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Data Article

Data on cytotoxic and antibacterial activity of synthesized Fe₃O₄ nanoparticles using *Malva sylvestris*

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ABSTRACT

The biosynthesis of materials using medicinal plants can be a low-cost and eco-friendly approach due to their extraordinary properties. Herein, we reported a facile synthesis of Fe₃O₄ nanoparticles using *Malva sylvestris*. The surface morphology, functional groups, and elemental analysis were done to characterize the synthesized nanoparticles. The cytotoxicity performance of the synthesized nanoparticles was analyzed by exposing nanoparticles to MCF-7 and Hep-G2 cancer cell lines through MTT colorimetric assay and the IC50 value was defined as 100 µg/mL and 200 µg/mL, respectively. The antibacterial performance of synthesized nanoparticles against four different bacterial strains including *Staphylococcus*

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aureus, *Corynebacterium*, *Pseudomonas aeruginosa*, and *Klebsiella pneumoniae* were assessed through microdilution broth method. The synthesized Fe₃O₄ nanoparticles using *Malva sylvestris* demonstrated higher antibacterial effects against Gram-positive strains with MIC values of 62.5 µg/mL and 125 µg/mL which increase the inhibitory percentage to more than 90%.
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Specifications Table

| | |
|--------------------------------|---|
| Subject | Biology |
| Specific subject area | Developmental biology |
| Type of data | Figure and table |
| How data were acquired | <ul style="list-style-type: none">- Cytotoxic and antibacterial tests.- X-Ray Diffraction (XRD)- Scanning Electron Microscopy (SEM)- Energy Dispersive X-Ray Spectroscopy (EDX)- Fourier-transform infrared spectroscopy (FT-IR). |
| Data format | Raw |
| Parameters for data collection | MIC and MBC methods for antibacterial tests and MTT assay for cytotoxic tests. |
| Description of data collection | The effects of cytotoxic and antibacterial activity of synthesized Fe ₃ O ₄ nanoparticles using <i>Malva sylvestris</i> were evaluated on different microorganisms by MIC and MBC methods. The cytotoxic effects were determined using two different cancer cell lines. |
| Data source location | Department of Medical Nanotechnology, School of Advanced Medical Sciences and Technologies, Shiraz University of Medical Sciences, Shiraz, Iran |
| Data accessibility | Data are provided in this article. The raw data source for this article is available in the Supplementary section . |

Value of the Data

- This research analyzes the cytotoxic and antibacterial effects of synthesized Fe₃O₄ nanoparticles using *Malva sylvestris* against four different microorganisms and two different cancer cell lines. Biologists, toxicologists, and pharmacists can pay close attention to the results of this study.
- Data from this work describe the anticancer activity of synthesized Fe₃O₄ Nanoparticles using *Malva sylvestris* in both human liver and breast cancer cell lines.
- The data are useful to compare the physicochemical characters, cytotoxic and antibacterial performances of the synthesized nanoparticles and can investigate the reliability of these synthesized nanoparticles in clinical applications.
- These data are relevant in developmental biology, especially for the understanding of the antibacterial and anticancer role of synthesized Fe₃O₄ nanoparticles using *Malva sylvestris* and their usage in therapeutic procedures for human cancers. So, it may be helpful for biology researchers to find a cancer therapy method.
- The synthesized nanoparticles can be applied as an antibacterial, antiseptic, and chemo-preventive agent in cancer treatment.

1. Data

The experimental data on cytotoxic and antibacterial activity of synthesized Fe₃O₄ nanoparticles using *Malva sylvestris* are reported in this dataset. To validate the successful synthesis of Fe₃O₄ nanoparticles using *Malva sylvestris*, Fourier transform infrared spectroscopy (FTIR) was applied to determine the functional groups in the nanoparticles (Fig. 1a). The X-ray powder diffraction (XRD) analysis is depicted in Fig. 1b. SEM images and EDX analysis of Fe₃O₄ nanoparticles and Fe₃O₄/*Malva sylvestris* nanoparticles are presented in Fig. 2.
The cytotoxic effects of synthesized Fe₃O₄ nanoparticles using *Malva sylvestris* against Hep-G2 and MCF-7 cell lines have demonstrated in Fig. 3. The inhibitory effect of synthesized Fe₃O₄ nanoparticles

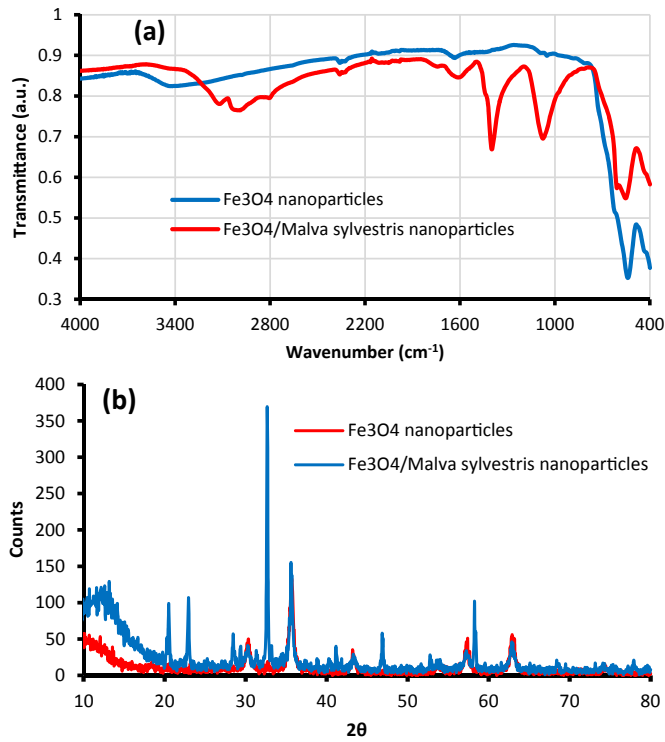


Fig. 1. (a) FTIR and (b) XRD results for the Fe₃O₄ and Fe₃O₄/Malva sylvestris nanoparticles.

using *Malva sylvestris* against four bacterial strains was analyzed through microdilution broth assay (see Fig. 4). The performance of nanoparticles against selected microorganisms is presented in Table 1. The raw data source for this dataset is available in the [Supplementary section](#).

2. Experimental design, materials, and methods

2.1. Preparation of Fe₃O₄/Malva sylvestris nanoparticles

All of the materials utilized in this investigation like Ferrous(II) Sulfate Heptahydrate (FeSO₄·7H₂O), Iron(III) Chloride Six hydrate (FeCl₃·6H₂O) and Ammonia solution were purchased from Merck Co. (Germany). First, 4.75 g FeCl₃·6H₂O, 3.89 g FeSO₄·7H₂O, and 320 mL deionized water were poured into a round-bottom flask and stirred for 1 h while the temperature was set to 80 °C [1,2]. Then, 0.5 g *Malva sylvestris* was ultrasonically mixed in 120 mL of deionized water for 30 min and then poured into the previous suspension. The obtained suspension was stirred under 80 °C for about 2 h and after that 40 mL NH₃ was gradually added to the suspension [3]. After filtration, the suspension was washed and the pH scale was set on 7 and finally dried in an oven at 100 °C for 2 h.

2.2. Characterization of nanoparticles

The synthesized nanoparticles were characterized using Fourier-transform infrared spectroscopy (FTIR, Tensor II FT-IR spectroscopy Bruker, Germany) in the region of 400–4000 cm⁻¹, X-ray diffraction (XRD, Panalytical model X'Pert Pro), scanning electron microscope (SEM, Tescan model Mira III), and EDX (Tescan model S Max detector Mira III).

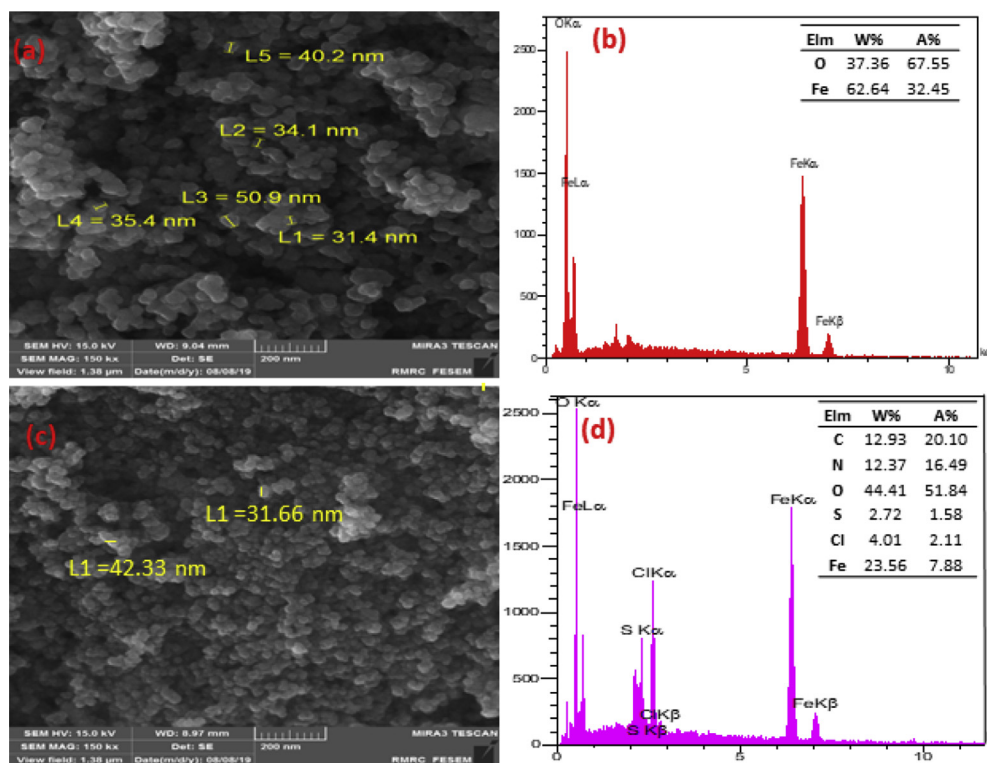


Fig. 2. FESEM images and EDAX analysis of (a,b) Fe_3O_4 nanoparticles and (c,d) $\text{Fe}_3\text{O}_4/\text{Malva sylvestris}$ nanoparticles.

2.3. MTT assay

The cytotoxicity of the synthesized nanoparticles was evaluated on Hep-G2 and MCF-7 cell lines using MTT colorimetric assay. In this method, hydrogen peroxide was considered as the positive control and culture medium was the negative control. Briefly, a certain number of Hep-G2 and MCF-7 cells (10^4) were placed in each well of a sterile 96-well microplate and incubated in a humidified atmosphere of 5% CO_2 , 95% air at 37 °C to reach about 75–90% confluence. Then, 100 μL of the synthesized nanoparticles in a wide range of concentrations was replaced with previous media. Afterward, 25 μL of the MTT 3-(4,5 Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium stock solution (with 4 mg/mL concentration) was transferred into each well and incubated for 4 h in standard condition. The mitochondrial performance of viable cells led to formation of purple formazan crystals, where for dissolving these crystals we applied 100 μL dimethyl sulfoxide (DMSO). In the final step, the absorption of solution was recorded at 570 nm wavelength using a microplate reader (Model 50, Bio-Rad Corp, Hercules, California, USA).

2.4. Minimum inhibitory concentrations (MICs) assay

In MICs test, all of the procedures were carried out according to the standards of the Clinical and Laboratory Standards Institute (CLSI) for assessing the antibacterial susceptibility of the synthesized nanoparticles [4]. Briefly, 2-fold serial dilutions of the testing compounds (at a descending concentration from 1000 $\mu\text{g}/\text{mL}$ to 7.8 $\mu\text{g}/\text{mL}$) and control groups were provided with Brain heart infusion (BHI) in 96-well microplates. Afterward, the microbial concentration was adapted to match the turbidity

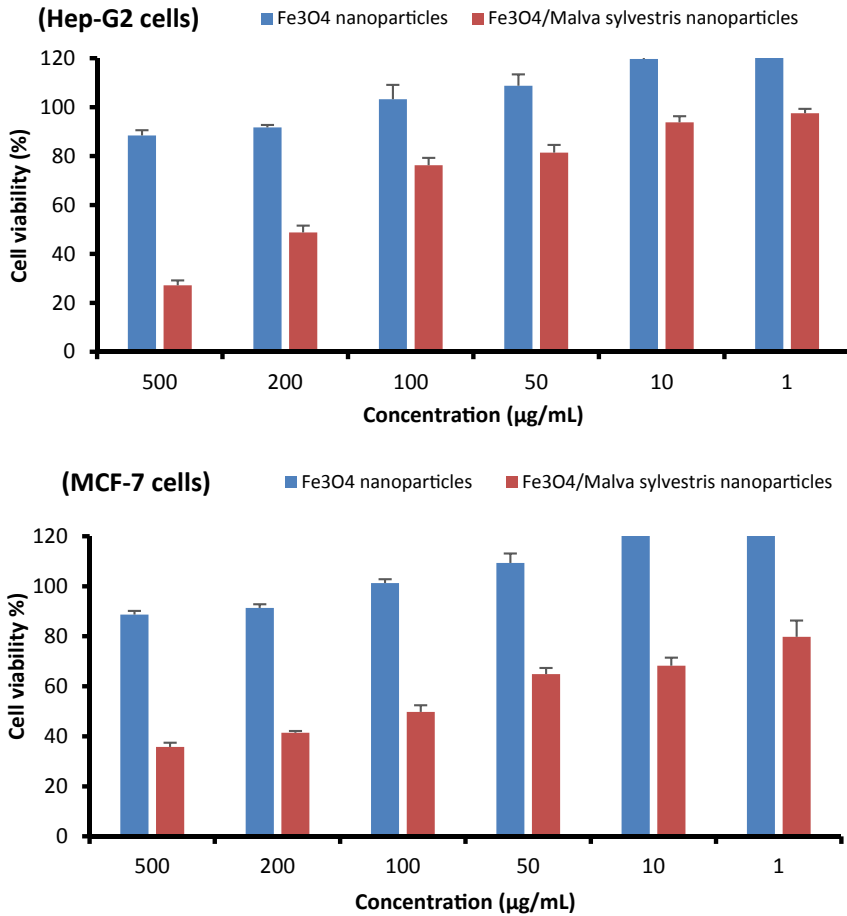


Fig. 3. Effect of nanoparticles on cell viability of MTT assay for all tested concentrations on Hep-G2 and MCF-7 cells after 24 h in comparison with control (untreated cell). Each bar represents the mean \pm SD (standard deviation) of three independent tests.

standard of 0.5 McFarland (OD600: 0.1–0.2) in a way that the concentration of the compounds was 1000 µg/mL in the first wells. The plates were incubated for 24 h at 37 °C. Later the optical density was measured at 600 nm by a microplate reader (BioTek, Power Wave XS2). This procedure was done in triplicate.

2.5. Minimum Bactericidal Concentrations (MBCs) assay

All the selected microorganisms were cultured overnight in BHI, and then stocks with the concentration of 10^5 – 10^6 CFU/mL were prepared for each one. The total of 90 µL of serially diluted concentrations of compounds (from 1000 µg/mL to 7.8 µg/mL) was added to a 96-well micro-plate consisting of 90 µg/mL BHI, then 10 µg/mL of bacteria were added to each cell. Micro-plates were incubated for 24 h at 37 °C. Then, 10 µL of each bacterial suspension was added to a newly prepared BHI and incubated for another 24 hours at 37 °C to exam bactericidal performance of each compound. The lowest concentration of compounds that leads no growth of bacteria was regarded as minimum bactericidal concentration (MBC). This procedure was also repeated three times.

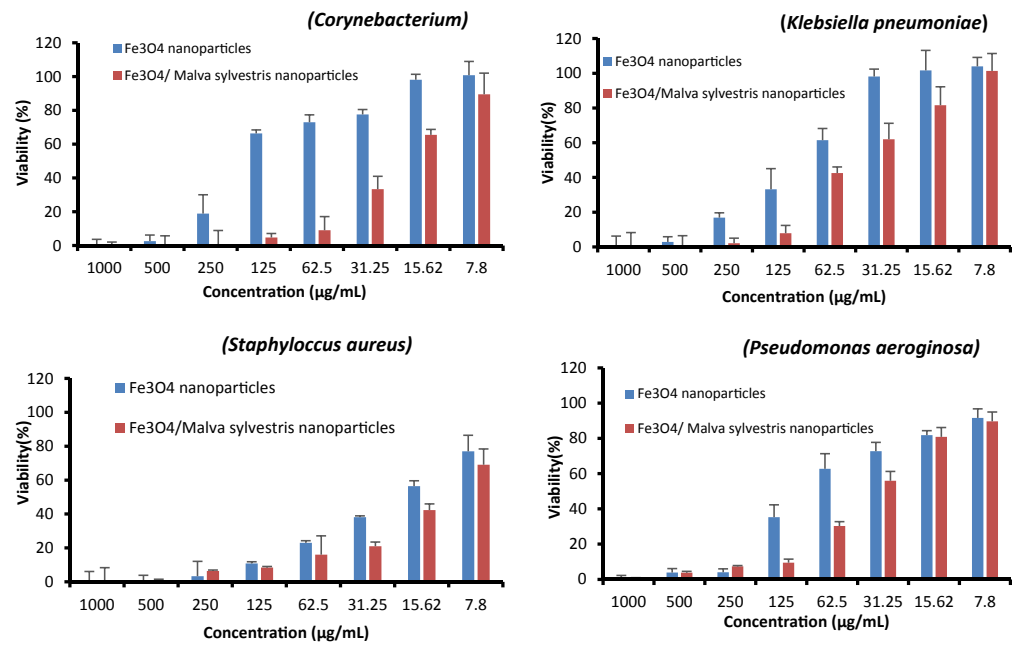


Fig. 4. Effect of nanoparticles on the viability percentages of *Corynebacterium*, *Klebsiella pneumoniae*, *Staphylococcus aureus*, *Pseudomonas aeruginosa* in tested concentrations (each bar represents the mean \pm SD (standard deviation) of three independent tests).

Table 1
Performance of nanoparticles against selected microorganisms.

| Microorganisms | Fe ₃ O ₄ nanoparticles (µg/mL) | | Fe ₃ O ₄ /Malva sylvestris nanoparticles (µg/mL) | |
|-------------------------------|--|------|--|------|
| | MIC | MBC | MIC | MBC |
| <i>Staphylococcus aureus</i> | 250 | 250 | 125 | >125 |
| <i>Corynebacterium</i> | 500 | 1000 | 62.5 | 125 |
| <i>Pseudomonas aeruginosa</i> | 250 | 500 | 125 | >125 |
| <i>Klebsiella pneumoniae</i> | 500 | 500 | 125 | >125 |

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Conflict of Interest

The authors declare that they have no known competing financial interests or personal relationships that could have appeared to influence the work reported in this paper.

Appendix A. Supplementary data

Supplementary data to this article can be found online at <https://doi.org/10.1016/j.dib.2019.104929>.

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