cancer. *Cancer Biol. Ther.* **10**, 520–527. (doi:10.4161/cbt.10.5.12726)
46. Hesson LB, Cooper WN, Latif F. 2007 The role of RASSF1A methylation in cancer. *Dis. Markers* **23**, 73–87. (doi:10.1155/2007/291538)
47. Dammann R, Schagdarsurengin U, Seidel C, Strunnikova M, Rastetter M, Baier K, Pfeifer GP. 2005 The tumor suppressor RASSF1A in human carcinogenesis: an update. *Histol. Histopathol.* **20**, 645–663. (doi:10.14670/HH-20.645)
48. Fenton SL *et al.* 2004 Identification of the E1A-regulated transcription factor p120 E4F as an interacting partner of the RASSF1A candidate tumor suppressor gene. *Cancer Res.* **64**, 102–107. (doi:10.1158/0008-5472.CAN-03-2622)
49. Baron R, Kneissel M. 2013 WNT signaling in bone homeostasis and disease: from human mutations to treatments. *Nat. Med.* **19**, 179–192. (doi:10.1038/nm.3074)
50. Meng Y, Wang Q-G, Wang J-X, Zhu S-T, Jiao Y, Li P, Zhang S-T. 2011 Epigenetic inactivation of the SFRP1 gene in esophageal squamous cell carcinoma. *Dig. Dis. Sci.* **56**, 3195–3203. (doi:10.1007/s10620-011-1734-7)
51. Hao X-W, Zhu S-T, He Y-L, Li P, Wang Y-J, Zhang S-T. 2012 Epigenetic inactivation of secreted frizzled-related protein 2 in esophageal squamous cell carcinoma. *World J. Gastroenterol.* **18**, 532–540. (doi:10.3748/wjg.v18.i6.532)
52. Schlenz M *et al.* In press. Epigenetic loss of putative tumor suppressor SFRP3 correlates with poor prognosis of lung adenocarcinoma patients. *Epigenetics*. (doi:10.1080/15592294.2016.1229730)
53. Yamamura A *et al.* 2017 NDRG2, suppressed expression associates with poor prognosis in pancreatic cancer, is hypermethylated in the second promoter in human gastrointestinal cancers. *Biochem. Biophys. Res. Commun.* **484**, 138–143. (doi:10.1016/j.bbrc.2017.01.055)
54. Yang S-H, Li S-L, Dong Z-M, Kan Q-C. 2012 Epigenetic inactivation of Wnt inhibitory factor-1 in human esophageal squamous cell carcinoma. *Oncol. Res.* **20**, 123–130. (doi:10.3727/096504012X13477145153039)
55. Li J, Ying J, Fan Y, Wu L, Ying Y, Chan AT. C., Srivastava G, Tao Q. 2010 WNT5A antagonizes WNT/ β -catenin signaling and is frequently silenced by promoter CpG methylation in esophageal squamous cell carcinoma. *Cancer Biol. Ther.* **10**, 617–624. (doi:10.4161/cbt.10.6.12609)
56. Kim T-H, Moon J-Y, Kim S-H, Paik SS, Yoon HJ, Shin DH, Park SS, Sohn JW. 2015 Clinical significance of aberrant Wnt7a promoter methylation in human non-small cell lung cancer in Koreans. *J. Korean Med. Sci.* **30**, 155–161. (doi:10.3346/jkms.2015.30.2.155)
57. Michailidi C, Theocharis S, Tsourouflis G, Pletsas V, Kouraklis G, Patsouris E, Papavassiliou AG, Troungos C. 2015 Expression and promoter methylation status of hMLH1, MGMT, APC, and CDH1 genes in patients with colon adenocarcinoma. *Exp. Biol. Med.* **240**, 1599–1605. (doi:10.1177/1535370215583800)
58. Swellam M, Abdelmaksoud MDE, Sayed Mahmoud M, Ramadan A, Abdel-Moneem W, Hefny MM. 2015 Aberrant methylation of APC and RAR β 2 genes in breast cancer patients. *IUBMB Life* **67**, 61–68. (doi:10.1002/iub.1346)
59. Ginesta MM *et al.* 2016 APC promoter is frequently methylated in pancreatic juice of patients with pancreatic carcinomas or periampullary tumors. *Oncol. Lett.* **12**, 2210–2216. (doi:10.3892/ol.2016.4868)
60. Cao B, Yang W, Jin Y, Zhang M, He T, Zhan Q, Herman JG, Zhong G, Guo M. 2016 Silencing NKD2 by promoter region hypermethylation promotes esophageal cancer progression by activating Wnt signaling. *J. Thorac. Oncol.* **11**, 1912–1926. (doi:10.1016/j.jtho.2016.06.015)
61. Paluszczak J, Wiśniewska D, Kostrzewska-Poczekaj M, Kiwerska K, Grénman R, Mielcarek-Kuchta D, Jarmuż-Szymczak M. 2017 Prognostic significance of the methylation of Wnt pathway antagonists-CXCC4, DACT2, and the inhibitors of sonic hedgehog signaling-ZIC1, ZIC4, and HHIP in head and neck squamous cell carcinomas. *Clin. Oral Investig.* **21**, 1777. (doi:10.1007/s00784-016-1946-5)
62. Ma SSQ, Srivastava S, Llamas E, Hawkins NJ, Hesson LB, Ward RL, Ford CE. 2016 ROR2 is epigenetically inactivated in the early stages of colorectal neoplasia and is associated with proliferation and migration. *BMC Cancer* **16**, 508. (doi:10.1186/s12885-016-2576-7)
63. Kalkat M, De Melo J, Hickman KA, Lourenco C, Redel C, Resetca D, Tamachi A, Tu WB, Penn LZ. 2017 MYC deregulation in primary human cancers. *Genes* **8**, 151. (doi:10.3390/genes8060151)
64. Anadón C *et al.* 2017 Epigenetic loss of the RNA decapping enzyme NUDT16 mediates C-MYC activation in T-cell acute lymphoblastic leukemia. *Leukemia* **31**, 1622–1625. (doi:10.1038/leu.2017.99)
65. Kaminsky VO, Surova OV, Vaculova A, Zhivotovsky B. 2011 Combined inhibition of DNA methyltransferase and histone deacetylase restores caspase-8 expression and sensitizes SCLC cells to TRAIL. *Carcinogenesis* **32**, 1450–1458. (doi:10.1093/carcin/bgr135)
66. Delmonico L *et al.* 2015 CDKN2A (p14(ARF)/p16(INK4a)) and ATM promoter methylation in patients with impalpable breast lesions. *Hum. Pathol.* **46**, 1540–1547. (doi:10.1016/j.humpath.2015.06.016)
67. Verdoodt B, Sommerer F, Palisaar R-J, Noldus J, Vogt M, Nambiar S, Tannapfel A, Mirmohammadsadegh A, Neid M. 2011 Inverse association of p16 INK4a and p14 ARF methylation of the CDKN2a locus in different Gleason scores of prostate cancer. *Nature* **14**, 295–301. (doi:10.1038/pcan.2011.45)
68. Davidović R, Sopta J, Mandušić V, Krajinović M, Stanojević M, Tulić G, Dimitrijević B. 2013 p14(ARF) methylation is a common event in the pathogenesis and progression of myxoid and pleomorphic liposarcoma. *Med. Oncol.* **30**, 682. (doi:10.1007/s12032-013-0682-9)
69. Wang Y-J *et al.* 2014 Epigenetic changes of TIMP-3, GSTP-1 and 14-3-3 sigma genes as indication of status of chronic inflammation and cancer. *Int. J. Biol. Markers* **29**, e208-14. (doi:10.5301/ijbm.5000104)
70. Zurita M, Lara PC, del Moral R, Torres B, Linares-Fernández JL, Arrabal SR, Martínez-Galán J, Oliver FJ, Ruiz de Almodóvar JM. 2010 Hypermethylated 14-3-3-sigma and ESR1 gene promoters in serum as candidate biomarkers for the diagnosis and treatment efficacy of breast cancer metastasis. *BMC Cancer* **10**, 217. (doi:10.1186/1471-2407-10-217)
71. Yi B *et al.* 2009 Inactivation of 14-3-3 sigma by promoter methylation correlates with metastasis in nasopharyngeal carcinoma. *J. Cell. Biochem.* **106**, 858–866. (doi:10.1002/jcb.22051)
72. Liu L, Yang X. 2015 Implication of reprimo and hMLH1 gene methylation in early diagnosis of gastric carcinoma. *Int. J. Clin. Exp. Pathol.* **8**, 14977–14982.
73. Díaz-Lagares A *et al.* 2016 Epigenetic inactivation of the p53-induced long noncoding RNA TP53 target 1 in human cancer. *Proc. Natl Acad. Sci. USA* **113**, E7535–E7544. (doi:10.1073/pnas.1608585113)
74. Ashkenazi A. 2008 Directing cancer cells to self-destruct with pro-apoptotic receptor agonists. *Nat. Rev. Drug Discov.* **7**, 1001–1012. (doi:10.1038/nrd2637)
75. Alipour M, Zargar SJ, Safarian S, Fouladdel S, Azizi E, Jafarholizadeh N. 2013 The study of DNA methylation of bax gene promoter in breast and

- colorectal carcinoma cell lines. *Iran J. Cancer Prev.* **6**, 59–64.
76. Hervouet E, Cheray M, Vallette FM, Cartron P-F. 2013 DNA methylation and apoptosis resistance in cancer cells. *Cells* **2**, 545–573. (doi:10.3390/cells2030545)
 77. Xiong S. 2011 MicroRNA-7 inhibits the growth of human non-small cell lung cancer A549 cells through targeting BCL-2. *Int. J. Biol. Sci.* **7**, 805–814. (doi:10.7150/ijbs.7.805)
 78. Piazza R *et al.* 2013 Epigenetic silencing of the proapoptotic gene BIM in anaplastic large cell lymphoma through an MeCP2/SIN3a deacetylating complex. *Neoplasia* **15**, 511–522. (doi:10.1593/neo.121784)
 79. Zhu X, Yi F, Chen P, Chen L, Zhang X, Cao C, Tan W. 2015 5-Aza-2'-deoxycytidine and CDDP synergistically induce apoptosis in renal carcinoma cells via enhancing the APAF-1 activity. *Clin. Lab.* **61**, 1821–1830. (doi:10.7754/Clin.Lab.2015.150429)
 80. Lee KH, Lim SW, Kim HG, Kim DY, Ryu SY, Joo JK, Kim JC, Lee JH. 2009 Lack of death receptor 4 (DR4) expression through gene promoter methylation in gastric carcinoma. *Langenbecks Arch. Surg.* **394**, 661–670. (doi:10.1007/s00423-009-0484-x)
 81. Skiriute D, Vaitkiene P, Saferis V, Asmoniene V, Skauminas K, Deltuva VP, Tamasauskas A. 2012 MGMT, GATA6, CD81, DR4, and CASP8 gene promoter methylation in glioblastoma. *BMC Cancer* **12**, 218. (doi:10.1186/1471-2407-12-218)
 82. Butler LM, Dobrovic A, Bianco T, Cowled PA. 2000 Promoter region methylation does not account for the frequent loss of expression of the Fas gene in colorectal carcinoma. *Br. J. Cancer* **82**, 131–135. (doi:10.1054/bjoc.1999.0889)
 83. Petak I, Danam RP, Tillman DM, Vernes R, Howell SR, Berdzi L, Kopper L, Brent TP, Houghton JA. 2003 Hypermethylation of the gene promoter and enhancer region can regulate Fas expression and sensitivity in colon carcinoma. *Cell Death Differ.* **10**, 211–217. (doi:10.1038/sj.cdd.4401132)
 84. Watson CJ *et al.* 2012 Identification of a methylation hotspot in the death receptor Fas/CD95 in bladder cancer. *Int. J. Oncol.* **40**, 645–654. (doi:10.3892/ijco.2011.1250)
 85. Saberi E, Kordi-Tamandani D-M, Jamali S, Rigi-Ladiz M-A. 2014 Analysis of methylation and mRNA expression status of FADD and FAS genes in patients with oral squamous cell carcinoma. *Med. Oral Patol. Oral Cir. Bucal.* **19**, e562–e568. (doi:10.4317/medoral.19805)
 86. Terasawa K *et al.* 2004 Epigenetic inactivation of TMS1/ASC in ovarian cancer. *Clin. Cancer Res.* **10**, 2000–2006. (doi:10.1158/1078-0432.CCR-0932-03)
 87. Stone AR, Bobo W, Brat DJ, Devi NS, Van Meir EG, Vertino PM. 2004 Aberrant methylation and down-regulation of TMS1/ASC in human glioblastoma. *Am. J. Pathol.* **165**, 1151–1161. (doi:10.1016/S0002-9440(10)63376-7)
 88. Sakata M *et al.* 2013 Methylation of the HACE1 gene is frequently detected in hepatocellular carcinoma. *HepatoGastroenterology* **60**, 781–783. (doi:10.5754/hge10439)
 89. Koo G-B *et al.* 2015 Methylation-dependent loss of RIP3 expression in cancer represses programmed necrosis in response to chemotherapeutics. *Cell Res.* **25**, 707–725. (doi:10.1038/cr.2015.56)
 90. Ferreira HJ *et al.* 2016 DNMT3A mutations mediate the epigenetic reactivation of the leukemogenic factor MEI51 in acute myeloid leukemia. *Oncogene* **35**, 3079–3082. (doi:10.1038/onc.2015.359)
 91. Ahn J-S *et al.* 2016 DNMT3A R882 mutation with FLT3-ITD positivity is an extremely poor prognostic factor in patients with normal-karyotype acute myeloid leukemia after allogeneic hematopoietic cell transplantation. *Biol. Blood Marrow Transplant.* **22**, 61–70. (doi:10.1016/j.bbmt.2015.07.030)
 92. Liu W-J, Tan X-H, Luo X-P, Guo B-P, Wei Z-J, Ke Q, He S, Cen H. 2014 Prognostic significance of Tet methylcytosine dioxygenase 2 (TET2) gene mutations in adult patients with acute myeloid leukemia: a meta-analysis. *Leuk. Lymphoma* **55**, 2691–2698. (doi:10.3109/10428194.2014.893308)
 93. Eden A, Gaudet F, Waghmare A, Jaenisch R. 2003 Chromosomal instability and tumors promoted by DNA hypomethylation. *Science* **300**, 455. (doi:10.1126/science.1083557)
 94. Tufarelli C, Cruickshanks HA, Meehan RR. 2013 LINE-1 activation and epigenetic silencing of suppressor genes in cancer: causally related events? *Mob. Genet. Elements* **3**, e26832. (doi:10.4161/mge.26832)
 95. Cruickshanks HA, Vafadar-Isfahani N, Dunican DS, Lee A, Sproul D, Lund JN, Meehan RR, Tufarelli C. 2013 Expression of a large LINE-1-driven antisense RNA is linked to epigenetic silencing of the metastasis suppressor gene TP53 in cancer. *Nucleic Acids Res.* **41**, 6857–6869. (doi:10.1093/nar/gkt438)
 96. Jeggo PA, Pearl LH, Carr AM. 2016 DNA repair, genome stability and cancer: a historical perspective. *Nat. Rev. Cancer* **16**, 35–42. (doi:10.1038/nrc.2015.4)
 97. Roos WP, Kaina B. 2013 DNA damage-induced cell death: from specific DNA lesions to the DNA damage response and apoptosis. *Cancer Lett.* **332**, 237–248. (doi:10.1016/j.canlet.2012.01.007)
 98. Gavande NS, VanderVere-Carozza PS, Hinshaw HD, Jalal SI, Sears CR, Pawelczak KS, Turchi JJ. 2016 DNA repair targeted therapy: the past or future of cancer treatment? *Pharmacol. Ther.* **160**, 65–83. (doi:10.1016/j.pharmthera.2016.02.003)
 99. Jackson SP, Bartek J. 2009 The DNA-damage response in human biology and disease. *Nature* **461**, 1071–1078. (doi:10.1038/nature08467)
 100. Treilleux I, Chapot B, Goddard S, Pisani P, Angèle S, Hall J. 2007 The molecular causes of low ATM protein expression in breast carcinoma; promoter methylation and levels of the catalytic subunit of DNA-dependent protein kinase. *Histopathology* **51**, 63–69. (doi:10.1111/j.1365-2559.2007.02726.x)
 101. Wang H *et al.* 2010 Chk2 down-regulation by promoter hypermethylation in human bulk gliomas. *Life Sci.* **86**, 185–191. (doi:10.1016/j.lfs.2009.11.023)
 102. Sartore-Bianchi A *et al.* 2017 Digital PCR assessment of MGMT promoter methylation coupled with reduced protein expression optimises prediction of response to alkylating agents in metastatic colorectal cancer patients. *Eur. J. Cancer* **71**, 43–50. (doi:10.1016/j.ejca.2016.10.032)
 103. Brigliadori G, Foca F, Dall'Agata M, Rengucci C, Melegari E, Cerasoli S, Amadori D, Calistri D, Faedi M. 2016 Defining the cutoff value of MGMT gene promoter methylation and its predictive capacity in glioblastoma. *J. Neurooncol.* **128**, 333–339. (doi:10.1007/s11060-016-2116-y)
 104. Yoon RG, Kim HS, Paik W, Shim WH, Kim SJ, Kim JH. 2017 Different diagnostic values of imaging parameters to predict pseudoprogression in glioblastoma subgroups stratified by MGMT promoter methylation. *Eur. Radiol.* **27**, 255–266. (doi:10.1007/s00330-016-4346-y)
 105. Singh KP, Treas J, Tyagi T, Gao W. 2012 DNA demethylation by 5-aza-2-deoxycytidine treatment abrogates 17 beta-estradiol-induced cell growth and restores expression of DNA repair genes in human breast cancer cells. *Cancer Lett.* **316**, 62–69. (doi:10.1016/j.canlet.2011.10.022)
 106. Howard JH *et al.* 2009 Epigenetic downregulation of the DNA repair gene MED1/MBD4 in colorectal and ovarian cancer. *Cancer Biol. Ther.* **8**, 94–100. (doi:10.4161/cbt.8.1.7469)
 107. Chaisaingmongkol J *et al.* 2012 Epigenetic screen of human DNA repair genes identifies aberrant promoter methylation of NEIL1 in head and neck squamous cell carcinoma. *Oncogene* **31**, 5108–5116. (doi:10.1038/onc.2011.660)
 108. Schärer OD. 2013 Nucleotide excision repair in eukaryotes. *Cold Spring Harb. Perspect. Biol.* **5**, a012609. (doi:10.1101/cshperspect.a012609)
 109. Wu Y-H, Tsai Chang J-H, Cheng Y-W, Wu T-C, Chen C-Y, Lee H. 2007 *Xeroderma pigmentosum* group C gene expression is predominantly regulated by promoter hypermethylation and contributes to p53 mutation in lung cancers. *Oncogene* **26**, 4761–4773. (doi:10.1038/sj.onc.1210284)
 110. Peng B, Hodge DR, Thomas SB, Cherry JM, Munroe DJ, Pompeia C, Xiao W, Farrar WL. 2005 Epigenetic silencing of the human nucleotide excision repair gene, hHR23B, in interleukin-6-responsive multiple myeloma KAS-6/1 cells. *J. Biol. Chem.* **280**, 4182–4187. (doi:10.1074/jbc.M412566200)
 111. Lima LMC, de Souza LR, da Silva TF, Pereira CS, Guimarães ALS, de Paula AMB, de Andrade Carvalho H. 2012 DNA repair gene excision repair cross complementing-group 1 (ERCC1) in head and neck squamous cell carcinoma: analysis of methylation and polymorphism (G19007A), protein expression and association with epidemiological and clinicopathological factors. *Histopathology* **60**, 489–496. (doi:10.1111/j.1365-2559.2011.04062.x)
 112. Muro Y, Sugiura K, Mimori T, Akiyama M. 2015 DNA mismatch repair enzymes: genetic defects and autoimmunity. *Clin. Chim. Acta* **442**, 102–109. (doi:10.1016/j.cca.2015.01.014)

113. Kidambi TD, Blanco A, Van Ziffle J, Terdiman JP. 2016 Constitutional MLH1 methylation presenting with colonic polyposis syndrome and not Lynch syndrome. *Fam. Cancer* **15**, 275–280. (doi:10.1007/s10689-016-9868-6)
114. Lee J-H, Paull TT. 2004 Direct activation of the ATM protein kinase by the Mre11/Rad50/Nbs1 complex. *Science* **304**, 93–96. (doi:10.1126/science.1091496)
115. Lord CJ, Ashworth A. 2016 BRCAness revisited. *Nat. Rev. Cancer* **16**, 110–120. (doi:10.1038/nrc.2015.21)
116. Liu S, Li Y, Xu H, Wang K, Li N, Li J, Sun T, Xu Y. 2016 Increased expression of SET domain-containing proteins and decreased expression of Rad51 in different classes of renal cell carcinoma. *Biosci. Rep.* **36**, e00349. (doi:10.1042/BSR20160122)
117. Suwaki N, Klare K, Tarsounas M. 2011 RAD51 paralogs: roles in DNA damage signalling, recombinational repair and tumorigenesis. *Semin. Cell Dev. Biol.* **22**, 898–905. (doi:10.1016/j.semcdb.2011.07.019)
118. Rieke DT, Ochsenreither S, Klinghammer K, Seiwert TY, Klauschen F, Tinhofer I, Keilholz U. 2016 Methylation of RAD51B, XRCC3 and other homologous recombination genes is associated with expression of immune checkpoints and an inflammatory signature in squamous cell carcinoma of the head and neck, lung and cervix. *Oncotarget* **7**, 75 379–75 393. (doi:10.18632/oncotarget.12211)
119. Pathania S, Nguyen J, Hill SJ, Scully R, Adelmant GO, Marto JA, Feunteun J, Livingston DM. 2011 BRCA1 is required for postreplication repair after UV-induced DNA damage. *Mol. Cell* **44**, 235–251. (doi:10.1016/j.molcel.2011.09.002)
120. O'Driscoll M, Jeggo PA. 2006 The role of double-strand break repair—insights from human genetics. *Nat. Rev. Genet.* **7**, 45–54. (doi:10.1038/nrg1746)
121. Moutinho C *et al.* 2014 Epigenetic inactivation of the BRCA1 interactor SRBC and resistance to oxaliplatin in colorectal cancer. *J. Natl Cancer Inst.* **106**, djt322. (doi:10.1093/jnci/djt322)
122. Masuda K *et al.* 2012 Association of epigenetic inactivation of the WRN gene with anticancer drug sensitivity in cervical cancer cells. *Oncol. Rep.* **28**, 1146–1152. (doi:10.3892/or.2012.1912)
123. Nogales V *et al.* 2016 Epigenetic inactivation of the putative DNA/RNA helicase SLFN11 in human cancer confers resistance to platinum drugs. *Oncotarget* **7**, 3084–3097. (doi:10.18632/oncotarget.6413)
124. Mu Y, Lou J, Srivastava M, Zhao B, Feng X-H, Liu T, Chen J, Huang J. 2016 SLFN11 inhibits checkpoint maintenance and homologous recombination repair. *EMBO Rep.* **17**, 94–109. (doi:10.15252/embr.201540964)
125. Zimmermann M, de Lange T. 2014 53BP1: pro choice in DNA repair. *Trends Cell Biol.* **24**, 108–117. (doi:10.1016/j.tcb.2013.09.003)
126. Davis AJ, Chen DJ. 2013 DNA double strand break repair via non-homologous end-joining. *Transl. Cancer Res.* **2**, 130–143. (doi:10.3978/j.issn.2218-676X.2013.04.02)
127. Zhou C, Tang H, Yu J, Zhuang D, Zhang H. 2015 Blood-based DNA methylation of DNA repair genes in the non-homologous end-joining (NEJ) pathway in patient with glioma. *Int. J. Clin. Exp. Pathol.* **8**, 9463–9467.
128. Carmona FJ *et al.* 2014 A comprehensive DNA methylation profile of epithelial-to-mesenchymal transition. *Cancer Res.* **74**, 5608–5619. (doi:10.1158/0008-5472.CAN-13-3659)
129. Yang M-H, Chen C-L, Chau G-Y, Chiou S-H, Su C-W, Chou T-Y, Peng W-L, Wu J-C. 2009 Comprehensive analysis of the independent effect of twist and snail in promoting metastasis of hepatocellular carcinoma. *Hepatology* **50**, 1464–1474. (doi:10.1002/hep.23221)
130. Li N, Li S. 2015 Epigenetic inactivation of SOX1 promotes cell migration in lung cancer. *Tumour Biol.* **36**, 4603–4610. (doi:10.1007/s13277-015-3107-x)
131. Li H *et al.* 2014 Epigenetic inactivation of KLF4 is associated with urothelial cancer progression and early recurrence. *J. Urol.* **191**, 493–501. (doi:10.1016/j.juro.2013.08.087)
132. Li P, Liu X, Dong Z-M, Ling Z-Q. 2015 Epigenetic silencing of HIC1 promotes epithelial-mesenchymal transition and drives progression in esophageal squamous cell carcinoma. *Oncotarget* **6**, 38 151–38 165. (doi:10.18632/oncotarget.5832)
133. Yan W, Wu K, Herman JG, Brock MV, Zhou Y, Lu Y, Zhang Z, Yang Y, Guo M. 2014 Epigenetic silencing of DACH1 induces the invasion and metastasis of gastric cancer by activating TGF- β signalling. *J. Cell. Mol. Med.* **18**, 2499–2511. (doi:10.1111/jcmm.12325)
134. Mustafa M, Lee J-Y, Kim MH. 2015 CTCF negatively regulates HOXA10 expression in breast cancer cells. *Biochem. Biophys. Res. Commun.* **467**, 828–834. (doi:10.1016/j.bbrc.2015.10.058)
135. Fang Z, Yao W, Xiong Y, Zhang J, Liu L, Li J, Zhang C, Wan J. 2010 Functional elucidation and methylation-mediated downregulation of ITGA5 gene in breast cancer cell line MDA-MB-468. *J. Cell. Biochem.* **110**, 1130–1141. (doi:10.1002/jcb.22626)
136. Lin Y-L, Gui S-L, Ma J-G. 2015 Aberrant methylation of CDH11 predicts a poor outcome for patients with bladder cancer. *Oncol. Lett.* **10**, 647–652. (doi:10.3892/ol.2015.3337)
137. van Baars R *et al.* 2016 CADM1 and MAL methylation status in cervical scrapes is representative of the most severe underlying lesion in women with multiple cervical biopsies. *Int. J. Cancer* **138**, 463–471. (doi:10.1002/ijc.29706)
138. Guo L-L, He Z-C, Yang C-Q, Qiao P-T, Yin G-L. 2015 Epigenetic silencing of olfactomedin-4 enhances gastric cancer cell invasion via activation of focal adhesion kinase signaling. *BMB Rep.* **48**, 630–635. (doi:10.5483/BMBRep.2015.48.11.130)
139. Lu C-H, Wu W-Y, Lai Y-J, Yang C-M, Yu L-C. 2014 Suppression of B3GNT7 gene expression in colon adenocarcinoma and its potential effect in the metastasis of colon cancer cells. *Glycobiology* **24**, 359–367. (doi:10.1093/glycob/cwu002)
140. Ding W, Fan X-L, Xu X, Huang J-Z, Xu S-H, Geng Q, Li R, Chen D, Yan G-R. 2015 Epigenetic silencing of ITGA2 by MiR-373 promotes cell migration in breast cancer. *PLoS ONE* **10**, e0135128. (doi:10.1371/journal.pone.0135128)
141. Seol HS, Akiyama Y, Shimada S, Lee HJ, Kim TI, Chun SM, Singh SR, Jang SJ. 2014 Epigenetic silencing of microRNA-373 to epithelial-mesenchymal transition in non-small cell lung cancer through IRAK2 and LAMP1 axes. *Cancer Lett.* **353**, 232–241. (doi:10.1016/j.canlet.2014.07.019)
142. Korpala M, Lee ES, Hu G, Kang Y. 2008 The miR-200 family inhibits epithelial-mesenchymal transition and cancer cell migration by direct targeting of E-cadherin transcriptional repressors ZEB1 and ZEB2. *J. Biol. Chem.* **283**, 14 910–14 914. (doi:10.1074/jbc.C800074200)
143. Planagumà, J., Liljeström M, Alameda F, Bützow R, Virtanen I, Reventós J, Hukkanen M. 2011 Matrix metalloproteinase-2 and matrix metalloproteinase-9 codistribute with transcription factors RUNX1/AML1 and ETV5/ERM at the invasive front of endometrial and ovarian carcinoma. *Hum. Pathol.* **42**, 57–67. (doi:10.1016/j.humpath.2010.01.025)
144. Dong W, Chen X, Xie J, Sun P, Wu Y. 2010 Epigenetic inactivation and tumor suppressor activity of HAI-2/SPINT2 in gastric cancer. *Int. J. Cancer* **127**, 1526–1534. (doi:10.1002/ijc.25161)
145. Yue D *et al.* 2014 Epigenetic inactivation of SPINT2 is associated with tumor suppressive function in esophageal squamous cell carcinoma. *Exp. Cell Res.* **322**, 149–158. (doi:10.1016/j.yexcr.2013.11.009)
146. Hwang S, Kim H-E, Min M, Raghunathan R, Panova IP, Munshi R, Ryu B. 2015 Epigenetic silencing of SPINT2 promotes cancer cell motility via HGF-MET pathway activation in melanoma. *J. Invest. Dermatol.* **135**, 2283–2291. (doi:10.1038/jid.2015.160)
147. Wang L, Ge J, Ma T, Zheng Y, Lv S, Li Y, Liu S. 2015 Promoter hypermethylation of the cysteine protease RECK may cause metastasis of osteosarcoma. *Tumour Biol.* **36**, 9511–9516. (doi:10.1007/s13277-015-3688-4)
148. Zhang Y, Huang Z, Zhu Z, Zheng X, Liu J, Han Z, Ma X, Zhang Y. 2014 Upregulated UHRF1 promotes bladder cancer cell invasion by epigenetic silencing of KiSS1. *PLoS ONE* **9**, e104252. (doi:10.1371/journal.pone.0104252)
149. Wu D, Li M, Wang L, Zhou Y, Zhou J, Pan H, Qu P. 2014 microRNA-145 inhibits cell proliferation, migration and invasion by targeting matrix metalloproteinase-11 in renal cell carcinoma. *Mol. Med. Rep.* **10**, 393–398. (doi:10.3892/mmr.2014.2149)
150. Zhou H, Huang S. 2011 Role of mTOR signaling in tumor cell motility, invasion and metastasis. *Curr. Protein Pept. Sci.* **12**, 30–42. (doi:10.2174/138920311795659407)
151. Wei F-Z *et al.* 2015 Epigenetic regulation of autophagy by the methyltransferase EZH2 through an mTOR-dependent pathway. *Autophagy* **11**, 2309–2322. (doi:10.1080/15548627.2015.1117734)
152. Chen L, Li X, Chen X. 2015 Prognostic significance of tissue miR-345 downregulation in non-small cell lung cancer. *Int. J. Clin. Exp. Med.* **8**, 20 971–20 976.
153. Yu M *et al.* 2017 miR-345 inhibits tumor metastasis and EMT by targeting IRF1-mediated mTOR/STAT3/AKT

- pathway in hepatocellular carcinoma. *Int. J. Oncol.* **50**, 975–983. (doi:10.3892/ijo.2017.3852)
154. Xu J *et al.* 2012 Epigenetic regulation of HIF-1 α in renal cancer cells involves HIF-1 α /2 α binding to a reverse hypoxia-response element. *Oncogene* **31**, 1065–1072. (doi:10.1038/onc.2011.305)
 155. Vander Heiden MG *et al.* 2011 Metabolic pathway alterations that support cell proliferation. *Cold Spring Harb. Symp. Quant. Biol.* **76**, 325–334. (doi:10.1101/sqb.2012.76.010900)
 156. Liu VM, Vander Heiden MG. 2015 The role of pyruvate kinase M2 in cancer metabolism. *Brain Pathol.* **25**, 781–783. (doi:10.1111/bpa.12311)
 157. Wamelink MMC, Struys EA, Jakobs C. 2008 The biochemistry, metabolism and inherited defects of the pentose phosphate pathway: a review. *J. Inherit. Metab. Dis.* **31**, 703–717. (doi:10.1007/s10545-008-1015-6)
 158. Lopez-Serra P *et al.* 2014 A DERL3-associated defect in the degradation of SLC2A1 mediates the Warburg effect. *Nat. Commun.* **5**, 3608. (doi:10.1038/ncomms4608)
 159. Mondesir J, Willekens C, Touat M, de Botton S. 2016 IDH1 and IDH2 mutations as novel therapeutic targets: current perspectives. *J. Blood Med.* **7**, 171–180. (doi:10.2147/JBM.S70716)
 160. Parker SJ, Metallo CM. 2015 Metabolic consequences of oncogenic IDH mutations. *Pharmacol. Ther.* **152**, 54–62. (doi:10.1016/j.pharmthera.2015.05.003)
 161. Viswanath P, Chaumeil MM, Ronen SM. 2016 Molecular imaging of metabolic reprogramming in mutant IDH cells. *Front. Oncol.* **6**, 60. (doi:10.3389/fonc.2016.00060)
 162. Ohba S *et al.* 2016 Mutant IDH1 expression drives TERT promoter reactivation as part of the cellular transformation process. *Cancer Res.* **76**, 6680–6689. (doi:10.1158/0008-5472.CAN-16-0696)
 163. Baggott JE, Tamura T. 2015 Folate-dependent purine nucleotide biosynthesis in humans. *Adv. Nutr.* **6**, 564–571. (doi:10.3945/an.115.008300)
 164. Gao X, Reid MA, Kong M, Locasale JW. 2017 Metabolic interactions with cancer epigenetics. *Mol. Aspects Med.* **55**, 20–25. (doi:10.1016/j.mam.2016.09.001)
 165. Zhao H *et al.* 2012 Frequent epigenetic silencing of the folate-metabolising gene cystathionine-beta-synthase in gastrointestinal cancer. *PLoS ONE* **7**, e49683. (doi:10.1371/journal.pone.0049683)
 166. Jeschke J *et al.* 2013 Frequent inactivation of cysteine dioxygenase type 1 contributes to survival of breast cancer cells and resistance to anthracyclines. *Clin. Cancer Res.* **19**, 3201–3211. (doi:10.1158/1078-0432.CCR-12-3751)
 167. Archer SL. 2016 Acquired mitochondrial abnormalities, including epigenetic inhibition of superoxide dismutase 2, in pulmonary hypertension and cancer: therapeutic implications. *Adv. Exp. Med. Biol.* **903**, 29–53. (doi:10.1007/978-1-4899-7678-9_3)
 168. Liu Q, Jin J, Ying J, Sun M, Cui Y, Zhang L, Xu B, Fan Y, Zhang Q. 2015 Frequent epigenetic suppression of tumor suppressor gene glutathione peroxidase 3 by promoter hypermethylation and its clinical implication in clear cell renal cell carcinoma. *Int. J. Mol. Sci.* **16**, 10 636–10 649. (doi:10.3390/ijms160510636)
 169. Damhofer H, Medema JP, Veenstra VL, Badea L, Popescu I, Roelink H, Bijlsma MF. 2013 Assessment of the stromal contribution to Sonic Hedgehog-dependent pancreatic adenocarcinoma. *Mol. Oncol.* **7**, 1031–1042. (doi:10.1016/j.molonc.2013.08.004)
 170. Tang S-N, Fu J, Nall D, Rodova M, Shankar S, Srivastava RK. 2012 Inhibition of sonic hedgehog pathway and pluripotency maintaining factors regulate human pancreatic cancer stem cell characteristics. *Int. J. Cancer* **131**, 30–40. (doi:10.1002/ijc.26323)
 171. Heiden KB, Williamson AJ, Doscas ME, Ye J, Wang Y, Liu D, Xing M, Prinz RA, Xu X. 2014 The sonic hedgehog signaling pathway maintains the cancer stem cell self-renewal of anaplastic thyroid cancer by inducing snail expression. *J. Clin. Endocrinol. Metab.* **99**, E2178–E2187. (doi:10.1210/jc.2014-1844)
 172. Schmidt-Wolf IGH, Plass C, Byrd JC, Frevel K, Pietsch T, Waha A. 2016 Assessment of promoter methylation identifies PTCH as a putative tumor-suppressor gene in human CLL. *Anticancer Res.* **36**, 4515–4519. (doi:10.21873/anticancer.10998)
 173. Chen J-S *et al.* 2014 Down-regulation of Gli-1 inhibits hepatocellular carcinoma cell migration and invasion. *Mol. Cell. Biochem.* **393**, 283–291. (doi:10.1007/s11010-014-2071-x)
 174. Singh I *et al.* 2014 Hmga2 is required for canonical WNT signaling during lung development. *BMC Biol.* **12**, 21. (doi:10.1186/1741-7007-12-21)
 175. Morishita A, Zaidi MR, Mitoro A, Sankarasharma D, Szabolcs M, Okada Y, D'Armiento J, Chada K. 2013 HMGA2 is a driver of tumor metastasis. *Cancer Res.* **73**, 4289–4299. (doi:10.1158/0008-5472.CAN-12-3848)
 176. Mayr C, Hemann MT, Bartel DP. 2007 Disrupting the pairing between let-7 and Hmga2 enhances oncogenic transformation. *Science* **315**, 1576–1579. (doi:10.1126/science.1137999)
 177. Aithal MG, S, Rajeswari N. 2013 Role of Notch signalling pathway in cancer and its association with DNA methylation. *J. Genet.* **92**, 667–675. (doi:10.1007/s12041-013-0284-5)
 178. Karon M, Sieger L, Leimbrock S, Finklestein JZ, Nesbit ME, Swaney JJ. 1973 5-Azacytidine: a new active agent for the treatment of acute leukemia. *Blood* **42**, 359–365.
 179. Ritchie EK, Feldman EJ, Christos PJ, Rohan SD, Lagassa CB, Ippoliti C, Scandura JM, Carlson K, Roboz GJ. 2013 Decitabine in patients with newly diagnosed and relapsed acute myeloid leukemia. *Leuk. Lymphoma* **54**, 2003–2007. (doi:10.3109/10428194.2012.762093)
 180. Damaskos C *et al.* 2016 Histone deacetylase inhibitors: a novel therapeutic weapon against medullary thyroid cancer? *Anticancer Res.* **36**, 5019–5024. (doi:10.21873/anticancer.11070)
 181. Brown JAL, Bourke E, Eriksson LA, Kerin MJ. 2016 Targeting cancer using KAT inhibitors to mimic lethal knockouts. *Biochem. Soc. Trans.* **44**, 979–986. (doi:10.1042/BST20160081)
 182. Song Y, Wu F, Wu J. 2016 Targeting histone methylation for cancer therapy: enzymes, inhibitors, biological activity and perspectives. *J. Hematol. Oncol.* **9**, 49. (doi:10.1186/s13045-016-0279-9)
 183. Pérez-Salvia M, Esteller M. 2016 Bromodomain inhibitors and cancer therapy: from structures to applications. *Epigenetics* **12**, 323–339. (doi:10.1080/15592294.2016.1265710)
 184. Enríquez P. 2016 CRISPR-mediated epigenome editing. *Yale J. Biol. Med.* **89**, 471–486.
 185. Delpu Y, Cordelier P, Cho WC, Torrisani J. 2013 DNA methylation and cancer diagnosis. *Int. J. Mol. Sci.* **14**, 15 029–15 058. (doi:10.3390/ijms140715029)
 186. Wang J, Han X, Sun Y. 2017 DNA methylation signatures in circulating cell-free DNA as biomarkers for the early detection of cancer. *Sci. China Life Sci.* **60**, 356–362. (doi:10.1007/s11427-016-0253-7)
 187. Morán S *et al.* 2016 Epigenetic profiling to classify cancer of unknown primary: a multicentre, retrospective analysis. *Lancet Oncol.* **17**, 1386–1395. (doi:10.1016/S1470-2045(16)30297-2)
 188. Vizoso M *et al.* 2015 Epigenetic activation of a cryptic TBC1D16 transcript enhances melanoma progression by targeting EGFR. *Nat. Med.* **21**, 741–750. (doi:10.1038/nm.3863)
 189. Esteller M, Garcia-Foncillas J, Andion E, Goodman SN, Hidalgo OF, Vanaclocha V, Baylin SB, Herman JG. 2000 Inactivation of the DNA-repair gene MGMT and the clinical response of gliomas to alkylating agents. *N. Engl. J. Med.* **343**, 1350–1354. (doi:10.1056/NEJM200011093431901)
 190. Sun-tzu. 2008 *The art of war*. London, UK: Penguin Books Ltd.